



<http://dx.doi.org/10.11646/zootaxa.3821.3.3>

<http://zoobank.org/urn:lsid:zoobank.org:pub:BFE89BF2-E024-4C1B-ABD5-1A2A286F0BE6>

## *Steinernema tophus* sp. n. (Nematoda: Steinernematidae), a new entomopathogenic nematode from South Africa

HARUN ÇIMEN<sup>1</sup>, MING-MIN LEE<sup>2</sup>, JUSTIN HATTING<sup>3</sup>, SELCUK HAZIR<sup>1</sup> & S. PATRICIA STOCK<sup>2,4</sup>

<sup>1</sup>Adnan Menderes University, Faculty of Arts and Science, Department of Biology, 09010 Aydin, Turkey

<sup>2</sup>Department of Entomology, University of Arizona. 1140 E. South Campus Dr. Tucson, AZ 85721-0036, Arizona, USA

<sup>3</sup>South African Agricultural Research Council, Small Grain Institute, Private Bag X29, Bethlehem, 9701, South Africa

<sup>4</sup>Corresponding author. E-mail: [spstock@email.arizona.edu](mailto:spstock@email.arizona.edu). Telephone: +1-520-626-3854, Fax: +1-520-621-1150

### Abstract

A new entomopathogenic nematode, *Steinernema tophus* n. sp. is described from South Africa. Morphological, molecular (ribosomal gene sequence data) together with cross-hybridization studies were used for diagnostics and identification purposes. Both molecular and morphological data indicate the new species belongs to the ‘*glaseri*-group’ of *Steinernema* spp. Key morphological diagnostic traits for *S. tophus* n. sp. include the morphology of the spicules and gubernaculum. Morphometric traits of third-stage infective juveniles, including total body length (average 1,046µm), tail length (average 70µm), location of the excretory pore (average 92 µm), D% (average 63), E% (average 132) and H% (average 32) values are definitive. In addition to these morphological characters, analysis of rDNA (28S and ITS) gene sequences depict this *Steinernema* species as a distinct and unique entity.

**Key words:** *Steinernema*, South Africa, rDNA genes, mitochondrial genes, morphology, cross hybridization

### Introduction

*Steinernema* nematodes Travassos, 1927 are obligate and lethal endoparasites that have a symbiotic relationship with Gram-negative  $\gamma$ -Proteobacteria in the genus *Xenorhabdus* Thomas & Poinar, 1979. This nematode-bacteria complex represents a mutualistic association, where the nematodes (third stage IJs<sup>1</sup>) vector the symbiotic bacteria between insects in a specialized intestinal receptacle (Stock & Goodrich-Blair, 2008). Once the bacteria are released in the insect’s hemocoel, the bacteria kill the insect host and create a favorable environment within the host cadaver for nematode growth and development.

This pairing is pathogenic for a wide range of insects and has successfully been implemented in biological control and integrated pest management programs worldwide (Gaugler & Kaya, 1993; Gaugler, 2002). Surveys for EPNs<sup>2</sup> have been conducted in temperate, subtropical and tropical regions (summarized by Hominick, 2002; Adams *et al.*, 2006). In South Africa, several surveys have been conducted to document the diversity of this group of nematodes, with the goal of finding locally-adapted species and/or isolates that can be assayed for control of native insect pests (Malan *et al.*, 2006, 2011, 2012; Hatting *et al.*, 2009; Pillay *et al.*, 2009).

Until now, five *Steinernema* and four *Heterorhabditis* species have been isolated and/or described in South Africa, and several of these species are being implemented in successful agricultural pest management programs (De Waal *et al.*, 2010; Malan *et al.*, 2009; Van Niekerk & Malan, 2012; Çimen *et al.* 2014).

This study describes a new *Steinernema* sp. originally recovered from a vineyard in Clanwilliam (Western Cape), South Africa. Differential interference contrast optics (DIC), DNA sequence analysis and cross-hybridisation assays were conducted to describe and illustrate this new *Steinernema* species.

1. Infective juvenile
2. Entomopathogenic nematode

related species by the morphology of the spicules and gubernaculum, the arrangement of the genital papillae and the values of ratios SW (average: 1.2, range: 1.0–1.6) and GS (average: 0.7, range: 0.6–0.9).

Phylogenetic analyses placed *S. tophus* n. sp. in clade V (as depicted by Spiridinov *et al.* 2004). Within this clade, the new species was found most closely related to a newly described South African species, *S. innovationi* followed by *S. khoisanae* (Nguyen *et al.* 2006) in the 28s topology. However, the new species can be differentiated from *S. khoisanae* and *S. innovationi* by morphological and morphometric differences of the IJ and first generation male (See Table 1). Furthermore, phylogenetic analysis of ITS and 28s datasets revealed distinct base pair differences between *S. tophus* n. sp. and members of the clade V *glaseri*-group (Tables 3 and 4). In particular, for the more variable gene, ITS dataset, there were 59 base pairs difference between *S. tophus* n. sp. and its sister species, *S. innovationi*.

*Steinernema tophus* n. sp. can be differentiated from *S. khoisanae* by the morphology of both infective juveniles and first generation adult males. For example *S. tophus* n. sp. infective juveniles are wider (average 38  $\mu\text{m}$  vs 33  $\mu\text{m}$ ) and slightly shorter than those *S. khoisanae* (average 1,046  $\mu\text{m}$  vs 1,075 $\mu\text{m}$ ). The excretory pore in the new species is generally located more posteriorly, and the tail is usually shorter than that of *S. khoisanae* (average 70  $\mu\text{m}$  vs 85  $\mu\text{m}$ ), though given the overlap in ranges for these features, they should not be considered key diagnostic traits, but only as general trends. Males of *S. tophus* n. sp. can be distinguished from *S. khoisanae* by the D% (average: 92 vs 88) and SW ratio (average: 1.2 vs 1.99). First-generation females of *S. tophus* n. sp. are characterized by having a conoid tail with a mucro, whereas in *S. khoisanae* the tail is digitated.

Third-stage infective juveniles of *S. tophus* n. sp. differ from those of *S. innovationi* by the location of the excretory pore (average 90  $\mu\text{m}$  vs 88  $\mu\text{m}$ ), the tail length (average 70  $\mu\text{m}$  vs 76  $\mu\text{m}$ ) and the values of D% (63 vs 58) and E% (132 vs. 115). First generation females of the new species and *S. innovationi* have a mucronated tail, however the new species does not have a digitate tail which is present in *S. innovationi*. Males of *S. tophus* n. sp. can be separated from those of *S. innovationi* by the morphology the spicules. Specifically, the spicule calomus is rhomboidal in *S. innovationi* but it is elongated in the new species. The curvature of the lamina is less pronounced in *S. tophus* n. sp. when compared to *S. innovationi*. Furthermore, the number and arrangement of postcloacal papillae is different when compared to *S. innovationi*. The new species has four pairs of postcloacal papillae of which one subventral, one subdorsal and two terminal) whereas *S. innovationi* has five postcloacal pairs (one subventral and two subdorsal and two terminal). First generation males of the new species differ from *S. tophus* n. sp. males also differ in the value of SW when compared to that of *S. innovationi* (average 1.2 vs. 1.4)

## Acknowledgements

This research was partially funded by the South African Agricultural Research Council (ARC) under Project GK05/14 to J. Hatting. Training of H. Çimen in P. Stock's Laboratory was supported in part by NemaSym Research Coordination Network grant (S. P. Stock, NSF IOS 0840932). The authors have no conflicts of interest to declare.

## References

- Adams, B.J., Fodor, A., Klein, M.G., Smith, H.L., Stackebrandt, E., Stock, S.P. & Klein, M.G. (2006) Biodiversity and systematics of nematode-bacterium entomopathogens. *Biological Control*, 37, 32–49.  
<http://dx.doi.org/10.1016/j.biocontrol.2005.11.008>
- Artyukhovsky, A.K., Kozodoi, E.M., Reid, A.P. & Spiridonov, S.E. (1997) Redescription of *Steinernema arenarium* (Artyukhovsky, 1967) topotype from Central Russia and a proposal for *S. anomalae* (Kozodoi, 1984) as a junior synonym. *Russian Journal of Nematology*, 5, 31–37.
- Bedding, R.A. & Akhurst, R.J. (1975) A simple technique for the detection of insect parasitic rhabditid nematodes in soil. *Nematologica*, 21, 109–110.  
<http://dx.doi.org/10.1163/187529275x00419>
- Çimen, H., Lee, M.-M., Hatting, J., Hazir, S. & Stock, S.P. (2014) *Steinernema innovationi* n. sp. (Panagrolaimomorpha: Steinernematidae) a new entomopathogenic nematode from South Africa. *Journal of Helminthology*, 1–4.  
<http://dx.doi.org/10.1017/s0022149x14000182>
- De Waal, J.Y., Malan, A.P., Levings, J. & Addison, M.F. (2010) Key elements in the successful control of diapausing codling moth, *Cydia pomonella* (Lepidoptera: Tortricidae) in wooden fruit bins with a South African isolate of *Heterorhabditis*

- zealandica* (Rhabditida: Heterorhabditidae). *Biocontrol Science and Technology*, 20, 489–502.  
<http://dx.doi.org/10.1080/09583151003599708>
- Edgar, R.C. (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research*, 32, 1792–1797.  
<http://dx.doi.org/10.1093/nar/gkh340>
- Franklin, M. & Goodey, J.B. (1949) A Cotton Blue-Lactophenol Method for Mounting Plant Parasitic Nematodes. *Journal of Helminthology*, 23, 175–178.  
<http://dx.doi.org/10.1017/s0022149x0003251x>
- Gaugler, R. (2002) *Entomopathogenic nematology*. CAB International Wallingford, UK, 400 pp.
- Gaugler, R. & Kaya, H.K. (1993) *Entomopathogenic nematodes in biological control*. CRC Press Boca Raton, Florida, 365 pp.
- Hatting, J., Stock, S.P. & Hazir, S. (2009) Diversity and distribution of entomopathogenic nematodes (Steinernematidae, Heterorhabditidae) in South Africa. *Journal of Invertebrate Pathology*, 102, 120–128.  
<http://dx.doi.org/10.1016/j.jip.2009.07.003>
- Hominick, W.M. (2002) Biogeography. In: Gaugler, R. (Ed.), *Entomopathogenic Nematology*. CABI Publishing Wallingford, UK, pp.115–143.
- Hominick, W.M., Briscoe, B.R., del Pino, F.G., Heng, J., Hunt, D.J., Kozodoy, E., Mráček, Z., Nguyen, K.B., Reid, A.P., Spiridonov, S., Stock, S.P., Sturhan, D., Waturu, C. & Yoshida, M. (1997) Biosystematics of entomopathogenic nematodes, current status, protocols and definitions. *Journal of Helminthology*, 71, 271–298.  
<http://dx.doi.org/10.1017/s0022149x00016096>
- Huelsenbeck, J.P. & Ronquist, F. (2001) MrBayes: Bayesian inference of phylogeny. *Bioinforma*, 17, 754–755.
- Kaya, H.K. & Stock, S.P. (1997) Techniques in insect nematology. In: Lacey, L. (Ed.) *Manual of techniques in insect pathology*. Academic Press Ltd. San Diego, California, pp. 281–324.
- Lee, M.M., Sicard, M., Skeie, M. & Stock, S.P. (2009) *Steinernema boemarei* n. sp. (Nematoda: Steinernematidae), a new entomopathogenic nematode from southern France. *Systematic Parasitology*, 72, 127–141.  
<http://dx.doi.org/10.1007/s11230-008-9166-2>
- Maddison, W.P. & Maddison, D.R. (2011) Mesquite: a modular system for evolutionary analysis. Version 2.75. Available from: <http://mesquiteproject.org> (accessed 10 June 2014)
- Malan, A.P., Nguyen, K.B. & Addison, M.F. (2006) Entomopathogenic nematodes (Steinernematidae and Heterorhabditidae) from the southwestern parts of South Africa. *African Plant Protection*, 12, 65–69.
- Malan, A.P. & Manrakhan, A. (2009) Susceptibility of the Mediterranean fruit fly (*Ceratitis capitata*) and the Natal fruit fly (*Ceratitis rosa*) to entomopathogenic nematodes. *Journal of Invertebrate Pathology*, 100, 47–49.  
<http://dx.doi.org/10.1016/j.jip.2008.09.007>
- Malan, A.P., Knoetze, R. & Moore, S.D. (2011) Isolation and identification of entomopathogenic nematodes from citrus orchards in South Africa and their biocontrol potential against false codling moth. *Journal of Invertebrate Pathology*, 108, 115–25.  
<http://dx.doi.org/10.1016/j.jip.2011.07.006>
- Malan, A.P., Knoetze, R. & Tiedt, L. (2012) *Heterorhabditis noenieputensis* n. sp. (Rhabditida: Heterorhabditidae), a new entomopathogenic nematode from South Africa. *Journal of Helminthology*, 12, 1–13.  
<http://dx.doi.org/10.1017/s0022149x12000806>
- McClure, M.J. & Stowell, L.J. (1978) A simple method of processing nematodes for electron microscopy. *Journal of Nematology*, 18, 376–377.
- Mráček, Z., Hernandez, E.A. & Boemare, N.E. (1994) *Steinernema cubana* sp. n. (Nematoda: Rhabditida: Steinernematidae) and the preliminary characterization of its associated bacterium. *Journal of Invertebrate Pathology*, 64, 123–129.  
<http://dx.doi.org/10.1006/jipa.1994.1080>
- Nadler, S.A., Bolotin, E. & Stock, S.P. (2006) Phylogenetic relationships of *Steinernema* (Cephalobina, Steinernematidae) based on nuclear, mitochondrial, and morphological data. *Systematic Parasitology*, 63, 159–179.  
<http://dx.doi.org/10.1007/s11230-005-9009-3>
- Nguyen, K.B., Maruniak, J. & Adams, B.J. (2001) The diagnostic and phylogenetic utility of the rDNA internal transcribed spacer sequences in *Steinernema*. *Journal of Nematology*, 33, 73–82.
- Nguyen, K.B. & Duncan, L.W. (2002) *Steinernema diaprepesi* n. sp. (Rhabditida: Steinernematidae), a parasite of the citrus root weevil *Diaprepes abbreviatus* (L) (Coleoptera: Curculionidae). *Journal of Nematology*, 34, 159–170.
- Nguyen, K.B., Malan, A. & Gozel, U. (2006) *Steinernema khoisanai* n. sp. (Rhabditida: Steinernematidae), a new entomopathogenic nematode from South Africa. *Nematology*, 8, 157–175.  
<http://dx.doi.org/10.1163/156854106777998728>
- Nylander, J.A.A., Ronquist, F., Huelsenbeck, J.P. & Nieves-Aldrey, J.L. (2004) Bayesian phylogenetic inference of combined data. *Systematic Biology*, 53, 47–67.  
<http://dx.doi.org/10.1080/10635150490264699>
- Pillay, U., Martin, L.A., Rutherford, R.S. & Berry, S.D. (2009) Entomopathogenic nematodes in sugar cane in South Africa. *Proceedings of the South African Sugar Technology Association*, 82, 538–541.
- Poinar, G.O. Jr. (1990) Entomopathogenic nematodes in biological control. In: Gaugler, R. & Kaya, K.H. (Eds.), *Taxonomy and biology of Steinernematidae and Heterorhabditidae*. CRC Press Boca Raton, Florida, pp. 23–74.

- Qiu, Y.F., Fang, Y., Zhou, Y., Pang, Y. & Nguyen, K.B. (2004) *Steinernema guangdongense* sp. n. (Nematoda: Steinernematidae), a new entomopathogenic nematode from southern China with a note on *S. serratum* (*nomen nudum*). *Zootaxa*, 704, 1–20.
- Rambaut, A. & Drummond, A. (2007) BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology*, 7, 214.  
<http://dx.doi.org/10.1186/1471-2148-7-214>
- Román, J. & Figueroa, W. (1994) *Steinernema puertoricensis* n. sp. (Rhabditida: Steinernematidae) a new entomopathogenic nematode from Puerto Rico. *Journal of Agriculture of the University of Puerto Rico*, 78, 167–175.
- Saitou, N. & Nei, M. (1987) The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4, 406–425.
- Seinhorst, J.W. (1959) A rapid method for the transfer of nematodes from fixative to anhydrous glycerin. *Nematologica*, 4, 67–69.
- Shen, C.P. & Wang, G.H. (1991) Description and study of an entomopathogenic nematode, *Steinernema longicaudum* sp. nov. and its application. In: Young and Middle Aged Science and Technology Works, Plant Protection (Ed.), *Proceedings of the first national academy association*. Chinese Science and Technology Press Beijing, China, pp. 220–231.
- Spiridinov, S.E., Reid, A.P., Podrucka, K., Subbotin, S.A. & Moens, M. (2004) Phylogenetic relationships within the genus *Steinernema* (Nematoda: Rhabditida) as inferred from analysis of sequences of the ITS-1-5.8S-ITS2 region of rDNA and morphological features. *Nematology*, 6, 547–566.
- Steiner, G. (1929) *Neoapectana glaseri* n. g. n. sp. (Oxyuridae), a new nemic parasite of the Japanese beetle (*Popillia japonica* Newm.). *Journal of the Washington Academy of Science*, 19, 436–440.
- Stock, S.P., Campbell, J.F. & Nadler, S.A. (2001a) Phylogeny of *Steinernema* Travassos, 1927 (Cephalobina, Steinernematidae) inferred from ribosomal DNA sequences and morphological characters. *Journal of Parasitology*, 87, 877–889.  
<http://dx.doi.org/10.2307/3285148>
- Stock, S.P., Heng, J., Hunt, D.J., Reid, A.P., Shen, X. & Choo, H.Y. (2001b) Redescription of *Steinernema longicaudum* Shen and Wang (Nematoda: Steinernematidae); geographic distribution and phenotypic variation between allopatric populations. *Journal of Helminthology*, 75, 81–92.  
<http://dx.doi.org/10.1079/joh200036>
- Stock, S.P. & Goodrich-Blair, H. (2008) Entomopathogenic nematodes and their bacterial symbionts: the inside out of a mutualistic symbiosis. *Symbiosis*, 46, 65–76.
- Stock, S.P. & Goodrich-Blair, H. (2012) Nematode parasites, pathogens and associates of insects and invertebrates of economic importance. In: Lacey, L.A. (Ed.), *Manual of Techniques in Invertebrate Pathology*. Elsevier Yakima, Washington, pp.373–426.
- Stock, S.P. & Koppenhöfer, A.M. (2003) *Steinernema scarabaei* n. sp. (Rhabditida: Steinernematidae), a natural pathogen of scarab beetle larvae (Coleoptera: Scarabaeidae) from New Jersey, USA. *Nematology*, 5, 191–204.  
<http://dx.doi.org/10.1163/156854103767139680>
- Stock, S.P., Griffin, C.T. & Chaenari, R. (2004) Morphological and molecular characterisation of *Steinernema hermaphroditum* n. sp. (Nematoda: Steinernematidae), an entomopathogenic nematode from Indonesia, and its phylogenetic relationships with other members of the genus. *Nematology*, 6, 401–412.  
<http://dx.doi.org/10.1163/1568541042360555>
- Stokwe, N., Malan A., Nguyen, K., Knoetze, R. & Tiedt, L. (2011) *Steinernema citrae* n. sp. (Rhabditida: Steinernematidae), a new entomopathogenic nematode from South Africa. *Nematology*, 13, 569–587.  
<http://dx.doi.org/10.1163/138855410x535714>
- Swofford, D.L. (2002) PAUP\*. *Phylogenetic analysis using parsimony (and other methods)*. Version 4. Sinauer Associates Sunderland, Mass., 257 pp.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M. & Kumar, S. (2011) MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Molecular Biology and Evolution*, 28, 2731–2739.  
<http://dx.doi.org/10.1093/molbev/msr121>
- Thompson, J., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D.G. (1997) The ClustalX windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Research*, 25, 4876–4882
- Uribe-Lorio, L., Mora, M. & Stock, S.P. (2007) *Steinernema costaricense* n. sp. and *Steinernema puntauvense* n.sp. (Rhabditida, Steinernematidae), two new entomopathogenic nematodes from Costa Rica. *Systematic Parasitology*, 68, 167–172.  
<http://dx.doi.org/10.1007/s11230-007-9098-2>
- van Niekerk, S. & Malan, A.P. (2012) Potential of South African entomopathogenic nematodes (Heterorhabditidae and Steinernematidae) for control of the citrus mealybug, *Planococcus citri* (Pseudococcidae). *Journal of Invertebrate Pathology*, 11, 166–174.  
<http://dx.doi.org/10.1016/j.jip.2012.07.023>
- Waturu, C.N., Hunt, D.J. & Reid, A.P. (1997) *Steinernema kari* sp n. (Nematoda: Steinernematidae), a new entomopathogenic nematode from Kenya. *International Journal of Nematology*, 7, 68–75.