

<http://dx.doi.org/10.11646/zootaxa.3949.2.4>
<http://zoobank.org/urn:lsid:zoobank.org:pub:F74491D7-DD55-4737-BA32-5E0B54A278FA>

Morphological and morphometric comparison of the first zoeal stage of the mangrove crabs of the genus *Aratus* H. Milne Edwards, 1853 (Decapoda: Sesarmidae)

ADRIANA P. REBOLLEDO^{1,4}, INGO S. WEHRTMANN^{1,2} & JOSE A. CUESTA³

¹Unidad de Investigación Pesquera y Acuicultura (UNIP) of the Centro de Investigación en Ciencias del Mar y Limnología (CIMAR), Universidad de Costa Rica, 11501-2060 San José, Costa Rica

²Museo de Zoología, Escuela de Biología, Universidad de Costa Rica, 11501-2060 San José, Costa Rica.

³Instituto de Ciencias Marinas de Andalucía (CSIC), Avenida República Saharaui, 2, E-11510 Puerto Real, Cádiz, Spain

⁴Corresponding author. E-mail: adripn@gmail.com

Abstract

The mangrove crab *Aratus pisonii* (H. Mile Edwards, 1837) was considered to have an amphi-American distribution but a recent genetic study revealed that the Eastern Tropical Pacific populations represent a new species, *A. pacificus* (Thiercelin & Schubart, 2014). These sister species separated by the Central American Isthmus have developed under different environmental conditions that may influence their larval development. A comparison of morphological and morphometric features (length and width of cephalothorax and length of rostral and dorsal spine, antenna, antennule, telson, and furcae) of recently-hatched larvae of *A. pacificus* (Pacific coast) and *A. pisonii* (Caribbean coast) from Costa Rica revealed that the setation pattern of the antennules differed between the species and the analyzed morphometric features were larger in *A. pisonii* larvae. Difference in size may be a response to different environmental conditions, as the lower primary production in coastal Caribbean waters, which may have forced females of *A. pisonii* to allocate more energy into the offspring, resulting in larger zoeal size. A greater endogenous reserve may allow the larvae to reduce the duration of the planktonic phase and increase the size at metamorphosis, thus enhancing their survival chances during the planktonic phase. These data regarding morphological and morphometric differences in recently-hatched larvae of the Pacific and Caribbean species support the conclusion that specimens of *Aratus* from both coasts of Costa Rica represent indeed different species.

Key words: Larvae, transisthmian sister species, interspecific variability, speciation

Introduction

Most marine invertebrates have a complex life cycle, usually characterized by larvae that drift in the plankton until they metamorphose into a juvenile stage that recruit the habitat of the adults (Giménez & Anger 2003; Anger 2006). The larval phase present a wide array of adaptive traits that differ from subsequent life history stages (Anger 2001, 2006). These traits are genetically heritable and influenced by past and present environmental conditions (Anger 2006; Giménez 2006). Larval descriptions are therefore often considered in the process to define phylogenetic relationships (Ng & Clark 2000; Felder & Martin 2003; Anger 2006), particularly in cases where adults are difficult to distinguish (Ng & Clark 2000; Cuesta & Anger 2001).

The emergence of the Central American Isthmus, roughly 3.5 million years ago, provides a unique opportunity to study how environmental conditions shaped the expression of early ontogenetic characters of species separated by this geographic barrier (Lessios 2008; McAlister 2008). The Isthmus divided a previously continuous oceanic region, causing the formation of two very different marine systems, the Eastern Tropical Pacific (ETP) and Caribbean Sea. Each of these systems presents particular conditions regarding temperature, salinity, seasonality, and primary productivity (Haugh & Tiedermann 1998; Steph *et al.* 2006; Jain & Collins 2007). The latter is especially important since food availability is considered one of the key factors determining the endurance of invertebrate larvae in the pelagic environment (Anger 2001; Giménez & Anger 2005).

endogenous nutrient reserves is advantageous to reduce their dependence on external food (Moran & Allen 2007) and improve their fitness in this more competitive environment (Allen *et al.* 2008). In areas with reduced food availability, larger offspring perform better than smaller conspecific; the greater energetic reserve provide them the advantage of reaching metamorphosis sooner, thereby shortening their planktonic phase, which in turn reduces the risk of predation and starvation (Hart 1995; Giménez & Anger 2003; 2005, Bas *et al.* 2007).

Temperature can also influences the embryo development and the larval phase of crustacean larvae (Shirley *et al.* 1987; Wehrtmann & López 2003; Barría *et al.* 2005). At higher temperatures the incubation period is reduced (Wear 1974; Hamasaki 2003), and the hatching larvae tend to attain smaller sizes than those where embryos were incubated at lower temperatures (Shirley *et al.* 1987; Wehrtmann & López 2003). This tendency can also be observed in the case of the two *Aratus* species studied herein: the zoea I obtained from the sampling site with considerably higher average temperature (Pacific; *Aratus pacificus*) were smaller than those (*A. pisonii*) collected from the Caribbean coast with lower average temperature.

Apart of these size differences, larvae from both species studied herein differed significantly in the ratios of dorsal/rostral spine length (Table 4), with *A. pisonii* zoeae having proportionally larger dorsal and rostral spines. Cuesta and Anger (2001) nevertheless compared the TFL/TBL ratio among different sesarmid species and reported for a Caribbean population of *A. pisonii* a lower value (<1.9) than those obtained in this study. Comparisons with additional populations are therefore needed to determine the degree of interpopulational variability of this parameter.

Larval studies between other decapod species occurring at both Caribbean and Pacific coasts have raised doubts about their conspecific status (i.e. Gore 1972; Thatje & Bacardit 2000), especially due to the fact that transisthmian sibling species showed clear form and size variations in early ontogenetic stages (Cuesta & Schubart 1999; Bartilotti *et al.* 2012). Thiercelin and Schubart (2014) more recently analyzed *Aratus* specimens collected across the Isthmus, and used genetic as well as male gonopod characteristics to distinguish two *Aratus* species and to describe *A. pacificus* for the Pacific coast of Central America.

The intrageneric variability is typically very low in the first zoea of decapods. As mentioned by Mantelatto *et al.* (2014), there are only very few characters where first zoeal stages show differences among closely related species. The setae and aesthetasc numbers of the zoea I antennules remained constant between specimens of the same coasts, but differed between the Pacific and Atlantic individuals (Table 2). The difference regarding this feature, together with the smaller size of the offspring of the Pacific corroborate the recent separation of the genus in a Pacific and Atlantic species.

Acknowledgments

We are grateful to the Instituto Costarricense de Electricidad (ICE), which provided logistic support for the collection of *Aratus pacificus* in the Térraba-Sierpe's mangrove as part of the Diquís Hydroelectric Project. Special thanks are due to the assistants that collaborated with the sampling of *A. pisonii* in the Gandoca's lagoon. We appreciate the comments of the anonymous reviewers and Zootaxa editor, Dr. Peter Castro, which served to improve the quality of the manuscript. The present study was completed by the first author to partially fulfill the requirements for a M.Sc. degree in Biology at the Escuela de Biología, Universidad de Costa Rica.

References

- Aikawa, H. (1929) On larval forms of some Brachyura. *Records of Oceanographic Works of Japan*, 2, 17–55.
- Allen, J.D., Zakas, C. & Podolsky, R.D. (2006) Effects of egg size reduction and larval feeding on juvenile quality for a species with facultative-feeding development. *Journal of Experimental Marine Biology and Ecology*, 331, 186–197.
<http://dx.doi.org/10.1016/j.jembe.2005.10.020>
- Allen, R.M., Buckley, Y.M., & Marshall, D.J. (2008) Offspring size plasticity in response to intraspecific competition: an adaptive maternal effect across life-history stages. *The American Naturalist*, 171, 225–237.
<http://dx.doi.org/10.1086/524952>
- Anger, K. (2001) The biology of decapod crustacean larvae. *Crustacean Issues* 14. Swets & Zeitlinger B.V., Lisse, The Netherlands.
- Anger, K. (2006) Contributions of larval biology to crustacean research: a review. *Invertebrate Reproduction &*

- Development*, 49, 175–205.
<http://dx.doi.org/10.1080/07924259.2006.9652207>
- Barría, E.M., Jara, C.G. & Paschke, K.A. (2005) La temperatura como factor de variabilidad en el desarrollo y morfología de larvas zoeas de *Acanthocyclus hassleri* Rathbun (Decapoda, Brachyura, Atelecyclidae) cultivadas en laboratorio. *Investigaciones Marinas*, 33, 25–41.
<http://dx.doi.org/10.4067/S0717-71782005000100002>
- Bartolotti, C., Calado, R., Rhyne, A. & Dos Santos, A. (2012) Shedding light on the larval genus *Eretmocaris*: morphological larval features of two closely related trans-isthmian *Lysmata* species (Decapoda: Caridea: Hippolytidae) described on the basis of laboratory cultured material. *Helgoland Marine Research*, 66, 97–115.
<http://dx.doi.org/10.1007/s10152-011-0251-6>
- Bas, C.C., Spivak, E.D. & Anger, K. (2007) Seasonal and interpopulational variability in fecundity, egg size, and elemental composition (CHN) of eggs and larvae in a grapsoid crab, *Chasmagnathus granulatus*. *Helgoland Marine Research*, 61, 225–237.
<http://dx.doi.org/10.1007/s10152-007-0070-y>
- Clark, P.F., Calazans, D. & Pohle, G.W. (1998) Accuracy and standardization of brachyuran larval descriptions. *Invertebrate Reproduction & Development*, 33, 127–144.
<http://dx.doi.org/10.1080/07924259.1998.9652627>
- Conde, J.E., Tognella, M.M.P., Paes, E.T., Soares, M.L.G., Louro, I.A. & Schaeffer-Novelli, Y. (2000) Population and life history features of the crab *Aratus pisonii* (Decapoda: Grapsidae) in a subtropical estuary. *Interciencia*, 25, 151–158.
- Cuesta, J.A. & Anger, K. (2001) Larval morphology of the sesarmid crab *Armases angustipes* Dana, 1852 (Decapoda, Brachyura, Grapoidea). *Journal of Crustacean Biology*, 21, 821–838.
<http://dx.doi.org/10.1163/20021975-99990175>
- Cuesta, J.A. & Schubart, C.D. (1999) First zoeal stages of *Geograpsus lividus* and *Goniopsis pulchra* from Panama confirm consistent larval characters for the subfamily Grapsinae (Crustacea: Brachyura: Grapsidae). *Ophelia*, 51, 163–176.
<http://dx.doi.org/10.1080/00785326.1999.10409406>
- Cuesta, J.A., García-Guerrero, M.U., Rodríguez, A. & Hendrickx, M.E. (2006a) Larval morphology of the sesarmid crab, *Aratus pisonii* (H. Milne Edwards, 1837) (Decapoda, Brachyura, Grapoidea) from laboratory-reared material. *Crustaceana*, 79, 175–196.
<http://dx.doi.org/10.1163/156854006776952838>
- Cuesta, J.A., Guerao, G., Liu, H.C. & Schubart C.D. (2006b) Morphology of the first zoeal stages of eleven Sesarmidae (Crustacea, Brachyura, Grapoidea) from the Indo-West Pacific, with a summary of familial larval characters. *Invertebrate Reproduction and Development*, 49, 151–173.
<http://dx.doi.org/10.1080/07924259.2006.9652206>
- Felder, D.L. & Martin, J.W. (2003) Establishment of a new genus for *Panopeus bedensis* Benedict & Rathbun, 1891 and several other xanthoid crabs from the Atlantic and Pacific oceans (Crustacea: Decapoda: Xanthoidea). *Proceedings of the Biological Society of Washington*, 116, 438–452.
- Flores, E.E. & Chien, Y.H. (2011) Chromatosomes in three phenotypes of *Neocardina denticulata* Kemp, 1918: morphological and chromatic differences measured non-invasively. *Journal of Crustacean Biology*, 31, 590–597.
<http://dx.doi.org/10.1651/11-3457.1>
- Fortunato, H. (2004) Reproductive strategies in gastropods across the Panama seaway. *Invertebrate Reproduction and Development*, 46, 139–148.
<http://dx.doi.org/10.1080/07924259.2004.9652617>
- Fransozo, A., Cuesta, J.A. & Negreiros-Fransozo, M.L. (1998) The first zoeal stage of two species of Grapsidae (Decapoda, Brachyura) and a key to such larvae from the Brazilian coast. *Crustaceana*, 71, 331–343.
<http://dx.doi.org/10.1163/156854098X00293>
- Giménez, L. (2006) Phenotypic links in complex life cycles: conclusions from studies with decapod crustaceans. *Integrative and Comparative Biology*, 46, 615–622.
<http://dx.doi.org/10.1093/icb/icb010>
- Giménez, L. & Anger, K. (2003) Larval performance in an estuarine crab, *Chasmagnathus granulata*, is a consequence of both larval and embryonic experience. *Marine Ecology-Progress Series*, 249, 251–264.
<http://dx.doi.org/10.3354/meps249251>
- Giménez, L. & Anger, K. (2005) Effects of temporary food limitation on survival and development of brachyuran crab larvae. *Journal of Plankton Research*, 27, 485–494.
<http://dx.doi.org/10.1093/plankt/fbi024>
- Giménez, K.A. & Torres, G. (2004) Linking life history traits in successive phases of a complex life cycle: effects of larval biomass on early juvenile development in an estuarine crab, *Chasmagnathus granulata*. *Oikos*, 104, 570–580.
<http://dx.doi.org/10.1111/j.0030-1299.2004.12957.x>
- Gore, R.H. (1972) *Petrolisthes armatus* (Gibbes, 1850): the development under laboratory conditions of larvae from a Pacific specimen (Decapoda, Porcellanidae). *Crustaceana*, 22, 67–83.
<http://dx.doi.org/10.1163/156854072X00688>
- Hamasaki, K. (2003) Effects of temperature on the egg incubation period, survival and developmental period of larvae of the

- mud crab *Scylla serrata* (Forskål) (Brachyura: Portunidae) reared in the laboratory. *Aquaculture*, 219, 561–572.
[http://dx.doi.org/10.1016/S0044-8486\(02\)00662-2](http://dx.doi.org/10.1016/S0044-8486(02)00662-2)
- Hart, M.W. (1995) What are the costs of small egg size for a marine invertebrate with feeding planktonic larvae? *The American Naturalist*, 146, 415–426.
<http://dx.doi.org/10.1086/285807>
- Hartnoll, R.G. (1965) Notes on the marine grapsid crabs of Jamaica. *Proceedings of the Linnean Society of London*, 176, 113–147.
<http://dx.doi.org/10.1111/j.1095-8312.1965.tb00940.x>
- Haugh, G.H. & Tielmann, R. (1998) Effect of the formation of the Isthmus of Panama on Atlantic Ocean thermohaline circulation. *Nature*, 393, 673–676.
<http://dx.doi.org/10.1038/31447>
- Ingle, R. (1992) *Larval stages of northeastern Atlantic crabs: an illustrated key*. Chapman & Hall, London, 363 pp.
- Jain, S. & Collins, L.S. (2007) Trends in Caribbean paleoproductivity related to the Neogene closure of the Central American Seaway. *Marine Micropaleontology*, 63, 57–74.
<http://dx.doi.org/10.1016/j.marmicro.2006.11.003>
- Kikkawa, T., Nakahara, Y., Hamano, T., Hayashi, K.I. & Miy, Y. (1995) Chromatophore distribution patterns in the first and second zoeae of atyid shrimps (Decapoda: Caridea: Atyidae): a new technique for larval identification. *Crustacean Research*, 24, 194–202.
- Lessios, H.A. (1990) Adaptation and phylogeny as determinants of egg size in echinoderms from two sides of the Isthmus of Panama. *The American Naturalist*, 135, 1–13
<http://dx.doi.org/10.1086/285028>
- Lessios, H.A. (2008) The great American schism: divergence of marine organisms after the rise of the Central American Isthmus. *Annual Review of Ecology, Evolution, and Systematics*, 39, 63–91.
<http://dx.doi.org/10.1146/annurev.ecolsys.38.091206.095815>
- Mantelatto, F.L., Reigada, Á.L., Gatti, A.C. & Cuesta, J.A. (2014) Morphology of the first zoeal stages of five species of the portunid genus *Callinectes* (Decapoda, Brachyura) hatched at the laboratory. *Anais da Academia Brasileira de Ciências*, 86, 755–768.
<http://dx.doi.org/10.1590/0001-3765201420130030>
- McAlister, J.S. (2008) Evolutionary responses to environmental heterogeneity in Central American echinoid larvae: plastic versus constant phenotypes. *Evolution*, 62, 1358–1372.
<http://dx.doi.org/10.1111/j.1558-5646.2008.00368.x>
- McEdward, L.R. & Miner, B.G. (2003) Fecundity-time models of reproductive strategies in marine benthic invertebrates: fitness differences under fluctuating environmental conditions. *Marine Ecology Progress Series*, 256, 111–121.
<http://dx.doi.org/10.3354/meps256111>
- Miura, O., Frankel, V. & Torchin, M.E. (2011) Different developmental strategies in geminate mud snails, *Cerithideopsis californica* and *C. pliculosa*, across the Isthmus of Panama. *Journal of Molluscan Studies*, 77, 255–258.
<http://dx.doi.org/10.1093/mollus/eyr012>
- Moran, A.L. (2004) Egg size evolution in tropical American arcid bivalves: the comparative method and the fossil record. *Evolution*, 58, 2718–2733.
<http://dx.doi.org/10.1111/j.0014-3820.2004.tb01624.x>
- Moran, A.L. & Allen, J.D. (2007) How does metabolic rate scale with egg size? An experimental test with sea urchin embryos. *The Biological Bulletin*, 212, 143–150.
<http://dx.doi.org/10.2307/25066591>
- Moran, A.L. & Emlet, R.B. (2001) Offspring size and performance in variable environments: field studies on a marine snail. *Ecology*, 82, 1597–1612.
[http://dx.doi.org/10.1890/0012-9658\(2001\)082\[1597:OSAPIV\]2.0.CO;2](http://dx.doi.org/10.1890/0012-9658(2001)082[1597:OSAPIV]2.0.CO;2)
- Ng, P.K. & Clark, P.F. (2000) The eumedonid file: a case study of systematic compatibility using larval and adult characters (Crustacea: Decapoda: Brachyura). *Invertebrate Reproduction and Development*, 38, 225–252.
<http://dx.doi.org/10.1080/07924259.2000.9652457>
- Shirley, S.M., Shirley, T.C. & Rice, S.D. (1987) Latitudinal variation in the Dungeness crab, *Cancer magister*: zoeal morphology explained by incubation temperature. *Marine Biology*, 95, 371–376.
<http://dx.doi.org/10.1007/BF00409567>
- Souza, A.S.D., Costa, R.M.D. & Abrunhos, F.A. (2013) Comparative morphology of the first zoea of twelve brachyuran species (Crustacea: Decapoda) from the Amazon region. *Zoologia Curitiba*, 30, 273–290.
<http://dx.doi.org/10.1590/S1984-4670201300030004>
- Steph, S., Tiedemann, R., Prange, M., Groeneveld, J., Nurnberg, D., Reuning, L., Schulz, M. & Haug, G. (2006) Changes in Caribbean surface hydrography during the Pliocene shoaling of the Central American Seaway. *Paleoceanography*, 21, 1–25.
<http://dx.doi.org/10.1029/2004PA001092>
- Terrossi, M., Cuesta, J.A., Wehrtmann, I.S. & Mantelatto, F.L. (2010) Revision of the larval morphology (Zoea I) of the family Hippolytidae (Decapoda, Caridea), with a description of the first stage of the shrimp *Hippolyte obliquimanus* Dana,

1852. *Zootaxa*, 2624, 49–66.
- Thatje, S. & Bacardit, R. (2000) Morphological variability in larval stages of *Nauticaris magellanica* (A. Milne Edwards, 1891) (Decapoda: Caridea: Hippolytidae) from South American waters. *Bulletin of Marine Science*, 66, 375–398.
- Thiercelin, N. & Schubart, C.D. (2014) Transisthmian differentiation in the tree-climbing mangrove crab *Aratus* H. Milne Edwards, 1853 (Crustacea, Brachyura, Sesarmidae), with description of a new species from the tropical eastern Pacific. *Zootaxa*, 3793 (1), 545–560.
<http://dx.doi.org/10.11646/zootaxa.3793.5.3>
- Warner, G.F. (1968) The larval development of the mangrove tree crab, *Aratus pisonii* (H. Milne Edwards), reared in the laboratory (Brachyura, Grapsidae). *Crustaceana, Supplement*, 2, 249–258.
- Wear, R.G. (1974) Incubation in British decapod Crustacea, and the effects of temperature on the rate and success of embryonic development. *Journal of the Marine Biological Association of the United Kingdom*, 54, 745–762.
<http://dx.doi.org/10.1017/S0025315400022918>
- Wehrtmann, I.S. & Albornoz, L. (2002) Evidence of different reproductive traits in the transisthmian sister species, *Alpheus saxidomus* and *A. simus* (Decapoda, Caridea, Alpheidae): description of the first postembryonic stage. *Marine Biology*, 140, 605–612.
<http://dx.doi.org/10.1007/s00227-001-0733-1>
- Wehrtmann, I.S. & López, G.A. (2003) Effects of temperature on the embryonic development and hatching size of *Betaeus emarginatus* (Decapoda: Caridea: Alpheidae). *Journal of Natural History*, 37, 2165–2178.
<http://dx.doi.org/10.1080/00222930210133291>
- Weiss, M., Thatje, S. & Heilmayer, O. (2010) Temperature effects on zoeal morphometric traits and intraspecific variability in the hairy crab *Cancer setosus* across latitude. *Helgoland Marine Research*, 64, 125–133.
<http://dx.doi.org/10.1007/s10152-009-0173-8>