



The echinoid fauna from middle and southern Japan: a preliminary report

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Abstract

Fifty-eight echinoid species from 22 families in nine orders were collected from middle and southern Japan. One species, *Aeropsis fulva*, is reported here for the first time from Japan, and four species, *Mespilia levituberculata*, *Pseudoboletia maculata*, *Lissodiadema purpureum* and *Echinodiscus tenuissimus* require systematic re-examination.

Introduction

In 2017 we launched a project aimed at summarizing the West Pacific echinoid fauna. As the first step, we have been reviewing the ecology and phylogeny of extant Japanese echinoids documented in the literature, supplemented by novel data obtained by fieldwork. These data are intended as a baseline for our future research on the echinoids living in greater West Pacific region.

Utinomi (1954, 1960) and Shigei (1981, 1986) summarized extant echinoid occurrences from the sea around Japan, and to date 112 echinoid species have been reported (Shigei 1986). However, in their works, ecological information of the species was scant or missing, especially concerning habitat information. Since these early studies, no comprehensive study of Japanese echinoids has been published. In the current project, sea urchins were collected from their natural habitats primarily by scuba diving and detailed ecological information was recorded. Apart from a thorough morphological taxonomic evaluation, freshly collected specimens are also subjected to molecular genetic analyses to validate their phylogenetic relationships. Here, we give a preliminary report of our fieldwork in middle and southern Japan in 2017 and 2018, during which we were able to observe and sample 58 different echinoid species accounting for more than half of currently described Japanese echinoids.

Sampling

Sampling was carried out in the following localities in middle to southern Japan (Fig. 1): by hand in the intertidal zone of Shirahama Beach (35°09'14" N, 139°08'41" E), Sesoko Island (26°38'04" N, 127°51'53" E) and Ginama (26°50'19" N, 128°15'00" E) and from drifting seaweeds in Owase Bay (34°04'22" N, 136°12'58" E); by scuba-diving in Aburatsubo (35°09'26" N, 139°36'40" E) (Misaki Marine Biological Station), Toi (34°54'24" N, 138°47'07" E), Takugi (36°09'49" N, 133°15'24" E), Komame (32°47'29" N, 132°41'42" E), Tachibanaura (32°49'57" N, 132°39'44" E), Kasiwa Island (32°46'31" N, 132°38'07" E), Issai (32°47'11" N, 132°38'04" E), Nishidomarisurugi-no-Hama (32°46'39" N, 132°43'55" E), Oura Bay (26°32'30" N, 128°03'15" E), Mizugama (26°21'36" N, 127°44'18" E), and Manzamo (26°30'15" N, 127°50'38" E); by

dredging in Sagami Bay using by aboard the vessel R/V Rinkai-maru of the Misaki Marine Biological Station, off Tosashimizu and in Kumanonada by aboard the vessel R/V Seisui-maru of Mie University.

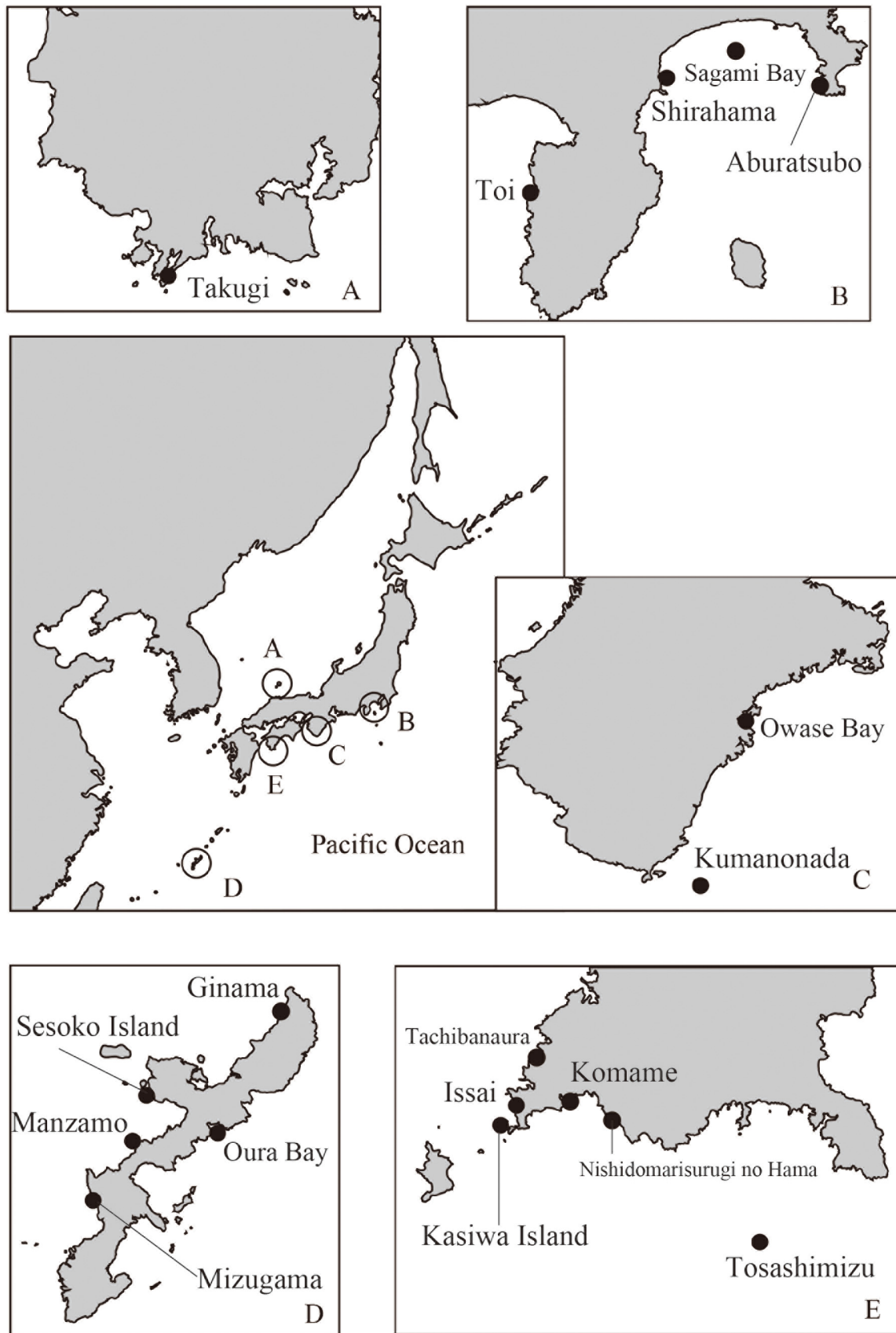


FIGURE 1. Map of sampling localities.

Sea urchins collected in this study

The fifty-eight species from 22 families of nine orders collected during our investigations are summarized in Table 1. *Aeropsis fulva* was recorded for the first time from Japan; a brief discussion on the new record is given below. Four species, *Mespilia levituberculata*, *Pseudoboletia maculata*, *Lissodiadema purpureum*, and *Sculpstechinus tenuissimus* require systematic re-examination. Brief comments for these species are given below.

TABLE 1. List of echinoid species collected from middle and southern Japan.

Order	Family	Species	Locality	Water Depth (m)	
Cidaroida	Cidaridae	<i>Acanthocidaris maculicolis</i>	Tosashimizu	115	
		<i>Eucidaris metularia</i>	Kasiwa Island	10	
		<i>Goniocidaris biserialis</i>	Tosashimizu	120	
		<i>Phyllacanthus</i> sp.	Mizugama	5	
		<i>Plococidaris verticillata</i>	Nishidomarisurugi-no-Hama	5–7	
		<i>Prionocidaris baculosa</i>	Kasiwa Island	10–20	
		<i>Rhopalocidaris gracilis</i>	Tosashimizu	120	
		<i>Stylocidaris bracteata</i>	Tosashimizu	115	
		<i>Chorocidaris fusispina</i>	Tosashimizu	110	
		<i>Stylocidaris reini</i>	Tosashimizu	120–150	
Echinothurioida	Echinothuriidae	<i>Araeosoma owstoni</i>	Kumanonada	740	
		<i>Asthenosoma ijimai</i>	Issai	40	
		<i>Calveriosoma gracile</i>	Kumanonada	450	
	Phormosomatidae	<i>Phormosoma bursarium</i>	Kumanonada	557	
Diadematoidea	Diadematidae	<i>Diadema clarki</i>	Kasiwa Island	10	
		<i>Diadema setosum</i>	Tachibanaura	6	
		<i>Echinothrix calamaris</i>	Komame	8	
		<i>Lissodiadema purpureum</i>	Mizugama	3–8	
Stomopneustoida	Stomopneustidae	<i>Stomopneustes variolaris</i>	Kasiwa Island	10	
Pedinoida	Pedinidae	<i>Caenopedina mirabilis</i>	Kumanonada	218	
Camarodonta	Echinidae	<i>Gracilechinus lucidus</i>	Kumanonada	217–557	
		Temnopleuridae	<i>Mespilia levituberculata</i>	Aburatsubo	5
			<i>Microcyphus excentricus</i>	Mizugama	3–8
	<i>Temnopleurus toreumaticus</i>		Owase Bay	1	
	Toxopneustidae	<i>Pseudoboletia maculata</i>	Tachibanaura	8	
		<i>Toxopneustes elegans</i>	Tachibanaura	8	
		<i>Toxopneustes pileolus</i>	Komame	8	
		<i>Tripneustes gratilla</i>	Komame	8	
	Strongylocentrotidae	<i>Pseudocentrotus depressus</i>	Shirahama-beach	Intertidal zone	
		<i>Hemicentrotus pulcherrimus</i>	Shirahama-beach	Intertidal zone	
	Echinometridae	<i>Colobocentrotus mertensii</i>	Ginama	Intertidal zone	
		<i>Echinometra mathaei</i>	Tachibanaura	5	
		<i>Echinometra oblonga</i>	Sesoko Island	Intertidal zone	
		<i>Echinometra</i> sp. A	Komame	5	
		<i>Echinometra</i> sp. C	Sesoko Island	Intertidal zone	
		<i>Echinostrephus aciculatus</i>	Tachibanaura	5	
<i>Echinostrephus molaris</i>		Mizugama	3–7		
<i>Heliocidaris crassispina</i>		Shirahama-beach	Intertidal zone		
<i>Heterocentrotus mamillatus</i>		Mizugama	2		

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TABLE 1. (Continued)

Order	Family	Species	Locality	Water Depth (m)
Clypeasteroida	Clypeasteridae	<i>Clypeaster japonicus</i>	Toi	10
		<i>Clypeaster virescens</i>	Tosashimizu	120
	Laganidae	<i>Laganum fudsiyama</i>	Kumanonada	190
		<i>Peronella japonica</i>	Aburatsubo	7
	Scutellidae	<i>Scaphechinus mirabilis</i>	Sagami Bay	3
	Astriclypeidae	<i>Astriclypeus manni</i>	Aburatsubo	7
<i>Sculpsitechinus</i> sp.		Oura Bay	10	
Holasteroida	Pourtalesiidae	<i>Pourtalesia laguncula</i>	Kumanonada	557
Spatangoida	Schizasteridae	<i>Moira lachesinella</i>	Takugi	10
		<i>Aeropsis fulva</i>	Kumanonada	834
	Brissidae	<i>Brissopsis luzonica</i>	Kumanonada	834
		<i>Brissus agassizii</i>	Takugi	10
		<i>Metalia</i> sp.	Takugi	9
	Eurypatagidae	<i>Linopneustes murrayi</i>	Kumanonada	740–834
		<i>Platybrissus roemeri</i>	Mizugama	8
	Maretiidae	<i>Maretia planulata</i>	Oura Bay	10
		<i>Nacospatangus altus</i>	Takugi	5–18
	Loveniidae	<i>Lovenia elongata</i>	Takugi	5–9
		<i>Lovenia gregalis</i>	Kumanonada	740

Aeropsis fulva (A. Agassiz, 1898)1898 *Aërope fulva* A. Ag.—Agassiz: 81; pl. 8: figs 5–6.1950 *Aëropsis fulva* (A. Agassiz).—Mortensen: 329.

Remarks: The highly modified test with a peculiar spindle-shaped form and position of the periproct above the ambitus make members of the genus *Aeropsis* immediately recognizable. Currently (Kroh & Mooi 2018) two species of the genus have been described, which can usually be distinguished based on their number of gonopores, being two in the Atlantic species *A. rostrata* (Norman, 1876) (see e.g. Mortensen 1907: fig. 15 or Kroh 2004: fig. 2/3d) and four in the Pacific *A. fulva* (see Agassiz 1904: figs. 281, 282). Although specimens with two gonopores have been reported in *A. fulva* too (e.g. Agassiz 1904: fig. 283), none with four pores are known in *A. rostrata*. Combined with the anterior position of the highest point of the test and the low test height (compare the key in Mortensen 1950: p. 327) the new Japanese specimen, which shows four gonopores thus can be confidently placed in *A. fulva*.

This species was initially described from the East Pacific, having been caught at several stations in the region south of Panama during the Albatross Cruise (Agassiz 1898: p. 81). Its range was later extended to deep waters off the coasts of Peru, the Bering Sea and Arafura Sea (see Mortensen 1950). Specimens from deep water off the coasts of California, Oregon and Washington State are recorded in collection of the California Academy of Sciences (<https://www.gbif.org/species/5188153>). The new occurrence in Japan is consistent with those earlier reports and suggests a wide distribution throughout the Pacific Ocean. The lack of records from large parts of this area might be explained by the depth inhabited by this species and its fragile test, both of which hamper collection.

Occurrence in Japan: Kumanonada, at a depth of 834 m.

Mespilia levituberculata Yoshiwara, 1898

Remarks: All echinoids of the genus *Mespilia* living around Japan have been attributed to *Mespilia globulus* since Shigei (1986) summarized Japanese echinoids, though he noted that Japanese specimens are morphologically very different from the specimens from other Indo-West-Pacific regions. Japanese *Mespilia* have originally been described as *M. levituberculata* by Yoshiwara (1898). However, this species has been synonymized with *M. globulus* by Clark (1912). Mortensen (1943), in contrast, suggested to “maintain the

Japanese form *Mespilia levituberculata* Yoshiwara to be, at least, a well characterized variety, if not rather a distinct species” and stated that “var. *levituberculata* is known only from Japan, from Sagami Bay to Kiushu”. Specimens of *Mespilia* collected in this study were provisionally identified as *M. levituberculata* due to their striking morphological difference to *Mespilia globulosa* from e.g., the Philippines. In-depth examinations of the phylogenetic relationship of the two forms are currently under way.

Pseudoboletia maculata Troschel, 1869

Remarks: Shigei (1986) described only one species from the genus *Pseudoboletia*, i.e. *P. indiana*, from Japan. *Pseudoboletia* specimens collected during the present study were identified as *P. maculata* based on their test bearing conspicuous dark spots. Utinomi (1954) was the first to report *Pseudoboletia maculata* from Japan and coined the Japanese name, “Madara-Uni” for this species. Shigei (1981) also reported that *Pseudoboletia* species collected around Japan as *P. maculata*. In Shigei’s (1986) faunal list of Japanese echinoids, however, *Pseudoboletia maculata* was missing, and instead *P. indiana* was included, associated with the Japanese name “Madara-Uni”. He noted that the specimens having a denuded test with whitish color should be attributed to *P. indiana*. A phylogenetic study to confirm the morphological identification of these specimens is currently in progress.

Lissodiadema purpureum (A. Agassiz & H.L. Clark, 1907)

Remarks: We collected the specimens from the same locality where Obuchi *et al.* (2013) reported the occurrence of *Lissodiadema lorioli*. Based on morphological observations, however, the newly collected material fits better with *L. purpureum* than *L. lorioli*. *L. purpureum* was originally described as *Leptodiadema purpureum* based on a single juvenile specimen from Hawaii. However, the genus *Leptodiadema* has since been synonymized with *Lissodiadema* (Schultz in Smith & Kroh, 2007) based on the examination of adult specimens found in Guam. A comprehensive description of this species, however, is still missing. We are currently working on a detailed re-description of the species including a molecular genetic analysis.

Sculpstechinus tenuissimus (L. Agassiz & Desor, 1847)

Remarks: This sand dollars with two lunules in the posterior ambulacra occurs around Okinawa. It was first recorded by Shigei (1986) under the name *Echinodiscus tenuissimus*. Stara & Sancier (2014) have recently revised the classification of fossil and recent astriclypeid echinoids and have established several new genera in this family. The Japanese specimens show features supporting the attribution to the genus *Sculpstechinus*; however, the neotype of *S. tenuissimus* from New Caledonia, designated by Stara & Sancier (2014) is very different from Japanese *Sculpstechinus* specimens and the conspecificity of the two populations needs to be re-evaluated and thoroughly explored.

Gracilechinus lucidus (Döderlein, 1885)

Remarks: Shigei (1986) included *Echinus lucidus* in the Japanese echinoid list. Currently this taxon is known as *Gracilechinus lucidus* (see Kroh & Mooi, 2018).

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