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# Phylum Nematoda: trends in species descriptions, the documentation of diversity, systematics, and the species concept

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## Abstract

This paper summarizes the trends in nematode species description and systematics emerging from a comparison of the latest comprehensive classification and census of Phylum Nematoda (Hodda 2022a, b) with earlier classifications (listed in Hodda 2007). It also offers some general observations on trends in nematode systematics emerging from the review of the voluminous literature used to produce the classification. The trends in nematodes can be compared with developments in the systematics of other organisms to shed light on many of the general issues confronting systematists now and into the future.

#### Introduction

The phylum Nematoda was only formally recognized relatively recently (Cobb 1932). Despite nematodes being known from ancient times, their classification has changed many times (summarized by Hodda 2007). Changes in classification have occurred at all taxonomic levels, from sub-specific to classes (and even included whether nematodes themselves should be considered a class within a larger phylum or a phylum themselves).

The frequent changes in classification have perhaps arisen because nematodes pose many systematic challenges. They are very diverse, with nearly 30,000 species described so that keeping track of the classification of the entire phylum is a substantial task: Nematoda is the phylum with 5<sup>th</sup> most described species, less than Arthropoda, Mollusca, Craniata and just less than Platyhelminthes, but more than Echinodermata, Annelida and Cnidaria (Hodda 2022a, Zhang 2011, 2013). Not only are there many species described, but there are probably a much larger number undescribed: even the lowest estimates of total nematode species numbers start at half a million species, and the highest estimates are more than 10 million species (Hodda 2022a). They are also mostly small, cryptic and morphologically simple, meaning that diagnosis by traditional morphological characters is often hard. But, for small, simple organisms, the genetic relationships among nematodes can be highly complex, meaning that molecular diagnoses also pose many challenges.

The original construction and more recent update of a comprehensive classification and census of all valid species and genera of nematodes (Hodda 2007, 2011, 2022a, b) has afforded the opportunity to examine trends in nematode systematics, and the nature of the challenges they reveal. Common trends and issues in nematode systematics emerging from the review of several thousand taxonomic publications are examined. Some of the issues raised in using these publications to construct a classification of all nematodes and a census of species have emerged recently and others are more long-standing. These trends are compared with some previous assessments of nematode species numbers and rates of species descriptions (Allen & Sher 1967, Hammond 1992, Hodda 2011, Hugot *et al.* 2001, Hyman 1951, Mayr *et al.* 1953, Stork 1993). The trends in nematode systemtics are compared and contrasted with trends and developments in the systematics of other organisms to perhaps illuminate many of the general issues confronting systematists now and those likely to emerge in the future. In this comparison, the emphasis is on the consequences of documented biological features of nematodes and other organisms for practical aspects of systematics. An extensive discussion of theoretical issues in the taxonomic system has been presented recently by Dubois *et al.* (2021).

## **Rate of species descriptions**

The new classification and census adds about 3800 new species in over 300 new genera, including fossil species as well (Hodda 2011, Hodda 2022a).

This rate of new nematode species descriptions can be compared with the rate of descriptions of other invertebrates detailed by Zhang (2021) when those data are adjusted for a similar time interval. When this is done, the absolute number of new nematode species descriptions from 2011 to 2019 is higher than that for many less speciose groups, but less than that in groups where more species have been described (Table 1). As a percentage of the number of described species, new descriptions of nematodes are near the high end of the range of values for other taxonomic groups (Table 1). This means that for many groups between about one sixth and one ninth of the currently recognized species have been described in the last decade: of the over 28 500 valid species and 3000 valid genera of nematodes currently recognized, about one eighth were described in the 9 years between the censuses—from end 2010 (Hodda 2011) to end 2019 (Hodda 2022a)—so nematodes were in the middle of this range. While this may appear encouraging, it is important to note that as a percentage of the estimated total number of species, the rate of new species descriptions for nematodes is the lowest of any in the table. The implications of this are discussed further below.



**FIGURE 1.** Descriptions of new nematode species per year. Circles are for all publications, triangles for the journal *Zootaxa* only. Filled symbols represent arithmetic mean rates per year for the period surrounding the point. Empty symbols represent estimates at the time. See text for full description. Trend lines are illustrative only.

Trends in the rate of nematode species and genus descriptions over time can be estimated thanks to some earlier summaries (Allen & Sher 1967, Hammond 1992, Hodda 2011, Hugot *et al.* 2001, Hyman 1951, Mayr *et al.* 1953, Stork 1993) (Figure 1). From the 3 nematode species named in the first formal binomial nomenclature (Linnaeus 1758) to the 10000 known in 1953 (Mayr *et al.* 1953) new nematode species descriptions therefore averaged about 50 per year for the nearly 200 first years of formal taxonomy. This figure of course obscures changes within the period, with an initially slower rate of species descriptions perhaps accelerating towards the end of the period. There were, however, periods within this time where large numbers of species were described in substantial monographs (e.g. Bastian 1865, Cobb 1920). By 1950, the total number of nematode species descriptions was estimated at about 200 per annum in the preceding few years (Hyman 1951). By 1966, the estimate for parasites of plants and free-

living nematodes only was about 200 new species per annum (Allen & Sher 1967), which is about 300 species per annum if corrected for the estimated proportion of animal parasites not included (Hodda 2022a). Between 1993 and 2010, several estimates put the rate of new nematode species descriptions substantially above 300 new species per annum (Hammond 1992, Hodda 2011, Hugot *et al.* 2001, Stork 1993). The current estimate of total number of nematode species descriptions per year has increased to very nearly 400 new species per annum (Hodda 2022a). Thus it appears that the overall rate of species descriptions of nematodes is increasing, but only relatively slowly: there has been no explosion in new species descriptions thanks to genetic technologies.

Taxonomic group	Described spp	Estimated total number of spp	New spp 2011–2019	New spp as % of described spp	New spp as % of total spp	References
Porifera	9,500	30,000	350	4	1.1	Hooper et al. 2021
Bryozoa	6,000	10,000	450	8	4.5	Bock & Gordon 2013, Gordon 1999, Gordon & Bock 2021
Annelida	20,000	30,000	1,600	8	5.3	Chapman 2009, Magalhaes et al. 2021
Tardigrada	1,500	15,000	200	13	1.3	McInnes et al. 2021
Nematoda	29,000	<sup>1</sup> 500,000	3,800	13	0.8	Hodda 2021, 2022a
Opiliones	7,000	10,000	350	5	3.5	Coddington & Colwell 2001, Kury 2013, Perez- Gonzalez <i>et al</i> . 2021
Araneae	44,000	100,000	7,000	16	7.0	Jaeger <i>et al.</i> 2021, Zhang 2013
Acari	55,000	500,000	6,000	11	1.2	Skoracka <i>et al.</i> 2015, Zhang 2013, Zhang <i>et al.</i> 2021
Minor Insect orders <sup>2</sup>	12,000	30,000	450	4	1.5	Bernard & Whittington 2021, Stork 2018
Blattodea	7,600	28,000	250	3	0.9	Beccaloni & Eggleton 2013, Djernaes 2018, Stork 2018
Ephemeroptera	4,000	5,000	600	15	12.5	Jacobus et al. 2021
Odonata	6,000	7,000	500	8	7.1	Dijkstra <i>et al.</i> 2013, Paulson & Marinov 2021
Diptera	160,000	800,000	12,400	8	1.6	Whitmore <i>et</i> al. 2021, Stork 2018, Zhang 2013
Amphibia	8,300	20,000	1,400	17	7.0	Rivera-Correa et al. 2021
Mammalia	6,500	7,000	300	5	5.0	Burgin <i>et al.</i> 2018, Cordiero-Estrela <i>et al.</i> 2021, Fisher <i>et al.</i> 2018

**TABLE 1.** Approximate numbers of new species described from 2011 to 2019 (inclusive) relative to numbers of described species and estimated total species numbers for selected animal groups. All numbers approximate and rounded to allow for uncertainty in any estimate of species numbers (Chapman 2009, May 1988).

The annual rate of new descriptions remains a very small proportion of the estimated total number of species less than 0.1% of an estimated 500 000 to 1 million species (Hugot *et al.* 2001, Lambshead 1993). (Note that the values in Table 1 are for the period 2011–2019 and not annual rates.) If the estimates of the total number of nematode species are accurate, this equates to over a millenium to describe all species at the current rate, even for the lower estimate! (See below for further discussion of the total numbers of nematode species.)

<sup>1</sup> Lowest estimate, see Hodda (2022a) and Hodda & Khudhir (2022) for discussion of higher estimates.

<sup>2</sup> Including the insects Archaeognatha (Microcoryphia), Dermaptera, Embioptera, Siphonaptera, Zoraptera and Zygentoma, as well as the hexapod classes Protura and Diplura.

Because the recent classification includes authority and date for all valid nematode genera, the increase in number of genera and hence rate of proposal of nematode genera can be tracked more precisely than number of species (Figure 2). Using this data, the number of genera proposed has shown a steady increase, but with a notable temporary decrease in the decade from 1940 to 1950, and also a more recent, consistent trend to fewer proposals in the 3 decades since 1990 (Figure 2). While the earlier decrease is undoubtedly related to wars disrupting descriptive taxonomy during the decade, the cause of the recent decrease is less clear. It may be related to uncertainty in converting clades identified by molecular analyses into higher taxonomic groups (Dubois *et al.* 2021).



**FIGURE 2.** Proposal of new nematode genera per decade (columns) and total number of valid genera (points and line). The columns represent the total increase in the decade. The points and line represent the total number of valid genera at the end of the decade; i.e. the column for 2010 represents the genera added from the beginning of 2001 to the end of 2010, and the point for 2010 represents the number of genera at the end of 2010.

## Sources of species descriptions

If the number of species described is not changing much, what does seem to be changing is the journals in which descriptions have been published. Summaries of publications describing nematodes in the journal *Zootaxa* (Hodda 2021, Xu *et al.* 2013, Zhao 2007), plus the current data, show a doubling of the rate of descriptions published in *Zootaxa* approximately every 3 to 4 years (Figure 1). This is after growing from 2 new species in the 3<sup>rd</sup> year of the journal in 2003, to 75 new species in 2020

Notwithstanding potential biases in some perceptions of *Zootaxa* as the only journal to publish species descriptions (Pinto *et al.* 2021), the publication seems to be responsible for an increasing proportion of new species descriptions (Figure 1). In the period from the last census (Hodda 2011) to the current one (Hodda 2022a), the journal *Zootaxa* has accounted for almost 20% of the total (over 600 species) (Hodda 2021). This proportion is increasing with some variation from year to year: in 2020 *Zootaxa* accounted for 25% of the total number of new nematode descriptions, which was just over 300. This is close to the proportion for the journal as a whole (Zhang 2021).

For genera proposed, *Zootaxa* has accounted for a slightly smaller but still substantial percentage (a total of 38 genera of the 285 proposed since 2000, or 13%). Although numbers of genera proposed per year are small and hence vary substantially from year to year, the proportion of genera published in *Zootaxa* seems to be growing as well: in 2020 *Zootaxa* published 4 of 11 new genera, or 36%.

The significance *Zootaxa* in documenting biodiversity has been discussed by several authors (Hodda 2021, Pinto *et al.* 2021, Zhang 2011, 2021).

## New descriptions vs synonymizations

The current total number of valid species of nematodes in the latest census (over 28 000 in over 3000 genera) includes a substantial number of reinstatements of previously characterized but subsequently synonymized species and genera. Analysis of molecular data has often implied that older taxa often consist of multiple species, and that proposed species from different geographic regions which appear very similar and are hence often synonymized, are in fact often really different (Brandt *et al.* 2007, Derycke *et al.* 2009, Grosemans *et al.* 2016, Guden *et al.* 2018, Perez-Ponce de Leon & Poulin 2018). However, this has not always proved the case, and molecular evidence has indicated that some species are very widespread and variable, and hence have been described under multiple names that should indeed be regarded as synonyms (De Groote *et al.* 2017, De Oliviera *et al.* 2014, 2017). Overall, reinstatements of previous names have outnumbered synonymizations, but proposals of new species based on new specimens have vastly outnumbered changes involving existing names.

# **Distribution of new descriptions**

In all publications since 2011 describing new nematode species, several trends in new species descriptions are apparent. These trends show that descriptions of new nematode species are often not a reflection of the true diversity of nematodes, but are heavily biased by geographic, economic and technical or methodological influences unrelated to the organisms themselves or their underlying diversity.

For example, most descriptions were from terrestrial habitats, freshwater aquatic environments or from terrestrial or freshwater aquatic hosts (2400 species). Only 30% of species were described from marine environments or hosts (about 1100 species). This is despite a much greater part of nematode phylogenetic diversity coming from marine habitats, as measured by the number of higher taxa (Hodda 2011, 2022a). Marine nematode diversity may be seriously underestimated (Brandt *et al.* 2007, Fonseca *et al.* 2010a, Hodda *et al.* 2009, Lambshead 2004, Lambshead & Boucher 2003). However, getting true estimates of the diversity of marine nematodes may be difficult because of the vast areas of coastline and ocean bed, and the logistic or methodological difficulties of sampling it (Hodda 1990, Hodda & Khudhir 2022, Lambshead & Hodda 1994). Marine nematodes were until recently underrepresented in molecular studies, too (Bik *et al.* 2010, Meldal *et al.* 2007).

The number of new species described seems strongly related to human resources applied to taxonomic effort. In the last few years, most new nematode species have been described from Asia, particularly the emerging economies of China and India (Table 2). By contrast, the fewest species by far were described from Africa, where nematological training and expertise remains sparse (Coyne *et al.* 2018, Wacekel *et al.* 2010). Relative to total surface of the earth or total land area, North America was also under-represented in new descriptions. One can only speculate on whether the underlying causes might be similar (because of little training in nematode taxonomy), or different (because of a much better-studied fauna meaning that there are fewer new species encountered). Notable also is that far more species were described from Australasia than would be expected from its scientific population (about 3% of the world's total overall: OECD 2011). This is related to several very active taxonomists having time to describe species in retirement freed from other commitments (Davies, Spratt, Smales). There are serious concerns that this situation will not continue, as there are few young people in training to replace them and succession planning has been limited (Hodda *et al.* 2017, 2019, Howie 2012).

Other phyla parasitic in vertebrates can have very different geographic distributions for new species descriptions (Poulin & Jorge 2019, Poulin *et al.* 2019), so the pattern of new species descriptions for nematodes may not be indicative of a more general trend.

On a continental scale, the geographic distribution of new nematode species descriptions is substantially different to both the land and total areas (including surrounding seas and oceans) (Table 2). Asia and Europe had a much larger proportion of species described than their areas, while the Americas and Africa had many fewer species described than their areas. The total proportion of species described from Australasia (14%) was close to the total area of land and water as a proportion of earth's surface (14%). However, water forms a large part of this area,

including large parts of the South Pacific and Indian Oceans, and compared to the land area (6% of the total), the number of species described was very large because most of the new species descriptions were terrestrial rather than marine. Few new species have been described from Antarctica and surrounding waters, although the number of new species is substantial relative to the small number of known species from Antarctica and the estimated total number of species on and around that continent.

Of particular note is that several species were described, not from their presumed native location, but from quarantine interceptions in other places (apparently completely remote from the assumed origin of the specimens) (Table 2). In all of these cases, the original source of the specimens could not be positively identified, even though the new species could be positively described and diagnosed from the specimens intercepted. They may have come from the origin of the produce on which they were found, they may have come locally from the destination, or any place along the transport route of the produce. As the movement of plant and animal products has expanded, trade networks have become increasingly important in the dispersal of organisms generally (Banks *et al.* 2015). Likewise trade hubs may become increasingly important as centres where produce from many different places not otherwise sampled can be examined (Hodda *et al.* submitted). The implication of this is that finding new species some distance from their natural geographic range and habitat may become an increasing trend.

**TABLE 2.** Total number and % of described new species of nematodes from 2011 to 2019 listed by continent, together with their total area as a % of total earth's surface, ocean area as a % of earths total surface, and land area as a % of earth's total land area. Note that some new species were described from more than one continent.

Continent	new species described	% new species described	% total surface area of earth	Ocean as % of total area of earth	% total land area
Asia <sup>1</sup>	1672	47	22	9	30
North America <sup>2</sup>	330	9	19	14	17
South America <sup>3</sup>	416	12	17	13	12
Africa <sup>4</sup>	191	5	15	9	20
Australasia <sup>5</sup>	486	14	14	11	6
Europe <sup>6</sup>	439	12	8	6	7
Antarctica <sup>7</sup>	7	0	5	4	9
Trade <sup>8</sup>	19	1	??	??	??

<sup>1</sup> including adjacent seas and parts of the North Pacific, Indian and Arctic Oceans

<sup>2</sup> including adjacent seas and parts of the North Pacific, North Atlantic and Arctic Oceans

<sup>3</sup> including adjacent seas and parts of the South Pacific and South Atlantic Oceans

<sup>4</sup> including adjacent seas and parts of the South Atlantic and Indian Oceans

<sup>5</sup> including adjacent seas and parts of the South Pacific and Indian Oceans

<sup>6</sup> including adjacent seas and parts of the North Atlantic and Arctic Oceans

<sup>7</sup> including adjacent seas and parts of the Southern Ocean

<sup>8</sup> also included in total for origin of material where known, but not in total for place of interception

## Ecological associations of new species

The description of new species was strongly biased by their life habits—whether they were free-living, or associated with vertebrates, invertebrates or plants. More free-living microbivorous or predatory nematodes were described than any other group, with nematodes associated with vertebrates having next most species described, and invertebrateand plant-associated nematodes fewest species described (Table 3). This is roughly the pattern in the currently described species, but is vastly different to the estimated total number of species that really exist, where there are far more free-living species than other types (Hodda *et al.* 2009).

Significantly, the estimated total number of nematode species associated with invertebrates has increased as a result of extensive collecting and detailed molecular work concentrated on particular taxonomic groups of nematodes or hosts (Bartholomaeus *et al.* 2012, Davies *et al.* 2010a, b, 2012a, b, 2013a, b, 2014a, b, c, d, e, 2015, Giblin-Davis *et al.* 2014, Kanzaki *et al.* 2014, Zhao *et al.* 2013a, b, 2015). Similar increases in numbers of species

have been observed in some nematode groups associated with particular vertebrates (Fenner *et al.* 2011, Justine & Iwaki 2014, Moravec & Ali 2014, Moravec & Barton 2015, Moravec & Diggles 2014a, b, 2015a, b, Moravec & Jirku 2014a, b, 2015, Moravec & Justine 2014a, b, 2015a, b, Moravec & Monoharan 2014a, b, Moravec & Van As 2015a, b, Moravec *et al.* 2014, 2015a, b, Purwaningsih & Smales 2010, 2011, 2014, Smales 2010, 2011a, b, c, 2012, 2013, 2014, 2015, Smales & Heinrich 2010, Spratt 2010, 2011, Weaver & Smales 2010, 2012). This highlights that the interests and areas of expertise of the most active systematists can have a strong influence on apparent diversity of particular groups, rather than the real biological diversity.

**TABLE 3.** Number and percentage of new nematode species described from 2011 to 2019 by gross ecological association.

Habitat	Species	percentage		
free-living	1595	44		
vertebrate-associated	1013	28		
invertebrate-associated	447	12		
plant-associated	535	15		
all	3590	100		

**TABLE 4.** Numbers of new species described from 2011 to 2019 in nematode genera with more than 5 species described, listed in decreasing order of number of newly-described species, with their classification and gross ecological association (f—free living; p—plant associated; i—invertebrate associated; v—vertebrate associated).

Genus	Order	f	р	i	v	new spp.	total spp in genus <sup>1</sup>
Philometra	Spirurida	0	0	0	1	<sup>2</sup> 57	135
Bursaphelenchus	Panagrolaimida	0	1	1	0	36	135
Steinernema	Panagrolaimida	0	0	1	0	36	105
Fergusobia	Panagrolaimida	0	1	0	0	32	49
Aphelenchoides	Panagrolaimida	0	1	0	0	26	193
Xiphinema	Dorylaimida	0	1	0	0	25	255
Longidorus	Dorylaimida	0	1	0	0	24	159
Pristionchus <sup>3</sup>	Diplogasterida	1	0	0	0	<sup>2</sup> 22	56
Cloacina	Rhabditida	0	0	0	1	19	140
Caenorhabditis <sup>3</sup>	Rhabditida	1	0	0	0	<sup>2</sup> 18	38
Cucullanus	Spirurida	0	0	0	1	16	173
Sabatieria	Monhysterida	1	0	0	0	16	90
Schistonchus	Panagrolaimida	0	1	1	0	13	21
Daptonema	Monhysterida	1	0	0	0	12	153
Aporcelinus	Dorylaimida	1	0	0	0	11	25
Oscheius	Rhabditida	0	0	1	0	11	47
Procamallanus	Spirurida	0	0	0	1	11	195
Rhabdochona	Spirurida	0	0	0	1	10	159
Ditylenchus	Panagrolaimida	0	1	0	0	9	80
Paratylenchus	Panagrolaimida	0	1	0	0	9	170
Pratylenchus	Panagrolaimida	0	1	0	0	9	122
Trichuris	Trichocephalida	0	0	0	1	9	98
Deladenus	Panagrolaimida	0	0	1	0	8	32
Ektaphelenchoides	Panagrolaimida	0	0	1	0	8	30
Ficophagus	Panagrolaimida	0	1	1	0	8	16
Meloidogyne	Panagrolaimida	0	1	0	0	8	100

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Genus	Order	f	р	i	v	new spp.	total spp in genus <sup>1</sup>
Oncholaimus	Oncholaimiida	1	0	0	0	8	121
Parapharyngodon	Spirurida	0	0	0	1	8	72
Rhabdias	Rhabditida	0	0	0	1	8	73
Syphacia	Spirurida	0	0	0	1	8	68
Tripylella	Tripylida	1	0	0	0	8	11
Aplectana	Spirurida	0	0	0	1	7	58
Aulolaimus	Mermithida	1	0	0	0	7	19
Hysterothylacium	Spirurida	0	0	0	1	7	115
Litinium	Ironida	1	0	0	0	7	13
Meteterakis	Spirurida	0	0	0	1	7	27
Phasmarhabditis	Rhabditida	0	0	1	0	7	18
Philometroides	Spirurida	0	0	0	1	7	32
Rotylenchus	Panagrolaimida	0	1	0	0	7	104
Sectonema	Dorylaimida	1	0	0	0	7	33
Trischistoma	Tripyloidida	1	0	0	0	7	16
Axonchium	Dorylaimida	1	0	0	0	6	37
Campylaimus	Monhysterida	1	0	0	0	6	23
Cosmocercoides	Spirurida	0	0	0	1	6	28
Crassonchus	Dorylaimida	1	0	0	0	6	6
Desmodora	Desmodoridae	1	0	0	0	6	72
Dichelyne	Spirurida	0	0	0	1	6	56
Dorylaimopsis	Monhysterida	1	0	0	0	6	26
Epacanthion	Enoplida	1	0	0	0	6	27
Geomonhystera	Monhysterida	1	0	0	0	6	33
Haliplectus	Plectida	1	0	0	0	6	25
Heth	Spirurida	0	0	0	1	6	44
Paramphimonhystrella	Monhysterida	1	0	0	0	6	9
Parasphaerolaimus	Monhysterida	1	0	0	0	6	9
Parodontophora	Monhysterida	1	0	0	0	6	36
Pseudaphelenchus	Panagrolaimida	0	0	1	0	6	6
Rhynchonema	Monhysterida	1	0	0	0	6	28
Ruehmaphelenchus	Panagrolaimida	0	0	1	0	6	8
Serpentirhabdias	Rhabditida	0	0	0	1	6	12
Tylencholaimus	Dorylaimida	1	0	0	0	6	52
Total 60		24	12	10	17	687	4093
% of total 24	1	<b>40</b> <sup>5</sup>	<b>20</b> <sup>5</sup>	<b>20</b> <sup>5</sup>	<b>30</b> <sup>5</sup>	<b>20</b> <sup>6</sup>	<b>15</b> <sup>7</sup>

## **TABLE 4.** (Continued)

<sup>1</sup> Mean number of species per genus for the whole Nematoda is 9, with a median of 3 and a mode of 1 (under the classification of Hodda 2022a)

<sup>2</sup> not all species formally or conventionally described

<sup>3</sup> model organism with significance for knowledge of other organisms or basic biological processes

<sup>4</sup> % of total number of described nematode genera to end 2019

<sup>5</sup>% of number of nematode genera with more than 5 species described from 2011 to 2019; note figures for f, p, i and v add to more than 100% because some species fall into more than 1 category

<sup>6</sup>% of total number of new nematode species described from 2011 to 2019

<sup>7</sup> % of total number of all nematode species described at the end of 2019

# Relationship of new descriptions with existing knowledge

The relationship between number of species described and the true species number within genera is difficult to untangle because the description of new species is heavily biased towards those genera with the most species already described. Just 2% of the total number of nematode genera together account for 20% of new descriptions (Table 4). This 2% includes 60 genera with more than 5 species described in the last 9 years, and all of the most species nematode genera are included in it, with the exception of *Helicotylenchus* (203 described species) (Table 4). The number of recently-described species in this 2% is considerably more than that expected if species descriptions were directly proportional to the number of species already described (15% of known species: Table 4). The large number of described species within these large genera could result from the effort being directed towards them, or else the substantial amount already known about them and the substantial taxonomic expertise developed in describing species. The taxonomic effort, knowledge and expertise could itself result from the genuinely substantial species diversity and the large number of species that really do exist.

The genera with most species newly recognized since 2010 also show heavy bias towards those with potential economic importance (Table 4). Of the 60 genera with more than 5 species described, only 40% were free-living, with 70% associated with plants, invertebrates or vertebrates. (Some genera are associated with more than one type of host.) The genus with the most species recognized was *Philometra*, parasites of fish. *Cucullanus*, another genus parasitic in fish, also had many species described. Plant parasites—genera *Bursaphelenchus*, *Aphelenchoides*, *Xiphinema*, *Longidorus*—have also had many species described. The former 2 genera are also associated with insect vectors, as is another genus having many species described—*Steinernema*. *Steinernema* is used for biocontrol of insects. Nematode genera used as model organisms—*Caenorhabditis* and *Pristionchus*—also had many species described, although the number is small relative to the large number and diversity of free-living genera and species thought to exist (see above). In some of the genera with many recent species descriptions, the large numbers of new species have been associated with only one or a few studies of limited geographic range (e.g. Siddiqi 2015).

Effort in discovering and describing vertebrate parasite species is not related to the diversity of potential hosts (Jorge & Poulin 2018).

## Nature of species descriptions

Another notable trend has been recognition of species without traditional full descriptions (Bouket 2014, Dey et al. 2012, Felix et al. 2014, Kiontke et al. 2011). These studies presented molecular, cross-breeding, morphological and biogeographic studies providing strong evidence of reproductive isolation, infertile progeny, character divergence These are basic criteria defining separate species under the molecular, biological, and geographic isolation. phenotypic and ecological species concepts (Coyne et al. 1988, Mayden 1997, Mayr & Ashlock 1991, Wilkins 2003, 2009, 2011). Hence there can be little doubt that these are real evolutionary entities. However, there have been considerable lags between identification of many of these species and their full description and diagnosis. Many have yet to be produced and presented. (Such time lags are not restricted to nematodes (Bebber et al. 2010, Fontaine et al. 2012)). Perhaps we have reached a point where to make tangible gains in recognizing and documenting any significant portion of the vast global nematode diversity with forseeable resources, then more flexible methods of documentation and incomplete data will be all that is possible (De Queiroz 2007, Ereschefsky 2001, Lorenzen 1983, 1994, 1996, 2000, Pleijel 1999, Renner 2016). New criteria for species description, such as those proposed for recently proposed species of *Caenorhabditis* (Felix et al. 2014), may be required. Interim designations of sibling species which are distinct genetically, biogeographically and in host range, but are indistinguishable morphologically have used identifiers such as "Caenorhabditis sp. 5" in numerous publications for 10 years before formal-albeit non-conventional-description and naming (Felix et al. 2014, Huang et al. 2014, Stevens et al. 2019). Similar designations were used for another 15 species of *Caenorhabditis* for a considerable time before formal descriptions (Felix et al. 2014, Slos et al. 2017). Very recent studies of the model nematode genus Pristionchus have resulted in some rapid descriptions of many species based on morphology, morphometrics and genome-wide sequence analysis (Kanzaki et al. 2018). This practice is continuing, for example, Rhabdias sp.4 and Rhabdias sp.5 (Mueller et al.

2018), or *Anisakis typica* species complex sp.T (Eamsobhana *et al.* 2018). It may even be taking more indefinite forms, such as *Contracaecum* type II Genotype B or *Hysterothylacum* type IV genotype B & C (Shamsi *et al.* 2018) or *Cercopithifilaria* sp. II (Maia *et al.* 2017, Otranto *et al.* 2013a, b). There have even been synonymizations of informal names (for exmple for *Hysterothylacum* larval types I to XIV: Pantoja *et al.* 2016).

Some recent descriptions of *Caenorhabditis* species have, however, taken a more conventional approach (Ferrari et al. 2017, Slos et al. 2017).

Evolutionary relationships and species concepts have been studied intensively in the genus Caenorhabditis because it is a model organism. Discovery of cryptic species has been expected, with concomitant conceptual and logistic difficulties in providing the diagnosis or description required for the name to become available (Mueller et al. 2018, Palomares-Rius et al. 2014, Perez-Ponce De Leon & Poulin 2018). But the long-term use of informal identifiers is not limited to model nematodes. "Contracaecum rudolphii A"; also "B", "C", "D", "E" and "F" have been used widely for a long time (D'Amelio et al. 2012, Li et al. 2005, Mattiucci et al. 2002, Zhang et al. 2009). A similar situation has occurred with Contracaecum osculatum with at least 5 sibling species being referred to using single letter identifiers (D'Amelio et al. 1995, Nascetti et al. 1993, Orecchia et al. 1994). These identifiers are still being used currently 20 years later (eg Karpiej et al. 2014, Shamsi 2014, Skrzypczak et al. 2014). Anisakis simplex sp "A", "B" and "C" likewise have been used for many years before formal description of one of them (Mattiucci et al. 2014). Recently, Cercopithifilaria sp. I and C. sp. II have been proposed and used in multiple publications over several years (e.g. Otranto et al. 2011, 2012, Maia et al. 2017). Likewise in Ditylenchus dipsaci (Kuehn 1857) Filipjev 1934, of 6 new species identified in 2005, only 2 have been given formal names to date (Chizhov et al. 2010, Vovlas et al. 2011). Two females described in 1973 (Zullini 1973) were given formal names in 2019 (Pena-Santiago 2019). Informal identifiers have also been necessary when new undescribed species are known or suspected and need to be referred to before subsequent formal publication: when many new species are found or suspected, not all of them can necessarily be described in a single publication (eg Davies et al. 2010a, 2015).

Informal identifiers have also been used where material is sufficient for species diagnosis and description but insufficient for definitive diagnosis or placement in a genus (eg Nippostrongylinae sp. 1, 2, 3: Smales & Heinrich 2010, Smales 2014, or *Halomonhystera* sp. A and B: Leduc 2014, or *Fissicauda* sp.1,2: Durette-Desset *et al.* 2017). This may be quite frequent. Nematodes in many habitats display high spatial and temporal patchiness which makes obtaining enough specimens often problematic and impractical when operating with limited time and resources (Hodda 1990, Hodda & Traunspurger 2021, Hodda & Khudhir 2022). Except for a relatively few species, repeated sampling at the same place may infrequently locate the same species (Hodda 1981, Hodda & Khudhir 2022, Khudhir *et al.* 2022, Nicholas 2001, Nicholas & Hodda 1999). Likewise, sampling at adjacent places frequently does not result in re-collection of more than a few species in many habitats (Bernard 1992, Fonseca *et al.* 2010b, Gingold *et al.* 2010, Hodda 1990, Traunspurger & Michiels 2006, Venekey *et al.* 2010). Without a sufficient number of specimens, assessing the variation within putative new taxa and how it relates to other nematode taxa is often not possible.

## What do nematode species tell us about evolution and species concepts?

There have been suggestions that among small organisms there are few true species. One suggestion is that small organisms are superabundant, can disperse with minimal energy cost suspended in the air or water or attached to drifting materials or other species, and can survive until they find suitable conditions (Costello & Wilson 2011, Costello *et al.* 2012, 2013, Finlay *et al.* 2001, Finlay & Fenchel 2004). This means there is constant mixing and few, widespread species. Nematodes fulfil all these criteria, being superabundant (Hodda & Davies 2019, Hoogen *et al.* 2019, Jairajpuri & Ahmad 1992, Platt 1994), able to disperse by wind, water or by using a vector organism (Banks *et al.* 2018, Fontaneto 2019, Kaya & Gaugler 1993, Kruitbos *et al.* 2009, Norton & Niblack 1991, Pena *et al.* 2011, Ptatscheck *et al.* 2018), and being able to survive for long periods if required (Gibbs 1982, Perry 1999, Wharton 1999, 2002). Although many nematodes are able to disperse by wind, water or biological vectors, it may be that many species do not (Hodda 1990).

Another suggestion is that high mutation rates and large population size mean that stable differences in fitness for different niches cannot evolve in different lineages for organisms smaller than 1mm body length, and hence species will not evolve (Rossberg *et al.* 2013). Most nematode species are less than 1mm long, meaning that conventional reproductively-isolated, genetically-distinct species should not exist under this theory.

The overwhelming evidence is contrary to both these theories. In at least some nematodes studied in detail, there are distinct groups based on molecular markers, reproductive isolation, morphological or biological differences and distribution. These are criteria used in most of the current species concepts, and they have been found in representatives of all types of nematodes: in the microbivorous free-living genera *Caenorhabditis, Micoletzkya* and *Pristionchus* (Cutter *et al.* 2019, Dey *et al.* 2012, Kanzaki *et al.* 2013, 2018, Kiontke *et al.* 2011, Ragsdale *et al.* 2013, Susoy & Herrmann 2014); in the mycetophagous plant- and insect-associated genus *Bursaphelenchus* (Abad *et al.* 1991, Beckenbach *et al.* 1999, de Guiran & Bruguier 1989, de Guiran & Ritter 1984, Fukushige 1991, Fukushige & Futai 1985, Kanzaki 2008, Lange *et al.* 2008, Mamiya 1986, Metge & Burgermeister 2008, Metge *et al.* 2008, Riga *et al.* 1992, Schauer-Blume 1990, Webster *et al.* 1990); in the obligate plant-parasitic genus *Globodera* (Chrisanfova *et al.* 2008, Fleming & Powers 1998a, b, Hoolahan *et al.* 2011, Picard & Plantard 2006, Picard *et al.* 2007, Leles *et al.* 2012, Peng *et al.* 1998, 2005). It also applies in marine environments as well as terrestrial (De Ley *et al.* 1999, Derycke *et al.* 2009, Eyualem & Blaxter 2003, Fonseca *et al.* 2008, Tandigan De Ley *et al.* 2007). Detailed studies have found large-scale diversification in some nematode groups without genetic isolation (Susoy *et al.* 2016).

Although there are at least some clear groups, in nematodes the patterns of variation found in molecular, reproductive, biological, evolutionary, morphological, ecological and biogeographic characters can be far from Some apparently similar groups prove on investigation to be clearly separated, and other apparently simple. different groups on detailed investigation prove clearly the same, with most characteristics concordant. However, many groups are differentiated only partly, with their characteristics for differentiating species either inconsistent, contradictory, unclear or unknown. For example, in the plant-parasitic genus *Radopholus*, one species (*R. similis*) occurs worldwide but with clear allopatric host races which appear to be the same genetically (Kaplan & Opperman 1997, 2000, Kaplan et al. 1996, 1997, 2000). Other morphologically indistinguishable allopatric host races do appear to differ genetically and so are regarded as different species (R. bridgei, R. daklakensis, R. duriophilus and R. arabocoffeae: Nguyen et al. 2003, Siddiqi & Hahn 1995, Trinh et al. 2004, 2012, Uribe et al. 2010). Yet other morphologically indistinguishable host races are intermediate (Turaganivalu et al. 2013). There are other biogeographic differences as well (Marin et al. 1999). Similarly, in the other plant-parasitic genera Ditylenchus and *Xiphinema*, some groups which differ in hosts or morphology are genetically indistinguishable and therefore deemed conspecific, while other groups with indistinguishable host ranges are genetically distinct and therefore deemed separate species (De Luca et al. 2013, Douda et al. 2013, Gutierrez-Gutierrez et al. 2010, 2012, He et al. 2005, Kumari & Di Cesare 2013, Leles et al. 2012, Meza et al. 2012, Pedram et al. 2012a, b, Prior et al. 2010, Sakai et al. 2012, Subbotin et al. 2011, Vovlas et al. 2011). In deep-sea nematodes, some species appear to be very localized and have little intra-specific variation in morphology or genetics, while others are very widespread and have considerable morphological and genetic variation within species (Brandt et al. 2007, De Groote et al. 2017, De Oliviera et al. 2014, 2017, Derycke et al. 2008, 2013, Lambshead 2004, Lambshead & Boucher 2003, Zeppelli et al. 2011). At least some freshwater aquatic nematodes may be similar (genus Chronogaster: Ettema et al. 2000). In a mostly free-living, but opportunistically vertebrate-parasitic species (Halicephalobus gingivalis Stefanski 1954), intraspecific morphological variability in a single population of a single species started from a single individual can be greater than what is regarded as inter-specific variation (Anderson et al 1998, Fonderie et al. 2013, Nadler et al. 2003). Patterns of variability within the genera Caenorhabditis, Pristionchus, Bursaphelenchus, Globodera, Radopholus, Ascaris and Trichuris are similarly complex (Caenorhabditis—Crombie et al. 2019, Cutter et al. 2019, Dey et al. 2012, Ferrari et al. 2017, Kanzaki et al. 2012, 2013, Kiontke et al. 2011; Pristionchus-Ragsdale et al. 2013; Bursaphelenchus-Abad et al. 1991, Beckenbach et al. 1999, de Guiran & Bruguier 1989, de Guiran & Ritter 1984, Fukushige & Futai 1985, Kanzaki 2008, Lange et al. 2008, Mamiya 1986, Metge & Burgermeister 2008, Metge et al. 2008, Riga et al. 1992, Schauer-Blume 1990, Webster et al. 1990; Globodera-Alenda et al. 2013, Blanchard 2006, Boucher et al. 2013, Handoo et al. 2012, Lax et al. 2014, Madani et al. 2010, Montary et al. 2015, 2019, Thevenoux et al. 2019; Radopholus—Elbadri et al. 1999a, b, 2002, Haegeman et al. 2010, Hahn et al. 1996, Huettel & Yaegashi 1988, Huettel et al. 1984, 1986, Plowright et al. 2013, Valette et al. 1998; Ascaris-Cavallero et al. 2013, Iniguez et al. 2012, Liu et al. 2012; Trichuris—Xie et al. 2018). Comparative studies have demonstrated that the amount of genetic variation within single interbreeding populations or biological species varies considerably across Phylum Nematoda (Gibson & Fuentes 2014). Differences in the amount of genetic variation within species may in fact be inevitable, and result from different selective evolutionary pressures on each species. Parasitic species—particularly on vertebrates—may be more variable than other groups of nematodes, but there are clearly other, as yet unidentified, relationships (Gibson & Fuentes 2014, Perez-Ponce de Leon & Poulin 2018).

As a measure of the complexity of relationships among closely-related nematodes, and the issues this can raise in defining taxonomic entities, hybridization with various success rates has been observed between well-recognized species of several nematode genera (*Globodera*—Douda *et al.* 2014, Mugniery 1979, Mugniery *et al.* 1992, Thiery *et al.* 1996, 1997; *Nacobbus*—Anthoine & Mugniery 2006; *Aphelenchoides*—Cayrol & Dalmasso 1975, Dalmasso & Cayrol 1973, *Fergusobia*—Ye *et al.* 2007, *Anisakis*—Simsek *et al.* 2020, and *Haemonchus*—Santos *et al.* 2019). There is molecular evidence suggesting that several species of both *Meloidogyne* and *Caenorhabditis* have arisen from hybridization of 2 ancestral species (Lamelza *et al.* 2020, Schoonmaker *et al.* 2020). Hybridization may be important in speciation in many organisms (Abbot *et al.* 2013, Roux *et al.* 2016, Seehausen *et al.* 2014). Notwithstanding the apparent frequency of hybridization of putative species of nematodes, genetic barriers to hybridization have been identified in several genera (e.g. *Caenorhabditis*—Bundus *et al.* 2018, Cutter 2018 and *Mesorhabditis*—Launay *et al.* 2020).

Complex patterns of cross compatibility and incompatibility in what are often considered races within species are also known in several nematode genera (*Globodera*—Franco & Evans 1978; *Nacobbus*—Anthoine & Mugniery 2006; *Pratylenchus*—Mizukubo *et al.* 2003, Perry *et al.* 1980; and *Ditylenchus*—Ladygina 1969, 1970, 1972a, b, c, 1973, 1974, 1976a,b, 1978, 1982, Ladygina & Barabashova 1976).

## Species definitions and boundaries

Defining species boundaries in nematodes is problematic because variation between individuals, populations, and reproductive, ecological or biogeographic groups is so complex (Bik *et al.* 2010, Thiery *et al.* 1997). But this situation is not limited to nematodes (Abbot *et al.* 2013, Carstens *et al.* 2013, Feder *et al.* 2012, Fujita *et al.* 2012, Lee 1998, Peccoud *et al.* 2009, Zhang *et al.* 2013). In other taxonomic groups as well the boundary between species, varieties, races and other sub-specific groups is a continuum as well (Barton 2001, Coyne & Orr 2004, Futuyma 2005, Mallet 2008, Seehausen 2004). Despite the uncertainties, the species is currently the basic unit for most studies of diversity, ecology and biology, as well as being used in biosecurity regulations and conservation planning. Hence, having a generally agreed concept of what is a species is important, even if this concept has to be a bit fuzzy. In this regard, the detailed studies on nematodes provide some of the best information available on which to base general models and concepts for other organisms (Cutter *et al.* 2019).

Although there are detailed studies of some species, nematode species descriptions based on detailed studies are not the norm (but see Felix et al. 2014, Hodda et al. 2014). The resources to investigate the details of biological, genetic, and ecological relationships of every putative species are generally not available, and are unlikely to be forthcoming any time soon. So a degree of uncertainty in the definition of most nematode species will remain, and the generally agreed concepts will need to be applied. A comprehensive cladistic phylogenetic analysis of free-living nematodes some years ago concluded that nematode classification was then an incomplete system, and was likely to remain so for some time (Lorenzen 1994). The present classifications show that the system is still incomplete for nematodes (Hodda 2022a). Even though over 28 000 valid species are recognized, and there are many more nematode species described but subsequently synonymized, this still represents only a small fraction of the estimated total number of species (Hugot et al. 2001, Hodda & Khudhir 2022). The task remains to describe as much of the huge nematode diversity as possible, even with the dual uncertainties of what is a nematode species and what is the taxonomic framework in which to place it evolutionarily. Too often nematode material is never recorded where specimens are unusual or rare, and the material never becomes available to scientists other than the collector (Khudhir et al. 2022). The tension between conservative approaches which only present taxonomic work that has a high degree of certainty as against presenting descriptions which are incomplete or uncertain (often because of a lack of material) was summarized recently by Payne (2015). Numerous examples of nematode taxa without full description and formal scientific name, but with an informal name and incomplete description testify to the issue in nematode taxonomy (eg Carta & Skantar 2014, Choudhury & Nadler 2016, Davies et al. 2015, De Oliviera et al. 2014, Kanzaki & Giblin-Davis 2014, Leduc 2014, Moravec & Barton 2015, Moravec & Jirku 2014a, Moravec & Manoharan 2014a, Moravec et al. 2016, Raymond et al. 2014, Smales 2015, Tchesunov et al. 2015, Thorne et al. 2017, Yong et al. 2015). There has also been a formal definition of what is a known species complex in the highlystudied model genus Pristionchus (Yoshida et al. 2018).

## The number of nematode species

Recognition of the difficulty in defining species has led to concerns about taxonomic inflation. These concerns have been expressed for nematodes, particularly the plant parasites (Luc *et al.* 1987, Maggenti 1991, Maggenti *et al.* 1987, 1988), as well as in some charismatic vertebrate groups like birds, amphibians and mammals. In primates, the number of species has doubled since 1985 through elevation of subspecies to species level even though few new taxa have been discovered (Burgin *et al.* 2018, Fisher *et al.* 2018, Isaac *et al.* 2004). Taxonomic inflation may be less of an issue for nematodes than for more charismatic groups because new species have been overwhelmingly based on new material rather than just revision of old material as with primates. Some new species have been identified from existing material (primarily culture collections), but most new species of nematodes have resulted from collections in new locations, habitats or hosts, as well as more intensive collecting from places sampled inadequately previously. Re-sampling of previous collecting sites, as well as collections from new sites in similar habitats and collections in different habtats are all likely to result in previously undescribed nematode species being found (Hodda 1990, Hodda & Khudhir 2022).

Notable has been the recent description of nematodes from up to 9 Km below the Earth's surface (Borgonie *et al.* 2011). At the other end of the altitude range, nematode survival has been documented in space and high in the earth's atmosphere after disintegration of a space-ship re-entering earth's atmosphere (Szewczyk *et al.* 2005). Together, these findings extend the overall habitat envelope for nematodes, and supports the claim, made many years ago, that nematodes occur everywhere on earth: "if all the matter in the universe except the nematodes were swept away, our world would still be dimly recognizable... we would find its mountains, hills valleys, rivers, lakes and oceans represented by a film of nematodes. The ghostly outlines of all the plants, animals and humans would appear outlined by their nematode parasites..." (Cobb, 1914, p 472).

## Nomenclatorial issues

The complexity of phylogenetic relationships in nematodes, together with their great diversity has also led to the suggestion of taxa above species and genus rank without taxonomic ranks, particularly in the species-rich Rhabditida (Sudhaus 2011). Such non-traditional approaches have been vigorously supported and equally vigorously disputed (Dubois *et al.* 2021, De Queiroz & Donohue 2001, 2013, Nixon & Carpenter 2000, Nixon *et al.* 2003, Platnik 2012, 2013). Higher taxa without ranks may be particularly useful in the largest nematode orders, the Spirurida, Rhabditida and Panagrolaimida. In these taxa the full range of traditional ranks has been required to reflect phylogeny, even with relatively conservative approaches assuming polytomies where evidence remains inconclusive or inconsistent (Hodda 2007, 2011, Hodda 2022a), but as the number of taxa known increases and the resolution improves, then the current number of ranks may prove inadequate. This does not seem to be necessary at the moment because, since previous synopses, there have been few studies or changes suggested to the composition of families, but these are relatively few. In few cases is there unequivocal, consistent genetic, morphological and other evidence to change families or resolve polytomies at higher levels of nematode classification. The concentration of new species descriptions in groups of economic, ecological or biological significance, rather than phylogenetic importance has not assisted resolving issues, notwithstanding initiatives to address this (Thomas 2007).

# Conclusion

A corollary of the large number of new descriptions, uncertainty over exactly what does constitute a nematode species, and uncertainties in higher-level classification mean that revisions and synonymization will continue to be as necessary as new descriptions. Organization of the species and genera into classifications of higher taxa will increasingly be necessary to keep track of, and enable access to, nematode diversity. Continual refinement of these nematode classifications will need to continue if we are to reflect more and more accurately the great diversity and complexity of relationships among nematodes. In all this uncertainty, one thing is certain: nematode classification will continue to challenge science, as well as the International Code of Zoological Nomenclature and taxonomic system, with its intricacies. Despite all this, progress has been made, and continues to be made.

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## References

- Abad, P., Tares, S., Burgier, N. & de Guiran, G. (1991) Characterization of the relationships in the pinewood nematode species complex (PWNSC) (*Bursaphelenchus* spp.) using a heterologous unc-22 DNA probe from *Caenorhabditis elegans*. *Parasitology*, 102, 303–308.
- Abbott, R., Albach, D., Ansell, S., Arntzen, J.W., Baird, S.J.E., Bierne, N. & Zinner, D. (2013) Hybridization and speciation. *Journal of Evolutionary Biology*, 26, 229–246.
- Alenda, C., Gallot-Legrand, A., Fouville, D. & Grenier, E. (2013). Sequence polymorphism of nematode effectors highlights molecular differences among the subspecies of the tobacco cyst nematode complex. *Physiological and Molecular Plant Pathology*, 84, 107–114.
- Allen, M.W. & Sher, S.A. (1967) Taxonomic problems concerning the phytoparasitic nematodes. Annual Review of Phytopathology, 5, 247–262.
- Anderson, R.C., Linder, K.E. & Peregrine, A.S. (1998) *Halicephalobus gingivalis* (Stefanski, 1954) from a fatal infection in a horse in Ontario, Canada with comments on the validity of *H. deletrix* and a review of the genus. *Parasite*, 5, 255–261.
- Anderson, T.J.C. (1995) *Ascaris* infections in humans from North America: molecular evidence for cross-infection. *Parasitology*, 110, 215–219.
- Anderson, T.J.C. (2001) The dangers of using single locus markers in parasite epidemiology: *Ascaris* as a case study. *Trends in Parasitology*, 17, 183–188.
- Anderson, T.J., Romero-Abal, M.E. & Jaenike, J. (1993) Genetic structure and epidemiology of Ascaris populations: patterns of host affiliation in Guatemala. Parasitology, 107, 319–334.
- Anderson, T.J.C., Romero-Abal, M.E. & Jaenike, J. (1995) Mitochondrial DNA and *Ascaris* epidemiology: the composition of parasite populations from individual hosts, families and villages. *Parasitology*, 110, 221–229.
- Anthoine, G. & Mugniery, D. (2006) Crossing experiments with South American populations of *Nacobbus aberrans* (Thorne, 1935) Thorne and Allen, 1944 (Nematoda : Pratylenchidae). *Nematropica*, 36, 67–77.
- Banks, N.C., Paini, D.R., Bayliss, K.L. & Hodda, M. (2015) The role of global trade and transport network topology in the human-mediated dispersal of alien species. *Ecology Letters*, 18, 188–199.
- Banks, N.C., Tangchitsomkid, N., Chanmalee, T., Sangsawang, T., Songvilay, P., Phannamvong, N., Thamakhot, S., Paini, D.R., Bayliss, K.L. & Hodda, M. (2018) Nematodes network too: diversity, abundance and dispersal via plant produce trade networks. *Plant Pathology*, 67, 1636–1644.
- Bartholomaeus, F., Davies, K.A., Ye, W. & Giblin-Davis, R.M. (2012) *Schistonchus* (Aphelenchoididae) from *Ficus benjamina* in Australia, with description of *S. benjamina* sp n. *Nematology*, 14, 1005–1013.
- Barton, N.H. (2001) The role of hybridization in evolution. *Molecular Ecology*, 10, 551–568.
- Bastian, H.C. (1865) Monograph on the Anguillulidae, or Free Nematoids, marine, land, and freshwater, with descriptions of 100 new species. *Transactions of the Linnean Society*, 25, 73–180.
- Bebber, D.P., Carine, M.A., Wood, J.R.I., Wortley, A.H., Harris, D.J., Prance, G.T., Davidse, G., Paige, J., Pennington, T.D., Robson, N.K.B. & Scotland, R.W. (2010) Herbaria are a major frontier for species discovery. *Proceedings of the National Academy of Science of the U.S.A.*, 107, 22169–22171.
- Beccaloni, G. & Eggleton, P (2013) Order Blattodea. Zootaxa, 3703, 46-48.
- Beckenbach, K., Blaxter, M. & Webster, J.M. (1999) Phylogeny of *Bursaphelenchus* species derived from analysis of ribosomal internal transcribed spacer DNA sequences. *Nematology*, 1, 539–548.
- Bernard, E.C. (1992) Soil nematode biodiversity. Biology & Fertility of Soils, 14, 99-103.
- Bernard, E.C. & Whittington, A.E. (2021) Papers and new species of minor insect orders published in *Zootaxa*, 2001–2020. *Zootaxa*, 4979 (1), 232–235.

https://doi.org/10.11646/zootaxa.4979.1.26

- Bik, H.M., Lambshead, P.J.D., Thomas, W.K. & Lunt, D.H. (2010) Moving towards a complete molecular framework of the Nematoda: a focus on the Enoplida and early-branching clades. *BMC Evolutionary Biology*, 10, 353.
- Blanchard, A. (2006) *Identification, polymorphism and molecular evolution of pathogenicity genes in the potato cyst nematode* Globodera pallida. Thesis, Université Rennes, Rennes.
- Bock, P.E. & Gordon, D.P. (2013) Phylum Bryozoa Ehrenberg, 1831. Zootaxa, 3703, 67–74. https:// doi.org/10.11646/zootaxa.3703.1.14
- Borgonie, G., Garcia-Moyano, A., Litthauer, D., Bert, W., Bester, A,., van Heerden, E., Moller, C., Erasmus M. & Onstott, T.C.

(2011) Nematoda from the terrestrial deep subsurface of South Africa. Nature, 474, 79–82.

- Boucher, A.C., Mimee, B., Montarry, J., Bardou-Valette, S., Belair, G., Moffett, P. & Grenier, E. (2013) Genetic diversity of the golden potato cyst nematode *Globodera rostochiensis* and determination of the origin of populations in Quebec, Canada. *Molecular Phylogenetics and Evolution*, 69, 75–82.
- Bouket, A.C. (2014) Hierarchical cluster analysis of *Criconemoides* species (Nematoda: Criconematidae), with a proposal for unknown species identification. *Archives of Phytopathology and Plant Protection*, 47, 90–105.
- Brandt, A., De Broyer, C., De Mesel, I., Ellingsen, K.E., Gooday, A.J., Hilbig, B., Linse, K., Thomson, M.R.A. & Tyler, P.A. (2007) The biodiversity of the deep Southern Ocean benthos. *Philosophical Transactions Of The Royal Society Series B*, 362, 39–66.
- Bundus, J.D., Wang, D. & Cutter, A.D. (2018) Genetic basis to hybrid inviability is more complex than hybrid male sterility in *Caenorhabditis* nematodes. *Heredity*, 121, 169–182.
- Burgin, C.J., Colella, J.P., Kahn, P.L. & Upham, N.S. (2018) How many species of mammals are there? *Journal of Mammalogy*, 99, 1–14.
- Carstens, B.C., Pelletier, T.A., Reid, N.M. & Satler, J.D. (2013) How to fail at sepecies delimitation. *Molecular Ecology*, 22, 4369–4383.
- Carta, L.K. & Skantar, A.M. (2014) A *Trichodorus* (Triplonchida: Trichodoridae) nematode from thrips (Thysanoptera: Panchaetothripinae). *Journal of Nematology*, 46, 302–308.
- Cavallero, S., Snabel, V., Pacella, F., Perrone, V. & D'Amelio, S. (2013) Phylogeographical studies of *Ascaris* spp. based on ribosomal and mitochondrial DNA sequences. *PLoS Neglected Tropical Diseases*, 7, e2170.
- Cayrol, J.-C. & Dalmasso, A. (1975) Interspecific affinities among 3 leaf nematodes *Aphelenchoides fragariae*, *Aphelenchoides ritzemabosi* and *Aphelenchoides besseyi*. *Cahiers O.R.S.T.O.M. Serie Biologie*, 10, 215–225.
- Chapman, A.D. (2009) *Numbers of living species in Australia and the world*. Second edition. Australian Biodiversity Information Services, Toowoomba.
- Chizhov, V.N., Borisov, B.A. & Subbotin, S.A. (2010) A new stem nematode, *Ditylenchus weischeri* sp. n. (Nematoda: Tylenchida), a parasite of *Cirsium arvense* (L.) Scop. in the central region of the non-chernozem zone of Russia. *Russian Journal of Nematology*, 18, 95–102.
- Choudhury, A. & Nadler, S.A. (2016) Phylogenetic relationships of Cucullanidae (Nematoda), with observations on Seuratoidea and the monophyly of *Cucullanus*, *Dichelyne* and *Truttaedacnitis*. *Journal of Parasitology*, 102, 87–93.
- Chrisanfova, G.G., Charchevnikov, D.A., Popov, I.O. & Zinovieva, S.V. (2008) Genetic variability and differentiation of three Russian populations of potato cyst nematode *Globodera rostochiensis* as revealed by nuclear markers. *Russian Journal of Genetics*, 44, 533–538.
- Cobb, N.A. (1914) Nematodes and their relationships. In: Anon (ed) Yearbook United States Department of Agriculture. United States Department of Agriculture, Beltsville MD, pp. 457–90.
- Cobb, N.A. (1920) One hundred new Nemas (type-species of 100 new genera). *In*: Cobb, N.A. (ed) *Contributions to a science of Nematology, IX*, Waverley Press, Baltimore, pp 217–343.
- Cobb, N.A. (1932) The english word "nema". Journal of the American Medical Association, 98, 75.
- Coddington, J.A. & Colwell, R.K. (2001) Arachnids. In: Levin, S.A. (ed) *Encyclopedia of Biodiversity*, Academic Press, Cambridge MS, pp 199–218.
- Cordeiro-Estrela, P., Feijo, A., Gaubert, P., Weksler, M., Hautier, L., Velazco, P.M., Teta, P., Fabre, P.-H., Veron, G. & Braun, J.K. (2021) The role and impact of Zootaxa in mammalogy in its first 20 years. *Zootaxa*, 4979 (1), 70–94. https://doi.org/10.11646/zootaxa.4979.1.10
- Costello, M.J., May, R.M., Stork, N.E. (2013) Response to Comments on "Can We Name Earth's Species Before They Go Extinct?". *Science*, 341, 237.
- Costello, M.J., Wilson, S.P. (2011) Predicting the number of known and unknown species in European seas using rates of description. *Global Ecology & Biogeography*, 20, 319–330.
- Costello, M.J., Wilson, S.P., Houlding, B. (2012) Predicting total global species richness using rates of species description and estimates of taxonomic effort. Systematic Biology, 61, 871–883.
- Coyne, D.L., Cortada, L., Dalzell, J.J., Claudius-Cole, A.O., Haukeland, S., Luambano, N. & Talwana, H. (2018) Plant-Parasitic Nematodes and Food Security in Sub-Saharan Africa. *Annual Review of Phytopathology*, 56, 381–403.
- Coyne, J.A. & Orr, H.A. (2004) Speciation. Sinauer, Sunderland MA, 545 pp.
- Coyne, J.A., Orr, H.A. & Futuyma, D.J. (1988) Do we need a new species concept? Systematic Zoology, 37, 190-200.
- Criscione, C.D., Anderson, J.D., Sudimack, D., Peng, W., Jha, B., Williams-Blangero, S. & Anderson, T.J.C. (2007) Disentangling hybridization and host colonization in parasitic roundworms of humans and pigs. *Proceedings of the Royal Society Series B*, 274, 2669–2677.
- Crombie, T.A., Zdraljevic, S., Cook, D.E., Tanny, R.E., Brady, S.C., Wang, Y., Evans, K.S., Hahnel, S., Lee, D., Rodriguez, B.C., Zhang, G.T., van der Zwagg, J., Kiontke, K. & Andersen, E.C. (2019) Deep sampling of *Caenorhabditis elegans* reveals high genetic diversity and admixture with global populations. *ELIFE*, 8, e50465.
- Cutter, A.D. (2018) X exceptionalism in Caenorhabditis speciation. Molecular Ecology, 27, 3925-3934.
- Cutter, A.D. (2019) Reproductive transitions in plants and animals: selfing syndrome, sexual selection and speciation. *New Phytologist*, 224, 1080–1094.
- Cutter, A.D., Morran, L.T. & Phillips, P.C. (2019) Males, Outcrossing, and Sexual Selection in Caenorhabditis Nematodes.

Genetics, 213, 27-57.

- D'Amelio, S., Mattiucci, S. Paggi, L., Koi, M., Podvyaznaya, I., Pugachev, O., Rusinek, O., Timoshkin, O. & Nascetti, G. (1995) Taxonomic rank and origin of *Contracaecum baicalensis* Mozgovoy & Ryjikov 1950, parasite of *Phoca sibirica* from Lake Baikal, with data on its occurrence in fish hosts. *In:* Anon.. (ed) *Proceedings of the IVth international symposium* of fish parasitology. IVth international symposium of fish parasitology, Munich, pp. 27.
- D'Amelio, S., Cavallero, S., Dronen, N.O., Barros, N.B. & Paggi, L. (2012) Two new species of *Contracaecum* Railliet & Henry 1912 (Nematoda: Anisakidae), *C. fagerholmi* n. sp. and *C. rudolphii* n. sp. from the brown pelican *Pelecanus occidentalis* in the norther Gulf of Mexico. *Systematic Parasitology*, 81, 1–16.
- Dalmasso, A. & Cayrol, J.C, (1973) Karyotypic discordances between 2 hybridizable nematodes *Aphelenchoides fragariae* and *A. ritzemabosi. Comptes Rendus Hebdomadaires des Seances de l'Academie des Sciences Serie D*, 276, 3171–3173.
- Davies, K.A., Bartholomaeus, F., Giblin-Davis, R.M., Ye, W., Taylor, G.S. & Thomas, W.K. (2014c) Nematodes from galls on Myrtaceae. VIII. *Fergusobia* from small galls on shoot buds, with descriptions of four new species. *Zootaxa*, 3857, 1–40.
- Davies, K.A., Bartholomaeus, F., Kanzaki, N. & Giblin-Davis, R.M. (2013a) *Schistonchus molochi* sp n. and *S. athertonensis* sp n. (Nematoda: Aphelenchoididae) from *Ficus watkinsiana* (Moraceae) in Australia. *Nematology*, 15, 389–400.
- Davies, KA., Bartholomaeus, F., Kanzaki, N., Ye, W. & Giblin-Davis, R.M. (2013b) Three new species of *Schistonchus* (Aphelenchoididae) from the *Ficus* subgenus *Sycomorus* (Moraceae) in northern Australia. *Nematology*, 15, 347–362.
- Davies, K.A., Giblin-Davis, R.M., Ye, W., Taylor, G.S. & Thomas, W.K. (2010b) Nematodes from galls on Myrtaceae. I. *Fergusobia/Fergusonina* galls on *Corymbia* spp., with re-description of *F. magna* and notes on its phylogenetic relationships. *Zootaxa*, 2634, 25–40.
- Davies, K.A., Giblin-Davis, R.M., Ye, W., Taylor, G.S. & Thomas, W.K. (2012a) Nematodes from galls on Myrtaceae. II. *Fergusobia/Fergusonina* from small axillary bud ('stem') and leaf ('pea') galls in Australia, with descriptions of two new species. *Zootaxa*, 3415, 1–22.
- Davies, K.A., Giblin-Davis, R.M., Ye, W., Taylor, G.S. & Thomas, W.K. (2012b) Nematodes from galls on Myrtaceae. III. *Fergusobia* from flower bud and stigma galls on *Eucalyptus*, with descriptions of four new species. *Zootaxa*, 3532, 1–36.
- Davies, K.A., Ye, W., Giblin-Davis, R.M., Taylor, G.S., Purcell, M. & Thomas, K. (2014a) Nematodes from galls on Myrtaceae. IX. *Fergusobia rosettae* n. sp. on *Melaleuca quinquenervia* and *F. tolgaensis* n. sp. on *Syzygium luehmannii*, from Queensland. *Zootaxa*, 3889, 214–236.
- Davies, K.A., Giblin-Davis, R.M., Ye, W. Taylor, G.S., Makinson, J. & Purcell, M. (2014b) Nematodes from galls on Myrtaceae. X. *Fergusobia* from galls on narrow-leaved *Melaleuca* spp. in Australia, with descriptions of three new species. *Zootaxa*, 3889, 237–258.
- Davies, K.A., Taylor, G.S., Nelson, L.A., Yeates, D. & Giblin-Davis, R.M. (2014e) Nematodes from galls on Myrtaceae. VI. Fergusobia from galls on Angophora in Australia, with description of F. colbrani n. sp. and key. Zootaxa, 3856, 326–348.
- Davies, K.A, Ye, W., Giblin-Davis, R.M., Taylor, G.S., Scheffer, S. & Thomas, W.K. (2010a) The nematode genus *Fergusobia* (Nematoda: Neotylenchidae): molecular phylogeny, descriptions of clades and associated galls, host plants and *Fergusonina* fly larvae. *Zootaxa*, 2633, 1–66.
- Davies, K.A., Ye, W. Giblin-Davis, R.M., Taylor, G.S., Hodda, M. & Thomas, W.K. (2014d) Nematodes from galls on Myrtaceae. VII. *Fergusobia* from 'leafy' leaf bud galls in Australia, with re-description of *Fergusobia tumifaciens* (Currie 1937) Wachek 1955 and descriptions of *Fergusobia planchonianae* n. sp. and *Fergusobia viminalisae* n. sp.. *Zootaxa*, 3856, 529–554.
- Davies, K.A., Ye, W., Kanzaki, N. Bartholomaeus, F., Zeng, Y. & Giblin-Davis, R.M. (2015) A review of the taxonomy, phylogeny, distribution and co-evolution of *Schistonchus* Cobb, 1927 with proposal of *Ficophagus* n. gen. and *Martininema* n. gen. (Nematoda: Aphelenchoididae). *Nematology*, 17, 761–829.
- De Groote, A., Hauquier, F., Vanreusel, A. & Derycke, S. (2017) Population genetic structure in *Sabatieria* (Nematoda) reveals intermediary gene flow and admixture between distant cold seeps from the Mediterranean Sea. *BMC Evolutionary Biology*, 17, PMID 154.
- De Guiran, G. & Brugier, N. (1989) Hybridization and phylogeny of the pine wood nematode (*Bursaphelenchus* spp.). *Nematologica*, 35, 321-330.
- De Guiran, G. & Ritter, M. (1984) Variabilite morphologique, pathologique et biochimique chez les nematodes des pins (*Bursaphelenchus* spp.). *Colloques de l'INRA*, 26, 41–42.
- De Ley, P., Felix, M.A., Frisse, L.M., Nadler, S.A., Sternberg, P.W. & Thomas, W.K. (1999) Molecular and morphological characterisation of two reproductively isolated species with mirror-image anatomy (Nematoda: Cephalobidae). *Nematology*, 1, 591–612.
- De Luca, F., Lazarova, S., Troccoli, A., Vovlas, N. & Peneva, V. (2013) Morphological and molecular characterisation of *Xiphinema macroacanthum* Lamberti, Roca & Agostinelli, 1989 (Nematoda: Longidoridae) from olive orchards in southern Italy. *Systematic Parasitology*, 85, 157–171.
- De Oliveira, D.A S., dos Santos, G.A.P., Derycke, S., Moens, T. & Decraemer, W. (2014) Biodiversity and connectivity of marine nematodes associated with algae from two tropical beaches. *Journal of Nematology*, 46, 152–152.
- De Oliveira, D.A S., Decraemer, W., Moens, T., dos Santos, G.A.P. & Derycke, S (2017) Low genetic but high morphological variation over more than 1000Km coastline refutes omnipresence of cryptic diversity in marine nematodes. *BMC Evolutionary Biology*, 17, 71.
- de Queiroz, K. (2007) Species concepts and species delimitation. Systematic Biology, 56, 879-886.
- de Queiroz, K. & Donoghue, M. (2001) Taxing debate for taxonomists. Science, 292, 2249-2249.

- de Queiroz, K. & Donoghue, M. (2013) Phylogenetic nomenclature, hierarchical information, and testability. *Systematic Biology*, 62, 167–174.
- Derycke, S., Backeljau, T. & Moens, T. (2013) Dispersal and gene flow in free-living marine nematodes. *Frontiers in Zoology*, 10, 1.
- Derycke, S., De Ley, P., Tandingan De Ley, I., Holovachov, O., Rigaux, A. & Moens, T. (2009) Linking DNA sequences to morphology: cryptic diversity and population genetic structure in the marine nematode *Thoracostoma trachygaster* (Nematoda, Leptosomatidae). *Zoologica Scripta*, 39, 276–289.
- Derycke, S., Fonseca, G., Vierstraete, A., Vanfleteren, J., Vincx, M. & Moens, T. (2008) Disentangling taxonomy within the *Rhabditis (Pellioditis) marina* (Nematoda, Rhabditidae) species complex using molecular and morphological tools. *Zoological Journal of the Linnean Society*, 152, 1–15.
- Dey, A., Jeon, Y., Wang, G.X. & Cutter, A.D. (2012) Global population genetic structure of *Caenorhabditis remanei* reveals incipient speciation. *Genetics*, 191, 1257.
- Dikstra, K.-D.B., Bechly, G., Bybee, S.M., Dow, R.A., Dumont, H.J., Fleck, G., Garrison, R.W., Hamalainen, M., Kalkman, V.J., Karube, H., May, M.L., Orr, A.G., Paulson, D.R., Rehn, A.C., Theischinger, G., Trueman, J.W.H., Van Tol, J., Von Ellenrieder, N. & Ware, J. (2013) The classification and diversity of dragonflies and damselflies (Odonata). *Zootaxa*, 3703, 36–45.
- Djernaes, M, (2018) Biodiversity of Blattodea the Cockroaches and Termites. In: Foottit, R.G & Adler, P.H. (eds) *Insect Biodiversity: Science and Society, Volume II, First Edition.* John Wiley & Sons Ltd, New Jersey, pp. 359–387.
- Douda, O., Marek, M., Zouhar, M. & Rysanek, P. (2013) Insights into the structure and phylogeny of the 28S rRNA expansion segments D2 and D3 of the plant-infecting nematodes from the genus *Ditylenchus* (Nematoda: Anguinidae). *Phytopathologia Mediterranea*, 52, 84–97.
- Douda, O., Zouhar, M., Renco, M. & Marek, M. (2014) Molecular and morphological exploration of a mixed population of two potato-parasiting nematode species, *Globodera rostochiensis* and *G. pallida. Helminthologia*, 51, 3–6.
- Dubois, A., Ohler, A. & Pyron, A. (2021) New concepts and methods for phylogenetic taxonomy and nomenclature in zoology, exemplified by a new ranked cladonomy of recent amphibians (Lissamphibia). *Megataxa*, 5, 1–738. https://doi.org/10.11646/megataxa.5.1.1
- Durette-Desset, M.-C., Digiani, M.C., Kilani, M. & Geffard-Kuriyama, D. (2017) Critical revision of the Heligmonellidae (Nematoda: Trichostrongylida: Heligmosomidae). *Memoires du Museum national d'histoire naturelle*, 211, 1–290.
- Eamsobhana, P., Yong, H.S., Song, S.L., Tungtrongchitr, A. & Roongruangchai, K. (2018) Genetic differentiation of *Anisakis* species (Nematoda: Anisakidae) in marine fish *Priacanthus tayensis* from Gulf of Thailand. *Tropical Biomedicine*, 35, 669–677.
- Elbadri, G.A.A., De Ley, P., Waeyenberge, L., Vierstraete, A., Moens, M. & Vanfleteren, J. (2002) Intraspecific variation in *Radopholus similis* isolates assessed with restriction fragment length polymorphism and DNA sequencing of the internal transcribed spacer region of the ribosomal RNA cistron. *International Journal for Parasitology*, 32, 199–205.
- Elbadri, G.A.A., Geraert, E. & Moens, M. (1999a) Morphological differences among *Radopholus* populations (Nematoda: Tylenchida) from banana in Africa. *Journal of Nematode Morphology and Systematics*, 2, 1–16.
- Elbadri, G.A.A., Geraert, E. & Moens, M. (1999b) Morphological differences among *Radopholus similis* (Cobb, 1893) Thorne, 1949 populations. *Russian Journal of Nematology*, 7, 139–153.
- Ereshefsky, M. (2001) *The Poverty of the Linnaean Hierarchy. A Philosophical Study of Biological Taxonomy*. Cambridge University Press, Cambridge UK, 316 pp.
- Ettema, C.H., Rathbun, S.L. & Coleman, D.C. (2000) On spatiotemporal patchiness and the coexistence of five species of *Chronogaster* (Nematoda: Chronogasteridae) in a riparian wetland. *Oecologia*, 125, 444–452.
- Eyualem, A. & Blaxter, M. (2003) Comparison of biological, molecular, and morphological methods of species identification in a set of cultured *Panagrolaimus* isolates. *Journal of Nematology*, 35, 119–128.
- Feder, J.L., Egan, S.P. & Nosil, P. (2012) The genomics of speciation-with-gene-flow. Trends in Genetics, 28, 342-350.
- Felix, M.-A., Braendle, C. & Cutter, A.D. (2014) A streamlined system for species diagnosis in *Caenorhabditis* (Nematoda: Rhabditidae) with name designations for 15 distinct biological species. *PLOS One*, 9, e94723.
- Fenner, A.L., Smales, L.R. & Bull, C.M. (2011) Pharyngodon asterostoma Adamson, 1984 (Nematoda: Pharyngodonidae), a new parasite record for the endangered Slater's Skink, *Liopholis slateri* Storr, 1968 (Sauria: Scincidae), from the Northern Territory, Australia. *Transactions of the Royal Society of South Australia*, 135, 140–142.
- Ferrari, C., Salle, R., Callemeyn-Torre, N., Jovelin, R., Cutter, A.D., Braendle, C. (2017) Ephemeral-habitat colonization and neotropical species richness of *Caenorhabditis* nematodes. *BMC Ecology*, 17, 43.
- Finlay, B.J., Esteban, G.F., Clarke, K.J. & Olmo, J.L. (2001) Biodiversity of terrestrial protozoa appears homogeneous across local and global spatial scales. *Protist*, 152, 355–366.
- Finlay, B.J., Fenchel, T. (2004) Cosmopolitan metapopulations of free-living microbial eukaryotes. Protist, 155, 237-244.
- Fisher, M.A., Vinson, J.E., Gittleman, J.L. & Drake, J.M. (2018) The description and number of undiscovered mammal species. *Ecology & Evolution*, 8, 3628–3635.
- Fleming, C.C. & Powers, T.O. (1998a) Potato cyst nematodes: species, pathotypes and virulence concepts. *In:* Marks, R.J. & Brodie, B.B. (Eds.), *Potato cyst nematodes, biology, distribution and control*. CABI, Wallingford UK, pp. 51–57.
- Fleming, C.C. & Powers, T.O. (1998b) Potato cyst nematode diagnostics: morphology, differential hosts and biochemical techniques. *In:* Marks, R.J. & Brodie, B.B. (Eds.), *Potato cyst nematodes, biology, distribution and control.* CABI,

Wallingford UK, pp. 91–114.

- Fonderie, P., Steel, H., Moens, T. & Bert, W. (2013) Experimental induction of intraspecific morphometric variability in a single population of *Halicephalobus* cf. *gingivalis* may surpass total interspecific variability. *Nematology*, 15, 529–544.
- Fonseca, V.G., Carvalho, G.R., Sung, W., Johnson, H.F., Power, D.M., Neill, S.P., Packer, M., Blaxter, M.L., Lambshead, P.J.D., Thomas, W.K. & Creer, S. (2010a) Second-generation environmental sequencing unmasks marine metazoan biodiversity. *Nature Communications*, 1, 98.
- Fonseca, G., Derycke, S. & Moens, T. (2008) Integrative taxonomy in two free-living nematode species complexes. *Biological Journal of the Linnaean Society*, 94, 737–753.
- Fonseca, G., Saltwedel, T. & Vanreusel, A (2010b) Variation in nematode assemblages over multiple spatial scales and environmental conditions in Arctic deep seas. *Progress in Oceanography* 84, 174–184.
- Fontaneto, D. (2019) Long-distance passive dispersal in microscopic aquatic animals. Movement Ecology, 7, 10.
- Fontaine, B., Perrard, A. & Bouchet, P. (2012) 21 years of shelf life between discovery and description of new species. *Current Biology*, 22, R943–R944.
- Franco, J.P. & Evans, K. (1978) Mating of British and Peruvian populations of potato cyst nematodes *Globodera* spp. *Nematropica*, 8, 5–9.
- Fukushige, H. (1991) Propagation of *Bursaphelenchus xylophilus* (Nematoda: Aphelenchoididae) on fungi growing in pineshoot segments. *Applied Entomology and Zoology*, 26, 371–376.
- Fukushige, H. & Futai, K. (1985) Characteristics of egg shells and the morphology of female tail-tips of *Bursaphelenchus xylophilus*, *B. mucronatus* and some strains of related species from France. *Japanese Journal of Nematology*, 15, 49–54.
- Fujita, M.K., Leache, A.D., Burbrink, F.T., McGuire, J.A. & Moritz, C. (2012) Coalescent-based species delimitation in an integrative taxonomy. *Trends in Ecology & Evolution*, 27, 480–488.
- Futuyma, D.J. (2005) Evolution. Sinauer, Sunderland MA, 603 pp.
- Gibbs, H.C. (1982) Mechanisms of survival of nematode parasites with emphasis on hypo-biosis. *Veterinary Parasitology*, 11, 25–48.
- Giblin-Davis, R.M., Kanzaki, N., Davies, K.A., Ye, W., Zeng, Y., Center, B.J., Esquivel, A. & Powers, T.O. (2014) Ficotylus laselvae n. sp (Tylenchomorpha: Anguinidae) associated with Ficus colubrinae in Costa Rica. Nematology, 16, 1139– 1151.
- Gibson, A.K. & Fuentes, J.A. (2014) A phylogenetic test of the Red Queen Hypothesis: outcrossing and parasitism in the Nematode phylum. *Evolution*, 69, 530–540.
- Gingold, R., Mundo-Ocampo, M., Holovachov, O. & Rocha-Olivares, A. (2010) The role of habitat heterogeneity in structuring the community of intertidal free-living marine nematodes. *Marine Biology*, 157, 1741–1753.
- Gordon, D.P. (1999) Bryozoan diversity in New Zealand and Australia. In: Ponder, W. & Lunney, D. (eds) *The Other 99%. The Conservation and Biodiversity of Invertebrates*. Royal Zoological Society of New South Wales, Mosman, pp. 199–204.
- Gordon, D.P. & Bock. P.E. (2021) Phylum Bryozoa Ehrenberg, 1831 in the first twenty years of Zootaxa. Zootaxa, 4979, 236–239.

https://doi.org/10.11646/zootaxa.4979.1.27

- Grosemans, T., Morris, K., Thomas, W.K., Rigaux, A., Moens, T. & Derycke, S. (2016) Mitogenomics reveals high synteny and long evolutionary histories of sympatric cryptic nematode species. *Ecology and Evolution*, 6, 1854–1870.
- Guden, R.M., Vafeiadou, A.-M., De Meester, N., Derycke, S. & Moens, T. (2018) Living apart-together: Microhabitat differentiation of cryptic nematode species in a saltmarsh habitat. *PLoS One*, 13, e0204750.
- Gutierrez-Gutierrez, C., Cantalapiedra-Navarrete, C., Decraemer, W., Vovlas, N., Prior, T., Palomares-Rius, J.E. & Castillo, P. (2012) Phylogeny, diversity, and species delimitation in some species of the *Xiphinema americanum*-group complex (Nematoda: Longidoridae), as inferred from nuclear and mitochondrial DNA sequences and morphology. *European Journal* of *Plant Pathology*, 134, 561–597.
- Gutierrez-Gutierrez, C., Palomares-Rius, J.E., Cantalapiedra-Navarrete, C., Landa, B., Esmenjaud, D. & Castillo, P. (2010) Molecular analysis and comparative morphology to resolve a complex of cryptic *Xiphinema* species. *Zoologica Scripta*, 39, 483–498.
- Haegeman, A., Elsen, A., De Waele, D. & Gheysen, G. (2010) Emerging molecular knowledge on *Radopholus similis*, an important nematode pest of banana. *Molecular Plant Pathology*, 11, 315–323.
- Hahn, ML., Wright, D.J. & Burrows, P.R. (1996) The chromosome number in *Radopholus similis*—A diagnostic feature? *Nematologica*, 42, 382–386.
- Hammond, P.M. (1992) Species inventory. *In*: Groombridge, B. (Ed), *Global diversity, status of the earth's living resources*. Chapman & Hall, London UK, pp 17–39.
- Handoo, Z.A., Carta, L.K., Skantar, A.M. & Chitwood, D.J. (2012) Description of *Globodera ellingtonae* n. sp. (Nematoda: Heteroderidae) from Oregon. *Journal of Nematology*, 44, 40–57.
- He, Y., Subbotin, S.A; Rubtsova, T.V., Lamberti, F., Brown, D.J.F. & Moens, M. (2005) A molecular phylogenetic approach to Longidoridae (Nematoda : Dorylaimida). *Nematology*, 7, 111–124.
- Hodda, M. (1981) *Behavioural adaptations of nematodes to a high-energy sandy beach*. Honours thesis, Australian National University, Canberra.
- Hodda, M. (1990) Variation in estuarine littoral nematode populations over 3 spatial scales. *Estuarine, Coastal & Shelf Science*, 30, 325–40.

Hodda M. (2007) Phylum Nematoda. Zootaxa, 1668, 265-293.

- Hodda, M. (2011) Phylum Nematoda Cobb 1932. Zootaxa, 3148, 63-95.
- Hodda, M. (2021) Papers published in Zootaxa concerning Nematoda from 2001 to 2020. Zootaxa, 4979, 95–101. https://doi.org/10.11646/zootaxa.4979.1.11
- Hodda, M. (2022a) Phylum Nematoda: a classification, catalogue and index of valid genera, with a census of valid species. *Zootaxa*, 5114 (1), 1–289.
  - https://doi.org/10.11646/zootaxa.5114.1.1
- Hodda, M. (2022b) Phylum Nematoda: feeding habits for all valid genera using a new, universal scheme encompassing the entire phylum, with descriptions of morphological characteristics of the stoma, a key, discussion of evidence for trophic relationships. *Zootaxa*, 5114 (1), 318–451. https://doi.org/10.11646/zootaxa.5114.1.3
- Hodda, M., Berrie, L., Banks, N.C., Paini, D.R., Bayliss, K.L. (submitted) Using network principles to survey efficiently for plant pests and diseases. *Phytopathology*
- Hodda, M., Collins, S.J., Vanstone, V.A., Hartley, D., Wanjura, W. & Kehoe, M. (2014) *Pratylenchus quasitereoides* n. sp. from cereals in Western Australia. *Zootaxa*, 3866, 277–288.
- Hodda, M. & Davies, K.A. (2019) Nematodes in cropping systems. University of Adelaide & CSIRO, Adelaide, 230 pp.
- Hodda, M., Escalona, H., Vossen, A. & Manwaring, A. (2019) *National Entomology Skills Analysis*. Australian Department of Agriculture, Canberra, 44 pp.
- Hodda, M. & Khudhir, M. (2022) Species richness of marine free-living nematodes in eastern Australia, and what it tells us about nematode diversity and its collection. *Hydrobiologia*, .
- Hodda, M., Peters, L. & Traunspurger, W. (2009) Nematode biodiversity: terrestrial, marine & freshwater. *In*: Wilson, M. & Kakouli Duarte, T. (eds) *Nematodes as Environmental Indicators*. CABI, Wallingford UK, pp. 45–93.
- Hodda, M. & Traunspurger, W. (2021) Nematodes from extreme and unusual freshwater habitats. *In*: Traunspurger, W. (ed) *Ecology of Freshwater Nematodes*. CABI, Wallingford, pp. 109–150.
- Hodda, M., Van Der Schyff, G. & Welsh, L. (2017) Enhancing Diagnostic Capability for Priority Pests: Collections and Capability Audit. CSIRO, Canberra. 49 pp.
- Hoogen, J. van den, Geisen, S., Routh, D., Ferris, H., Traunspurger, W., Wardle, D.A., de Goede, R.G.M., Adams, B.J., Ahmad, W., Andriuzzi, W.S., Bardgett, R.D., Bonkowski, M., Campos-Herrera, R., Cares, J.E., Caruso, T., de Brito Caixeta, L., Chen, X., Costa, S.R., Creamer, R., Mauro da Cunha Castro, J., Dam, M., Djigal, D., Escuer, M., Griffiths, B.S., Gutiérrez, C., Hohberg, K., Kalinkina, D., Kardol, P., Kergunteuil, A., Korthals, G., Krashevska, V., Kudrin, A.A., Li, Q., Liang, W., Magilton, M., Marais, M., Martín, J.A.R., Matveeva, E., Mayad, E.H., Mulder, C., Mullin, P., Neilson, R., Nguyen, T.A.D., Nielsen, U.N., Okada, H., Rius, J.E.P., Pan, K., Peneva, V., Pellissier, L., Carlos Pereira da Silva, J., Pitteloud, C., Powers, T.O., Powers, K., Quist, C.W., Rasmann, S., Moreno, S.S., Scheu, S., Setälä, H., Sushchuk, A., Tiunov, A.V., Trap, J., van der Putten, W., Vestergård, M., Villenave, C., Waeyenberge, L., Wall, D.H., Wilschut, R., Wright, D.G., Yang, J.-I. & Crowther, T.W. (2019) Soil nematode abundance and functional group composition at a global scale. *Nature*, 572, 194–198.
- Hoolahan, A.H., Blok, V.C., Gibson, T. & Dowton, M. (2011) Paternal leakage of mitochondrial DNA in experimental crosses of populations of the potato cyst nematode *Globodera pallida*. *Genetica*, 139, 1509–1519.
- Hooper, J.N.A., Woerheide, G., Hajdu, E., Erpenbeck, D., De Voogd, N.J. & Klautau, M. (2021) Zootaxa 20 years: Phylum Porifera. *Zootaxa*, 4979 (1), 38–56.

https://doi.org/10.11646/zootaxa.4979.1.8

- Howie, B. (2012) Plant Pathology and Entomology Capability Study 2012. Plant Biosecurity CRC, Canberra, 16 pp.
- Huang, R.-E., Ren, X., Qiu, Y. & Zhao, Z. (2014) Description of *Caenorhabditis sinica* sp n. (Nematoda: Rhabditidae), a nematode species used in comparative biology for *C.elegans*. *PLOS One*, 9, e110957.
- Huettel, R.N., Dickson, D.W. & Kaplan, D.T. (1984) *Radopholus citrophilus* sp. n. (Nematoda), a sibling species of *Radopholus similis*. *Proceedings of the Helminthological Society of Washington*, 51, 32–35.
- Huettel, R.N., Kaplan, D.T. & Dickson, D.W. (1986) Characterization of a new burrowing nematode population, *Radopholus citrophilus*, from Hawaii. *Journal of Nematology*, 18, 50–54.
- Huettel, R.N. & Yaegashi, T. (1988) Morphological differences between *Radopholus citrophilus* and *Radopholus similis*. Journal of Nematology, 20, 150–157.
- Hugot, J.P., Baujard, P. & Morand, S. (2001) Biodiversity in helminths and nematodes as a field of study: an overview. *Nematology*, 3, 199–208.
- Hyman, L.H. (1951) The Invertebrates. III. Acanthocephala, Aschelminthes and Entoprocta. The Pseudocoelomate Bilateria. McGraw-Hill, New York, 572 pp.
- Isaac, N.J.B., Mallet, J. & Mace, G.M. (2004) Taxonomic inflation: its influence on macroecology and conservation. *Trends in Ecology & Evolution*, 19, 464–469.
- Iniguez, A.M., Leles, D., Jaeger. L.H., Carvalho-Costa, F.A. & Araújo, A. (2012) Genetic characterisation and molecular epidemiology of Ascaris spp. from humans and pigs in Brazil. Transactions of the Royal Society for Tropical Medicine & Hygiene, 106, 604–612.
- Jacobus, L.M., Salles, F.F., Price, B., Pereira-Da-Conceicoa, L., Dominguez, E., Suter, P.J., Molineri, C., Tiunova, T.M. & Sartori, M. (2021) Mayfly taxonomy (Arthropoda: Hexapoda: Ephemeroptera) during the first two decades of the twenty-

first century and the concentration of taxonomic publishing. *Zootaxa*, 4979 (1), 25–30. https://doi.org/10.11646/zootaxa.4979.1.6

- Jaeger, P., Arnedo, M.A., Azevedo, G.M.H.F., Baehr, B., Bonaldo, A.B., Haddad, C.R., Harms, D., Hormiga, G., Labarque, F.M., Muster, C., Ramírez, M.J. & Santos, A.O J. (2021) Twenty years, eight legs, one concept: describing spider biodiversity in Zootaxa (Arachnida: Araneae). *Zootaxa*, 4979 (1), 131–146. https://doi.org/10.11646/zootaxa.4979.1.14
- Jairajpuri, M.S. & Ahmad, W. (1992) *Dorylaimida Free-living, predaceous and plant-parasitic nematodes*. Oxford and IBH Publishing Co., Delhi, 458 pp.
- Jorge, F. & Poulin, R. (2018) Poor geographical match between the distributions of host diversity and parasite discovery effort. *Proceedings of the Royal Society B-Biological Sciences*, 285, 20180072.
- Justine, J.-L. & Iwaki, T. (2014) *Huffmanela hamo* sp n. (Nematoda: Trichosomoididae: Huffmanelinae) from the dagger-tooth pike conger *Muraenesox cinereus* off Japan. *Folia Parasitologica*, 61, 267–271.
- Kanzaki, N. (2008) Taxonomy and systematics of the nematode genus *Bursaphelenchus* (Nematoda: Parasitaphelenchidae). *In:* Zhao, BG, Futai K, Sutherland, J.R. & Takeuchi Y (Eds.), *Pine Wilt Disease*. Springer, Tokyo, pp. 44–66.
- Kanzaki, N. & Giblin-Davis, R.M. (2014) Phylogenetic status and morphological characters of *Rhabditolaimus anoplophorae* (Rhabditida: Diplogastridae). *Journal of Nematology*, 46, 44–49.
- Kanzaki, N., Herrmann, M., Yoshida, K., Weiler, C., Roedelsperger, C. & Sommer, R.J. (2018) Samplings of millipedes in Japan and scarab beetles in Hong Kong result in five new species of *Pristionchus* (Nematoda: Diplogastridae). *Journal of Nematology*, 50, 587–610.
- Kanzaki, N., Ragsdale, E.J., Herrmann, M., Mayer, W.E. & Sommer, R.J. (2012) Description of three *Pristionchus* species (Nematoda: Diplogastridae) from Japan that form a cryptic species complex with the model organism *P. pacificus*. *Zoological Science*, 29, 403–417.
- Kanzaki, N., Ragsdale, E.J., Herrmann, M., Roseler, W. & Sommer, R.J.(2013) Two new species of *Pristionchus* (Nematoda: Diplogastridae) support the biogeographic importance of Japan for the evolution of the genus *Pristionchus* and the model system *P. pacificus*. *Zoological Science*, 30, 680–692.
- Kanzaki, N., Tanaka, R., Giblin-Davis, R.M. & Davies, K.A. (2014) New plant-parasitic nematode from the mostly mycophagous genus *Bursaphelenchus* discovered inside figs in Japan. *PLOS one*, 9, e99241.
- Kaplan, D.T. & Opperman, C.H. (1997) Genome similarity implies that citrus-parasitic burrowing nematodes do not represent a unique species. *Journal of Nematology*, 29, 430–440.
- Kaplan, D.T. & Opperman, C.H. (2000) Reproductive strategies and karyotype of the burrowing nematode, *Radopholus similis*. *Journal of Nematology*, 32, 126–133.
- Kaplan, D.T., Thomas, W.K., Frisse, L.M., Sarah, J.L., Stanton, J.M., Speijer, P.R., Marin, D.H. & Opperman, C.H. (2000) Phylogenetic analysis of geographically diverse *Radopholus similis* via rDNA sequence reveals a monomorphic motif. *Journal of Nematology*, 32, 134–142.
- Kaplan, D.T., Vanderspool, M.C., Garrett, C., Chang, S. & Opperman, C.H. (1996) Molecular polymorphisms associated with host range in the highly conserved genomes of burrowing nematodes, *Radopholus* spp. *Molecular Plant-Microbe Interactions*, 9, 32–38.
- Kaplan, D.T., Vanderspool, M.C. & Opperman, C.H. (1997) Sequence tag site and host range assays demonstrate that *Radopholus similis* and *R. citrophilus* are not reproductively isolated. *Journal of Nematology*, 29, 421–429.
- Karpiej, K., Simard, M., Pufall, E. & Rokicki, J. (2014) Anisakids (Nematoda: Anisakidae) from ringed seal, *Pusa hispida*, and bearded seal, *Erignathus barbatus* (Mammalia: Pinnipedia) from Nunavut region. *Journal of the Marine Biological Association of the United Kingdom*, 94, 1237–1241.
- Kaya, H.K & Gaugler, R. (1993) Entomopathogenic nematodes. Annual Review of Entomology, 38, 181-206.
- Khudhir, M., Nicholas, E., Campbell, J., Nicholas, W.L. & Hodda, M. (2022) A catalogue of the nematode slide collection from the late W.L. Nicholas. *Zootaxa*, in press.
- Kiontke, K.C., Felix, M.A., Ailion, M., Rockman, M.V., Braendle, C., Penigault, J.B. & Fitch, D.H.A. (2011) A phylogeny and molecular barcodes for *Caenorhabditis*, with numerous new species from rotting fruits. *BMC Evolutionary Biology*, 11, 339.
- Kruitbos, L.M., Heritage, S. & Wilson, M.J. (2009) Phoretic dispersal of entomopathogenic nematodes by *Hylobius abietis*. *Nematology*, 11, 419–427.
- Kumari, S. & Di Cesare, A. (2013) Nicotinamide dehydrogenase subunit 4 analysis of *Xiphinema diversicaudatum* and *Xiphinema simile* (Nematoda: Longidoridae). *European Journal of Plant Pathology*, 136, 803–810.
- Kury, A.B. (2013) Order Opiliones Sundevall, 1833. Zootaxa, 3703, 27–33.
- Ladygina, N.M. (1969) Physiological compatibility of various forms of stem nematodes. 1. Crossing of *Ditylenchus* from onion, strawberry and red clover. *Parazitologiya*, 3, 559–568.
- Ladygina, N.M. (1970) Physiological compatibility of various forms of stem eelworms. Parazitologiya, 4, 133-135.
- Ladygina, N.M. (1972a) Results of an investigation into the physiological compatibility of stem nematodes. *Nematodnye bolezni* sel'skokhozyaistvennykh kul'tur i mery bor'by s nimi. Tezisy soveshchaniya. Moskva, dekabr' 1972, 118–119.
- Ladygina, N.M. (1972b) Physiological compatibility of stem nematodes. *Problemy parazitologii. Trudy VII Nauchnoi Konferentsii Parazitologov USSR*, 1, 460–461.
- Ladygina, N.M. (1972c) Physiological compatibility of different forms of stem nematode: 2. Crossing of stem eelworms of

onion, garlic and potato. Russian Translation Program NL, 7086, 1-7.

- Ladygina, N.M. (1973) Physiological compatibility of different forms of stem eelworms. Part 3. Crossing of parsley parsnip onion and strawberry eelworms. *Parazitologiya*, 7, 67–71.
- Ladygina, N.M. (1974) Genetic physiological compatibility of various forms of stem eelworms. Part 4. The crossing of the phlox eelworm with other stem eelworms. *Parazitologiya*, 8, 63–69.
- Ladygina, N.M. (1976a) The crossing of the stem nematode *Ditylenchus destructor* Thorne 1945 from the potato with the nematode *D. dipsaci* (Kuhn 1957) Filipjev 1936 from parsley and parsnip. *Nauchnye Doklady Vysseh Shkoly Biologiceskie Nauki*, 10, 118–120.
- Ladygina, N.M. (1976b) Genetic and physiological compatibility of different forms of stem eelworms. Part 5. The crossing of the red clover race with other stem eelworms. *Parazitologiya*, 10, 40–47.
- Ladygina, N.M. (1978) The genetic and physiological compatibility of different forms of stem nematodes. VI. Crossbreeding of *Ditylenchus* from cultivated plants and from weeds. *Parazitologiya*, 12, 349–353.
- Ladygina, N.M. (1982). [Biological races, karyotypes and hybridization. *In*: 'Gubina, V.G. (ed). [*Plant and soil nematodes, genus* Ditylenchus.] Izdatel'stvo "Nau-ka", Moscow, pp 69–86.
- Ladygina, N.M. & Barabashova, V.N. (1976) Genetic and physiological compatibility and karyotypes of stem eelworms. *Parazitologiya*, 10, 449–456.
- Lambshead, P.J.D. (1993). Recent developments in marine benthic biodiversity research. *Recent developments in benthology*, 19, 5–24.
- Lambshead, P.J.D. (2004) Marine nematode biodiversity. In: Chen, Z.X., Chen, S.Y. & Dickson, D.W. (eds) Nematology: Advances And Perspectives, Vol 1: Nematode Morphology, Physiology and Ecology. CABI, Wallingford UK, pp. 438– 468.
- Lambshead, P.J.D. & Boucher, G. (2003) Marine nematode deep-sea biodiversity hyperdiverse or hype. *Journal of Biogeography*, 30, 475–485.
- Lambshead, P.J.D. & Hodda, M. (1994) Impact of disturbance on measurements of variability in marine nematode populations. *Vie et Milieu*, 44, 21–27.
- Lamelza, P., Young, J.M., Noble, L.M., Caro, L., Isakharov, A., Palanisamy, M., Rockman, M.V., Malik, H.S. & Ailion, M. (2019) Hybridization promotes asexual reproduction in *Caenorhabditis* nematodes. *PLOS Genetics*, 15, e1008520.
- Lange, C., Burgermeister, W., Metge, K. & Braasch H. (2008) Molecular characterization of isolates of the Bursaphelenchus sexdentati group using ribosomal DNA sequences and ITS-RFLP. In: Mota, M.M. & Vieira, P. (eds) Pine Wilt Disease—A Worldwide Threat to Forest Ecosystems. Gulbenkian Foundation, Lisbon, pp. 165–173.
- Launay, C., Felix, M.-A., Dieng, J. & Delattre, M. (2020) Diversification and hybrid incompatibility in auto-pseudogamous species of *Mesorhabditis* nematodes. *BMC Evolutionary Biology*, 20, 105.
- Lax, P., Duenas, J.C.R., Franco-Ponce, J., Gardenal, C.N. & Doucet, M.E. (2014) Morphology and DNA sequence data reveal the presence of *Globodera ellingtonae* in the Andean region. *Contributions to Zoology*, 83, 227–243.
- Leduc, D. (2014) Free-living nematodes of the genus *Halomonhystera* (Monhysteridae) from the Southwest Pacific region and Ross Sea. *New Zealand Journal of Zoology*, 41, 46–57.
- Lee, M.S.Y. (1998) Phylogenetic uncertainty, molecular sequences, and the definition of taxon names. *Systematic Biology*, 47, 719–726.
- Leles, D., Gardner, S.L., Reinhard, K., Iñiguez, A. & Araujo, A. (2012) Are Ascaris lumbricoides and Ascaris suum a single species? Parasites & Vectors 5, 42.
- Li, A., D'Amelio, S., Paggi, L., He, F., Gasser, R.B., Lun, Z., Abollo, E., Turchetto, M. & Zhu, X. (2005) Genetic evidence for the existence of sibling species within *Contracauecum rudolphii* (Hartwich 1964) and the variability of *Contracaecum septentrionale* (Kreis 1955) (Nematoda: Anisakidae). *Parasitology Research*, 96, 361–366.
- Linnaeus, C. (1758) Systema naturae regna tria naturae, secundum classes, ordines, genera, species, cum characteribus differentiis, synonymis, locis. 10th edition. Impensis direct. Laurentii Salvii, Holmiae, 824 pp.
- Liu, G.H., Wu, C.Y., Song, H.Q., Wei, S.J., Xu, M.J., Lin, R.Q., Zhao, G.H, Huang, S.Y. & Zhu, X.Q. (2012) Comparative analyses of the complete mitochondrial genomes of *Ascaris lumbricoides* and *Ascaris suum* from humans and pigs. *Gene*, 492, 110–116.
- Lorenzen, S. (1983) Phylogenetic systematics: problems, achievements and its application to the Nematoda. *In* : Stone, A.R., Platt, H.M. & Khalil, L.F. (eds) *Concepts in nematode systematics*. Systematics Association Special Volume 22, Academic, London UK, pp. 11–23.
- Lorenzen, S. (1994) *The phylogenetic systematics of freeliving nematodes*. Ray Society Publication 162, Ray Society, London, 383 pp.
- Lorenzen, S. (1996) The metamorphosis of traditional into advanced phylogenetic systematics and its impact on nematode systematics. *Russian Journal of Nematology*, 4, 61–70.
- Lorenzen, S. (2000) The role of the biogenetic convergence rule in polarizing transformation series—Arguments from nematology, chaos science, and phylogenetic systematics. *Annales Zoologici*, 50, 267–275.
- Luc, M., Maggenti, A.R., Fortuner, R., Raski D. & Geraert, E. (1987) A reappraisal of Tylenchina (Nemata) 1. For a new approach to the taxonomy of Tylenchina. *Revue de Nematologie*, 10, 127–134.
- Madani, M., Subbotin, S.A., Ward, L.J., Li, X. & De Boer, S.H. (2010) Molecular characterization of Canadian populations of potato cyst nematodes, *Globodera rostochiensis* and *G. pallida* using ribosomal nuclear RNA and cytochrome b genes.

Canadian Journal of Plant Pathology, 32, 252-263.

- Magalhaes, W.F., Hutchings, P., Oceguera-Figueroa, A., Martin, P., Schmelz, R.M., Wetzel, M.J., Wiklund, H., Maciolek, N.J., Kawauchi, G.Y. & Williams, J.D. (2021) Segmented worms (Phylum Annelida): a celebration of twenty years of progress through Zootaxa and call for action on the taxonomic work that remains. *Zootaxa*, 4979, 190–211. https://doi.org/10.11646/zootaxa.4979.1.18
- Maggenti, A.R. (1991) Nemata: higher classification. *In*: Nickle, W.R. (ed) *Manual of agricultural nematology*. Marcel Dekker, New York, pp. 147–187.
- Maggenti, A.R., Luc, M., Raski, D.J., Fortuner, R. & Geraert, E. (1987) A reappraisal of Tylenchina (Nemata). 2. Classification of the suborder Tylenchina (Nemata: Diplogasteria). *Revue de Nematologie*, 10, 135–142.
- Maggenti, A.R., Luc, M., Raski, D.J., Fortuner, R. & Geraert, E. (1988) A reappraisal of Tylenchina (Nemata). 11. List of generic and supra-generic taxa, with their junior synonyms. *Revue de Nematologie*, 11, 177–188.
- Maia, C., Casero, M., Annoscia, G., Latrofa, M.S., Colella, V., Pereira, A., Azevedo, F. & Otranto, D. (2017) *Cercopithifilaria* sp II in *Vulpes vulpes*: new host affiliation for an enigmatic canine filarioid. *Parasitology Research*, 116, 441–443.
- Mallet, J. (2008) Hybridization, ecological races and the nature of species: empirical evidence for the ease of speciation. *Philosophical Transactions of the Royal Society Series B*, 363, 2971–2986.
- Mamiya, Y. (1986) Interspecific hybridization between *Bursaphelenchus xylophilus* and *B. mucronatus* (Aphelenchida: Aphelenchoididae). *Applied Entomology and Zoology*, 21, 159–163.
- Marin, D.H., Kaplan, D.T. & Opperman, C.H. (1999) Randomly amplified polymorphic DNA differs with burrowing nematode collection site, but not with host range. *Journal of Nematology*, 31, 232–239.
- Mattiucci, S., Turchetto, M., Bragantini, F. & Nascetti, G. (2002) On the occurrence of the sibling species *Contracaecum rudolphii* complex (Nematoda: Anisakidae) in cormorants (*Phalacrocorax carbo sinensis*) from Venice and Caorle lagoons: genetic markers and ecological studies. *Parassitologia*, 44, 105.
- Mattiucci, S., Cipriani, P., Webb, S.C., Paoletti, M., Marcer, F., Bellisario, B., Gibson, D.I. & Nascetti, G. (2014) Genetic and morphological approaches distinguish the three sibling species of the *Anisakis simplex* species complex, with a species designation as *Anisakis berlandi* n. sp for *A. simplex* sp C (Nematoda: Anisakidae). *Journal of Parasitology*, 100, 199– 214.
- May, R.M. (1988) How many species are there on earth. Science, 241, 1441-1449.
- Mayden, R.L. (1997) A hierarchy of species concepts: the denouement in the saga of the species problem. *Systematics Association Special Volume Series*, 54, 381–424.
- Mayr, E. & Ashlock, P.D. (1991) Principles of Systematic Zoology. McGraw-Hill, New York, 475 pp.
- Mayr, E., Linsley, E.G. & Usinger, R.L. (1953) *Methods and Principles of Systematic Zoology*. McGraw-Hill, New York, 328 pp.
- McInnes, S.J., Jorgensen, A. & Michalczyk, U. (2021) 20 years of Zootaxa: Tardigrada (Ecdysozoa: Panarthropoda). Zootaxa, 4979 (1), 23–24.

https://doi.org/10.11646/zootaxa.4979.1.5

- Meldal, B.H.M., Debenham, N.J., De Ley, P., De Ley, I.T., Vanfleteren, J.R., Vierstraete, A.R., Bert, W., Borgonie, G., Moens, T., Tyler, P.A., Austen, M.C., Blaxter, M.L., Rogers, A.D. & Lambshead, P.J.D. (2007) An improved molecular phylogeny of the Nematoda with special emphasis on marine taxa. *Molecular Phylogenetics and Evolution*, 42, 622–636.
- Metge, K., Braasch, H., Gu, J. & Burgermeister, W. (2008) Variation in ITS and 28S rDNA of Bursaphelenchus Species (Nematoda: Parasitaphelenchidae). In: Mota, M.M. & Vieira, P. (eds) Pine Wilt Disease—A Worldwide Threat to Forest Ecosystems. Gulbenkian Foundation, Lisbon, pp. 151–164.
- Metge, K. & Burgermeister, W. (2008) Analysis of *Bursaphelenchus xylophilus* (Nematoda: Parasitaphelenchidae) provenances using ISSR and RAPD fingerprints. *In*: Mota, M.M. & Vieira, P. (eds) *Pine Wilt Disease—A Worldwide Threat to Forest Ecosystems*. Gulbenkian Foundation, Lisbon, pp. 175–186.
- Meza, P., Aballay, E. & Hinrichsen, P. (2012) Morphological and molecular characterisation of *Xiphinema index* Thorne and Allen, 1950 (Nematoda: Longidoridae) isolates from Chile. *Nematropica*, 42, 41–47.
- Mizukubo, T., Orui, Y., Hanada, K. & Sana, Z. (2003) Microevolutionary trend in *Pratylenchus coffeae* sensu stricto (Nematoda: Pratylenchidae): the diversity in PCR-RFLP phenotype, compatibility on host plants and reproductive segregation. *Japanese Journal of Nematology*, 33, 57–76.
- Montarry, J., Bardou-Valette, S., Mabon, R., Jan, P.L., Fournet, S., Grenier, E. & Petit, E.J. (2019) Exploring the causes of small effective population sizes in cyst nematodes using artificial *Globodera pallida* populations. *Proceedings of the Royal Society B-Biological Sciences*, 286, 9.
- Montarry, J., Jan, P.L., Gracianne, C., Overall, A.D., Bardou-Valette, S., Olivier, E. & Petit, E.J. (2015) Heterozygote deficits in cyst plant-parasitic nematodes: Possible causes and consequences. *Molecular Ecology*, 24, 1654–1677.
- Moravec, F. & Ali, A.H. (2014) Additional observations on *Philometra* spp. (Nematoda: Philometridae) in marine fishes off Iraq, with the description of two new species. *Systematic Parasitology*, 87, 259–271.
- Moravec, F., Ali, A.H., Abed, J.M. & Shaker, S.J. (2016) New records of philometrids (Nematoda: Philometridae) from marine fishes off Iraq, with the erection of two new species and the first description of the male of *Philometroides eleutheronemae* Moravec & Manoharan, 2013. *Systematic Parasitology*, 93, 129–144.
- Moravec, F., Bakenhaster, M. & Fajer-Avila, E.J. (2014) Three new gonad-infecting species of *Philometra* (Nematoda: Philometridae) parasitic in *Lutjanus* spp. (Lutjanidae) in the northern Gulf of Mexico off Florida, USA. *Folia Parasitologica*,

61, 355-369.

- Moravec, F. & Barton, D.P. (2015) Two gonad-infecting species of *Philometra* (Nematoda: Philometridae) from marine fishes off the northern coast of Australia. *Parasite*, 22. https://doi.org/10.1051/parasite/2015008
- Moravec, F. & Diggles, B.K. (2014a) Two new gonad-infecting species of *Philometra* Costa, 1845 (Nematoda: Philometridae)
- from marine fishes off the northern coast of Australia. Systematic Parasitology, 89, 33–44.
  Moravec, F. & Diggles, B.K. (2014b) Philometrid nematodes (Philometridae) from marine fishes off the northern coast of Australia, including three new species. *Folia Parasitologica*, 61, 37–54.
- Moravec, F. & Diggles, B. (2015a) *Philometra mirabilis* sp n. (Nematoda: Philometridae), a new gonad-infecting parasite from the freshwater fish *Cichla mirianae* (Cichlidae) in Brazilian Amazon. *Parasitology Research*, 114, 1929–1932.
- Moravec, F. & Diggles, B.K. (2015b) A new gonad-infecting species of *Philometra*, *P.barnesi* sp n. (Nematoda: Philometridae), from the marine fish *Pomadasys argenteus* (Haemulidae) off the northern coast of Australia. *Parasitology Research*, 114, 4121–4126.
- Moravec, F., Diggles, B., Barnes, L. & Macbeth, W. (2015a) Buckleyella ornata n. sp. (Nematoda: Philometridae) from the abdominal cavity of the talang queenfish Scomberoides commersionnianus (Perciformes: Carangidae) off the northern coast of Australia. Helminthologia, 51, 230–235.
- Moravec, F. & Jirku, M. (2014a) *Rhabdochona* spp. (Nematoda: Rhabdochonidae) from fishes in the Central African Republic, including three new species. *Folia Parasitologica*, 61, 157–172.
- Moravec, F. & Jirku, M. (2014b) *Dujardinascaris mormyropsis* n. sp (Nematoda: Anisakidae) from the osteoglossiform fish *Mormyrops anguilloides* (Linnaeus) (Mormyridae) in Central Africa. *Systematic Parasitology*, 88, 55–62.
- Moravec, F. & Jirku, M. (2015) Two *Procamallanus (Spirocamallanus)* species (Nematoda: Camallanidae) from freshwater fishes in the Lower Congo River. *Acta Parasitologica*, 60, 226–233.
- Moravec, F. & Justine, J.L. (2014a) *Capillaria plectropomi* n. sp (Nematoda: Capillariidae), a new intestinal parasite of the leopard coral grouper *Plectropomus leopardus* (Serranidae) off New Caledonia. *Parasite*, 21, 76.
- Moravec, F. & Justine, J.L. (2014b) Philometrids (Nematoda: Philometridae) in carangid and serranid fishes off New Caledonia, including three new species. *Parasite*, 21, 21.
- Moravec, F. & Justine, J.L. (2015a) New records of species of *Philometra* (Nematoda: Philometridae) from marine fishes off New Caledonia, including *P. cephalopholidis* sp n. from *Cephalopholis sonnerati* (Serranidae). *Parasitology Research*, 114, 3223–3228.
- Moravec, F. & Justine, J.L. (2015b) Anisakid nematodes (Nematoda: Anisakidae) from the marine fishes *Plectropomus laevis* Lacepede (Serranidae) and *Sphyraena qenie* Klunzinger (Sphyraenidae) off New Caledonia, including two new species of *Hysterothylacium* Ward & Magath, 1917. *Systematic Parasitology*, 92, 181–195.
- Moravec, F., Kamchoo, K. & Pachanawan, A. (2015b) New nematode species, *Orientatractis mekongensis* n. sp (Atractidae) and *Neosynodontisia suratthaniensis* n. g., n. sp (Pharyngodonidae) from freshwater fishes in Thailand. *Systematic Parasitology*, 92, 197–209.
- Moravec, F., Khosheghbal, M. & Pazooki, J. (2014) Dichelyne (Dichelyne) spinigerus sp nov (Nematoda: Cucullanidae) from the marine fish Otolithes ruber (Sciaenidae) off Iran and first description of the male of Philometra otolithi Moravec et Manoharan, 2013 (Nematoda: Philometridae). Acta Parasitologica, 59, 229–237.
- Moravec, F. & Manoharan, J. (2014a) Gonad-infecting species of *Philometra* (Nematoda: Philometridae) from groupers *Epinephelus* spp. (Osteichthyes: Serranidae) in the Bay of Bengal, India. *Acta Parasitologica*, 59, 596–605.
- Moravec, F. & Manoharan, J. (2014b) Two new gonad-infecting species of *Philometra* (Nematoda: Philometridae) parasitic in *Lutjanus* spp. (Osteichthyes: Lutjanidae) in the Bay of Bengal, India. *Parasitology Research*, 113, 3299–3307.
- Moravec, F. & Van As, L.L. (2015a) Procamallanus (Procamallanus) spp. (Nematoda: Camallanidae) in fishes of the Okavango River, Botswana, including the description of P. (P.) pseudolaeviconchus n. sp parasitic in Clarias spp. (Clariidae) from Botswana and Egypt. Systematic Parasitology, 90, 137–149.
- Moravec, F. & Van As, L.L. (2015b) Procamallanus (Spirocamallanus) spp. (Nematoda: Camallanidae) from fishes of the Okavango River, Botswana, including P. (S.) serranochromis n. sp parasitic in Serranochromis spp. (Cichlidae). Systematic Parasitology, 90, 151–164.
- Mugniery, D. (1979) Hybridization between *Globodera rostochiensis* and *Globodera pallida*. *Revue de Nematologie*, 2, 153–160.
- Mugniery, D., Bossis, M., & Pierre, J.-S. (1992) Hybridization between *Globodera rostochiensis* (Wollenweber), *G. pallida* (Stone), *G. virginiae* (Miller & Gray), *G. solanacearum* (Miller & Gray) and *Globodera mexicana* (Campos-Vela). Description and future of the hybrids. *Fundamental and Applied Nematology*, 15, 375–382.
- Mueller, M.I., Morais, D.H., Costa-Silva, G.J., Aguiar, A., Avila, R.W. & da-Silva, R.J. (2018) Diversity in the genus *Rhabdias* (Nematoda, Rhabdiasidae): Evidence for cryptic speciation. *Zoologica Scripta*, 47, 595–607.
- Nadler, S.A., Carreno, R.A., Adams, B.J., Kinde, H., Baldwin, J.G. &Mundo-Ocampo, M. (2003) Molecular phylogenetics and diagnosis of soil and clinical isolates of *Halicephalobus gingivalis* (Nematoda: Cephalobina: Panagrolaimoidea), an opportunistic pathogen of horses. *International Journal for Parasitology*, 33, 1115–1125.
- Nascetti, G., Cianchi, R., Mattiucci, S., D'Amelio, S., Orecchia, P., Paggi, L., Brattey, J., Berland, B., Smith, J.W. & Bullini, L. (1993) Three sibling species within *Contracaecum osculatum* (Nematoda, Ascarida, Ascaridoidea from the atlantic arcticboreal region: reproductive isolation and host preferences. *International Journal for Parasitology*, 23, 105–120.

- Nguyen, C.N., Subbotin, S.A., Madani, M., Trinh, P.Q. & Moens, M. (2003) *Radopholus duriophilus* sp. n. (Nematoda : Pratylenchidae) from western highland of Vietnam. *Nematology*, 5, 549–558.
- Nicholas, W.L. (2001) Seasonal variations in nematode assemblages on an Australian temperate ocean beach; the effect of heavy seas and unusually high tides. *Hydrobiologia*, 464, 17–26.
- Nicholas, W.L. & Hodda, M. (1999) The free-living nematodes of a temperate, high energy, sandy beach: faunal composition and variation over space and time. *Hydrobiologia*, 394, 113–127.
- Nixon, K.C. & Carpenter, J.M. (2000) On the other "phylogenetic systematics". Cladistics, 16, 298-318.
- Nixon, K.C., Carpenter, J.M. & Stevenson, D.W. (2003) The PhyloCode is fatally flawed, and the "Linnaean" system can easily be fixed. *Botanical Review*, 69, 111–120.
- Norton, D.C. & Niblack, T.L. (1991) Biology and Ecology of nematodes. *In*: Nickle, W.L. (ed) *Manual of Agricultural Nematology*. Marcel Dekker, New York, pp. 47–72.
- OECD (2011) OECD Science, Technology and Industry Scoreboard 2011. OECD Publishing, Paris, 204 pp.
- Orecchia, P., Mattiucci, S. & D'Amelio, S. (1994) Two new members in the *Contracaecum osculatum* complex (Nematoda, Ascaridoidea) from the Antarctic. *International Journal for Parasitology*, 24, 367–377.
- Otranto, D., Brianti, E., Dantas-Torres, F., Weigl, S., Latrofa, M.S., Gaglio, G., Cauquil, L., Giannetto, S. & Bain, O. (2011) Morphological and molecular data on the dermal microfilariae of a species of *Cercopithifilaria* from a dog in Sicily. *Veterinary Parasitology*, 182, 221–229.
- Otranto, D., Brianti, E., Latrofa, M.S., Annoscia, G., Weigl, S., Lia, R.P., Gaglio, G., Napoli, E., Giannetto, S., Papadopoulos, E., Miro, G., Dantas-Torres, F. & Bain, O. (2012) On a *Cercopithifilaria* sp transmitted by *Rhipicephalus sanguineus*: a neglected, but widespread filarioid of dogs. *Parasites & Vectors*, 5, 1.
- Otranto, D., Brianti, E., Dantas-Torres, Miro, G., Ltrofa, M.S., Mutafchiev, Y. & Bain, O. (2013a) Species diversity of dermal microfilariae of the genus *Cercopithifilaria* infesting dogs in the Mediterranean region. *Parasitology*, 140, 99–108.
- Otranto, D., Varcasia, A., Solinas, C., Scala, A., Brianti, E., Dantas-Torres, Annoscia, G., Martin, C., Mutafchiev, Y. & Bain, O. (2013b) Redescription of *Cercopithifilaria bainae* Almeida & Vicente, 1984 (Spirurida, Onchocercidae) from a dog in Sardinia, Italy. *Parasites and Vectors*, 6, 11.
- Palomares-Rius, J.E., Cantalapiedra-Navarrete, C. & Castillo, P. (2014) Cryptic species in plant-parasitic nematodes. *Nematology*, 16, 1105–1118.
- Pantoja, C.S., Perieira, F.B., Santos, C.P. & Luque, J.L. (2016) Morphology and molecular characterization hold hands: clarifying the taxonomy of *Hysterothylacium* (Nematoda: Anisakidae) larval forms. *Parasitology Research*, 115, 4353–4364.
- Paulson, D.R. & Marinov, M. (2021) Zootaxa 20th Anniversary Celebration: Odonata section. *Zootaxa*, 4979 (1), 218–221. https://doi.org/10.11646/zootaxa.4979.1.21
- Payne, A. (2015) Why do taxonomists write the meanest obituaries? Nautilus, 35, 1.
- Peccoud, J., Ollivier, A., Plantegenest, M. & Simon, J.C. (2009) A continuum of genetic divergence from sympatric host races to species in the pea aphid complex. *Proceedings of the National Academy of Science of the USA*, 106, 7495–7500.
- Pedram, M., Pourjam, E., Palomares-Rius, J.E., Ghaemi, R., Cantalapiedra-Navarrete, C. & Castillo, P. (2012a) Molecular and morphological characterisation of *Xiphinema granatum* n. sp and *Longidorus pisi* Edward, Misra & Singh, 1964 (Dorylaimida: Longidoridae) from Iran. *Nematology*, 14, 949–960.
- Pedram, M., Pourjam, E., Robbins, R.T., Ghaemi, R., Cantalapiedra-Navarrete, C. & Castillo, P. (2012b) Morphological and molecular characterisation of *Xiphinema mazandaranense* n. sp (Dorylaimida: Longidoridae), a new member of the *Xiphinema pyrenaicum* species complex. *Nematology*, 14, 109–119.
- Pena, E.de la, Vandegehuchte, M.L., Bonte, D. & Moens, M. (2011) Nematodes surfing the waves: long-distance dispersal of soil-borne microfauna via sea swept rhizomes. *Oikos*, 120, 1649–1656.
- Pena-Santiago, R. (2019) Taxonomy of the genus *Labronema* Thorne, 1939 (Nematoda: Dorylaimida: Dorylaimidae) with redescription of its type species *L. ferox* Thorne, 1939. *Nematology*, 21, 23–34.
- Peng, W., Anderson, T.J.C., Zhou, X. & Kennedy, M.W. (1998) Genetic variation in sympatric Ascaris populations from humans and pigs in China. Parasitology, 17, 355–361.
- Peng, W., Yuan, K., Hu. M. & Gasser, R.B. (2005) Mutation scanning-coupled analysis of haplotypic variability in mitochondrial DNA regions reveals low gene flow between human and porcine *Ascaris* in endemic regions of China. *Electrophoresis*, 26, 4317–4326.
- Perez-Gonzalez, A., Acosta, L.E., Proud, D.N & Hultz, J.W. (2021) Harvestmen in the first twenty years: a scientometric analysis of Zootaxa's contribution to opilionology (Arthropoda, Arachnida, Opiliones). Zootaxa, 4979, 102–114. https://doi.org/10.11646/zootaxa.4979.1.12
- Perez-Ponce de Leon, G. & Poulin, R. (2018) An updated look at the uneven distribution of cryptic diversity among parasitic helminths. *Journal of Helminthology*, 92, 197–202.
- Perry, R.N. (1999) Desiccation survival of parasitic nematodes. Parasitology, 119, S19-S30.
- Perry, R.N., Plowright, R.A. & Webb, R.M. (1980) Mating between *Pratylenchus penetrans* and *Pratylenchus fallax* in sterile culture. *Nematologica*, 26, 125–129.
- Picard, D. & Plantard, O. (2006) What constitutes a population for the plant parasitic nematode *Globodera pallida* in its native area (Peru)? *International Journal for Parasitology*, 36, 115–122.
- Picard, D., Plantard, O., Scurrah, M. & Mugniery, D. (2004) Inbreeding and population structure of the potato cyst nematode (*Globodera pallida*) in its native area (Peru). *Molecular Ecology*, 13, 2899–2908.

- Pinto, A.P., Mejdalani, G., Mounce, R., Silveira, L.F., Marinoni, L. & Rafael, J.A. (2021) Are publications on zoological taxonomy under attack? *Royal Society Open Science*, 8, 201617.
- Platnick, N.I. (2012) The Poverty of the Phylocode: A Reply to de Queiroz and Donoghue. Systematic Biology, 61, 360-361.
- Platnick, N.I. (2013) The Information Content of Taxon Names: A Reply to de Queiroz and Donoghue. *Systematic Biology*, 62, 175–176.
- Platt, H.M. (1994) Foreword. *In*: Lorenzen, S. *The phylogenetic systematics of freeliving nematodes (English translation: Platt H.M. (ed))*. Ray Society Publications, London, pp 5–6.
- Pleijel, F. (1999) Phylogenetic taxonomy, a farewell to species, and a revision of Heteropodarke (Hesionidae, Polychaeta, Annelida). *Systematic Biology*, 48, 755–789.
- Plowright, R., Dusabe, J., Coyne, D. & Speijer, P. (2013) Analysis of the pathogenic variability and genetic diversity of the plant-parasitic nematode *Radopholus similis* on bananas. *Nematology*, 15, 41–56.
- Poulin, R., Hay, E. & Jorge, F. (2019) Taxonomic and geographic bias in the genetic study of helminth parasites. *International Journal for Parasitology*, 49, 429–435.
- Poulin, R. & Jorge, F. (2019) The geography of parasite discovery across taxa and over time. Parasitology, 146, 168–175.
- Prior, T., Hockland, S. & Decraemer, W. (2010) A new approach to identify species in *Xiphinema americanum* sensu lato. *Communications in agricultural and applied biological sciences*, 75, 459–61.
- Ptatscheck, C., Gansfort, B. & Traunspurger, W. (2018) The extent of wind-mediated dispersal of small metazoans, focusing on nematodes. *Scientific Reports*, 8, 6814.
- Purwaningsih, E. & Smales, L.R. (2010) Two new species of *Dorcopsistrongylus* (Strongylida: Strongyloidea) from *Dorcopsis muelleri* (Marsupialia: Macropodidae) from Papua Indonesia. *Journal of Parasitology*, 96, 596–601.
- Purwaningsih, E. & Smales, L.R. (2011) Two new species of Labiostrongylinea (Nematoda: Cloacininae) from Salawati Island, Indonesia. *Transactions of the Royal Society of South Australia*, 135, 124–133.
- Purwaningsih, E. & Smales, L.R. (2014) New species of *Dorcopsinema* and *Paralabiostrongylus* (Nematoda: Chabertiidae: Cloacininae) from Indonesia, with a key to species of *Dorcopsinema*. *Zootaxa*, 3857, 591–598.
- Ragsdale, E.J., Kanzaki, N., Roseler, W., Herrmann, M. & Sommer, R.J.(2013) Three new species of *Pristionchus* (Nematoda: Diplogastridae) show morphological divergence through evolutionary intermediates of a novel feeding-structure polymorphism. *Zoological Journal of the Linnean Society*, 168, 671–698.
- Raymond, M.R., Wharton, D.A. & Marshall, C.J. (2014) Nematodes from the Victoria Land coast, Antarctica and comparisons with cultured *Panagrolaimus davidi*. *Antarctic Science*, 26, 15–22.
- Renner, S.S (2016) A Return to Linnaeus's Focus on Diagnosis, Not Description: The Use of DNA Characters in the Formal Naming of Species. *Systematic Biology*, 65, 1085–1095.
- Riga, E., Beckenbach, K. & Webster, J.M. (1992) Taxonomic relationships of *Bursaphelenchus xylophilus* and *B. mucronatus* based on interspecific and intraspecific cross-hybridization and DNA analysis. *Fundamental and Applied Nematology*, 15, 391–395.
- Rivera-Correa, M., Baldo, D., Candioti, F.V., Orrico, V.G.D., Blackburn, D.C., Castroviejo-Fisher, S., Chan, K.O., Gambale, P., Gower, D.J., Quah, E.S.H., Jodi, J., Rowley, L., Twomey, E. & Vences, M. (2021) Amphibians in *Zootaxa*: 20 years documenting the global diversity of frogs, salamanders, and caecilians. *Zootaxa*, 4979, 59–69. https://doi.org/10.11646/zootaxa.4979.1.9
- Rossberg, A.G., Rogers, T. & McKane, A.J. (2013) Are there species smaller than 1 mm? *Proceedings of the Royal Society Series B Biological sciences*, 280, 20131248.
- Roux, C., Fraisse, C., Romiguier, J., Anciaux, Y., Galtier, N. & Bierne, N. (2016) Shedding light on the grey zone of speciation along a continuum of genomic divergence. *PLoS Biology*, 14, e2000234.
- Sakai, H., Takeda, A. & Mizukubo, T. (2012) Intra-specific variation of *Xiphinema brevicolle* Lordello et Costa, 1961 (Nematoda: Longidoridae) in Japan. *Nematological Research*, 42, 1–7.
- Santos, M.C., Amarante, M.R.V. & Amarante, A.F.T. (2019) Establishment of co-infection and hybridization of *Haemonchus* contortus and *Haemonchus placei* in sheep. Journal of Helminthology, 93, 697–703.
- Schauer-Blume, M. (1990) Preliminary investigations on pathogenicity of European *Bursaphelenchus* species in comparison to *Bursaphelenchus xylophilus* from Japan. *Revue de Nematologie*, 13, 191–195.
- Schoonmaker, A., Hao, Y., Bird, D.McK. & Conant, G.C. (2020) A single, shared triploidy in three species of parasitic nematodes. *G3-Genes Genomes Genetics*, 10, 225–233.
- Seehausen, O. (2004) Hybridization and adaptive radiation. Trends in Ecology & Evolution, 19, 198-207.
- Seehausen, O., Butlin, R.K., Keller, I., Wagner, C.E., Boughman, J.W., Hohenlohe, P.A. & Widmer, A. (2014) Genomics and the origin of species. *Nature Reviews Genetics*, 15, 176.
- Shamsi, S. (2014) Recent advances in our knowledge of Australian anisakid nematodes. *International Journal for Parasitology: Parasites and Wildlife*, 178–187.
- Shamsi, S., Steller, E. & Chen, Y. (2018) New and known zoonotic nematode larvae within selected fish species from Queensland waters in Australia. *International Journal of Food Microbiology*, 272, 73–82.
- Siddiqi, M.R. (2015) Descriptions of seven new species of the nematode genus *Miconchus* Andrassy, 1958 (Mononchida). *International Journal of Nematology*, 25, 145–165.
- Siddiqi, M.R. & Hahn, M.L. (1995) *Radopholus bridgei* sp. n. (Tylenchida: Pratylenchidae) from Indonesia and its differentiation by morphological and molecular characters. *Afro-Asian Journal of Nematology*, 5, 38–43.

- Simsek, E., Pekmezci, G.Z., Yildirim, A., Duzlu, O., Onder, Z., Ciloglu, A., Sursal, N., Yilmaz, E., Gonulalan, Z. & Inci, A. (2020) Investigation of *Anisakis* larvae in different products of ready-to-eat fish meat and imported frozen fish in Turkey. *International Journal of Food Microbiology*, 333, 108829.
- Skoracka, A., Magalhaes, S., Rector, B.G. & Kuczynski, N. (2015) Cryptic speciation in the Acari: a function of species lifestyles or our ability to separate species? *Experimental & Applied Acarology*, 67, 165–182.
- Skrzypczak, M., Rokicki, J., Pawliczka, I., Najda, K. & Dzido, J. (2014) Anisakids of seals found on the southern coast of Baltic Sea. Acta Parasitologica, 59, 165–172.
- Slos, D., Sudhaus, W., Stevens, L., Bert, W. & Blaxter, M. (2017) Caenorhabditis monodelphis sp. n.: defining the stem morphology and genomics of the genus Caenorhabditis. BMC Zoology, 2,4.
- Smales, L.R. (2010) The gastrointestinal helminths of *Lorentzimys nouhuysi* (Rodentia: Muridae) with descriptions of two new genera and three new species (Nematoda) from Papua New Guinea. *Journal of Parasitology*, 96, 602–613.
- Smales, L.R. (2011a) New species and new locality records of the nematode genus *Labiosimplex* (Strongylida: Chabertiidae) from macropodid marsupials in Western Australia. *Records of the Western Australian Museum*, 26, 183–190.
- Smales, L.R. (2011b) The gastrointestinal helminths of *Chiruromys vates* (Rodentia: Muridae) with the description of a new species (Nematoda: Heligmonellidae) from Papua New Guinea. *Comparative Parasitology*, 78, 327–332.
- Smales, L.R. (2011c) Gastrointestinal nematodes of Coccymys ruemmleri (Rodentia, Muridae) with the description Montistrongylus giluwensis sp nov (Heligmonellidae) and Syphacia coccymyos sp nov (Oxyuridae) from Papua New Guinea. Acta Parasitologica, 56, 418–426.
- Smales, L.R. (2012) Helminths from Abeomelomys sevia (Rodentia: Muridae) with the description of a new species of Montistrongylus (Nematoda: Heligmonellidae) from Papua New Guinea. Comparative Parasitology, 79, 214–218.
- Smales, L.R. (2013) Nematodes from the caecum and colon of *Pogonomys* (Muridae: Anisomyini) from Papua New Guinea with the descriptions of a new genus of Oxyuridae (Nematoda: Oxyurida) and a new species of Trichuridae (Nematoda: Enoplida). *Zootaxa*, 3599, 577–587.
- Smales, L.R. (2014) Gastrointestinal helminths (Cestoda, Chabertiidae and Heligmonellidae) of *Pogonomys loriae* and *Pogonomys macrourus* (Rodentia: Muridae) from Papua Indonesia and Papua New Guinea with the description of a new genus and two new species. *Zootaxa*, 3889, 92–106.
- Smales, L.R. (2015) Nematodes of Heligmonellidae (Strongylida) of *Pogonomys championi* Flannery and *Pogonomys sylvestris* Thomas (Rodentia: Muridae) from Papua New Guinea with descriptions of five new species. *Systematic Parasitology*, 92, 113–129.
- Smales, L.R. & Heinrich, B. (2010) Gastrointestinal nematodes of *Paramelomys rubex* (Rodentia: Muridae) from Papua Indonesia and Papua New Guinea with the descriptions of three new genera and four new species of Heligmonellidae and Herpetostrongylidae (Nematoda: Trichostrongylida). *Zootaxa*, 2672, 1–28.
- Spratt, D.M. (2010) *Pelecitus bartneri* sp nov (Nematoda: Filarioidea) from the subcutaneous tissues of the leg of *Psephotus chrysopterygius* Gould, 1858 (Psittaciformes). *Transactions of the Royal Society of South Australia*, 134, 172–176.
- Spratt, D.M. (2011) New records of filarioid nematodes (Nematoda: Filarioidea) parasitic in Australasian monotremes, marsupials and murids, with descriptions of nine new species. *Zootaxa*, 2860, 1–61.
- Stevens, L., Felix, M.A., Beltran, T., Braendle, C., Caurcel, C., Fausett, S., Fitch, D., Frezal, L., Gosse, C., Kaur, T., Kiontke, K., Newton, M.D., Noble, L.M., Richaud, A., Rockman, M.V., Sudhaus, W. & Blaxter, M. (2019) Comparative genomics of 10 new *Caenorhabditis* species. *Evolution Letters*, 3, 217–236.
- Stork, N.E. (1993) How many species are there? *Biodiversity Conservation*, 2, 215–232.
- Stork, N.E. (2018) How many species of insects and other terrestrial arthropods are there on earth? *Annual Review of Entomology*, 63, 31–45.
- Subbotin, S.A., Deimi, A.M., Zheng, J. & Chizhov, V.N. (2011) Length variation and repetitive sequences of Internal Transcribed Spacer of ribosomal RNA gene, diagnostics and relationships of populations of potato rot nematode, *Ditylenchus destructor* Thorne, 1945 (Tylenchida: Anguinidae). *Nematology*, 13, 773–785.
- Sudhaus, W. (2011) Phylogenetic systematisation and catalogue of paraphyletic "Rhabditidae" (Secernentea, Nematoda). *Journal of Nematode Morphology and Systematics*, 14, 113–178.
- Susoy, V. & Herrmann, M. (2014) Preferential host switching and codivergence shaped radiation of bark beetle symbionts, nematodes of *Micoletzkya* (Nematoda: Diplogastridae). *Journal of Evolutionary Biology*, 27, 889–898.
- Susoy, V., Herrmann, M., Kanzaki, N., Kruger, M., Nguyen, C.N., Roedelserger, C., Roeseler, W., Weiler, C., Giblin-Davis, R.M., Ragsdale, E.J. & Sommer, R.J. (2016) Large-scale diversification without genetic isolation in nematode symbionts of figs. *Science Advances*, 2, e1501031.
- Szewczyk, N.J., Mancinelli, R.L., McLamb, W., Reed, D., Blumberg, B.S. & Conley, C.A. (2005) Caenorhabditis elegans survives atmospheric breakup of STS-107, space shuttle Columbia. Astrobiology, 5, 690–705. https://doi.org/10.1089/ast.2005.5.690
- Tandingan De Ley, I., Mundo-Ocampo, M., Yoder, M. & De Ley, P. (2007) Nematodes from vernal pools in the Santa Rosa Plateau Ecological Reserve, California I. *Hirschmanniella santarosae* sp. n. (Nematoda: Pratylenchidae), a cryptic sibling species of *H. pomponiensis* Abdel-Rahman & Maggenti, 1987. *Nematology*, 9, 405–429.
- Tchesunov, A.V., Portnova, D.A. & van Campenhout, J. (2015) Description of two free-living nematode species of *Halomonhystera disjuncta* complex (Nematoda: Monhysterida) from two peculiar habitats in the sea. *Helgoland Marine Research*, 69, 57–85.

- Thevenoux, R., Folcher, L., Esquibet, M., Fouville, D., Montarry, J. & Grenier, E. (2019) The hidden diversity of the potato cyst nematode *Globodera pallida* in the south of Peru. *Evolutionary Applications*, 13, 727–737.
- Thiery, M., Mugniery, D., Fouville, D. & Schots, A. (1996) Natural hybridization between *Globodera rostochiensis* and *G. pallida. Fundamental and Applied Nematology*, 19, 437–442.
- Thiery, M., Mugniery, D., Bossis, M. & SosaMoss, C. (1997) Crossing between *Globodera pallida* Stone and *G.* "mexicana" Campos-Vela: species concept and heritability of the development on potato. *Fundamental and Applied Nematology*, 20, 551–556.
- Thomas, W.K. (2007) NemAToL: Database for the nematode branch of the tree of life. Phytopathology, 97, S157-S157.
- Thorne, M.A.S., Seybold, A., Marshall, C. & Wharton, D. (2017) Molecular snapshot of an intracellular freezing event in an Antarctic nematode. *Cryobiology*, 75, 117–124.
- Traunspurger, W. & Michiels, I.C. (2006) Composition and distribution of free-living freshwater nematodes: global and local perspectives. In: Eyualem, A., Andrassy, I. & Traunspurger, W. (eds) *Freshwater nematodes: ecology and taxonomy*. CABI, Wallingford UK, pp. 46–76.
- Trinh, P.Q., Nguyen, C.N., Waeyenberge, L., Subbotin, S.A., Karssen, G. & Moens, M. (2004) Radopholus arabocoffeae sp n. (Nematoda : Pratylenchidae), a nematode pathogenic to Coffea arabica in Vietnam, and additional data on R. duriophilus. Nematology, 6, 681–693.
- Trinh, P.Q., Waeyenberge, L., Nguyen, C.N & Moens, M. (2012) Morphological and molecular diversity of *Radopholus* on coffee in Vietnam and description of *R. daklakensis* sp. n. from Robusta coffee. *Nematology*, 14, 65–83.
- Turaganivalu, U., Stirling, G.R. & Smith, M.K. (2013) Burrowing nematode (*Radopholus similis*): a severe pathogen of ginger in Fiji. *Australasian Plant Pathology*, 42, 431–436.
- Uribe, G.E.M., Bert, W., Vierstraete, A.R., de la Pena, E., Moens, M. & Decraemer, W. (2010) Burrowing nematodes from Colombia and their relationship with *Radopholus similis* populations, *R. arabocoffeae* and *R. duriophilus. Nematology*, 12, 619–629.
- Valette, C., Mounport, D., Nicole, M., Sarah, J.L. & Baujard, P. (1998) Scanning electron microscope study of two African populations of *Radopholus similis* (Nematoda : Pratylenchidae) and proposal of *R. citrophilus* as a junior synonym of *R. similis. Fundamental and Applied Nematology*, 21, 139–146.
- Vovlas, N., Troccoli, A., Palomares-Rius, J.E., De Luca, F., Liebanas, G., Landa, B.B., Subbotin, S.A. & Castillo, P. (2011) Ditylenchus gigas n. sp. parasitizing broad bean: a new stem nematode singled out from the Ditylenchus dipsaci species complex using a polyphasic approach with molecular phylogeny. Plant Pathology, 60, 762–775
- Venekey, V., Fonseca-Genevois, V.G. & Santos, P.J.P. (2010) Biodiversity of free-living marine nematodes on the coast of Brazil: a review. Zootaxa, 2568, 39–66.
- Wacekel, W.J, Kimenju, J.W., Sibanda, Z. & Talwana, H. (2010) Building capacity in plant nematology in sub-saharan Africa: contributions by nematology initiative of east and southern Africa (NIESA). *Journal of Nematology*, 42, 275.
- Weaver, H.J. & Smales, L.R. (2010) Three new species of *Syphacia* (Syphacia) (Oxyurida: Oxyuridae) from Queensland, Australia, and a key to the species present in the Australian bioregion. *Comparative Parasitology*, 77, 9–19.
- Weaver, H.J. & Smales, L.R. (2012) Parasite assemblages of Australian species of *Pseudomys* (Rodentia: Muridae: Murinae). Journal of Parasitology, 98, 30–35.
- Webster, J.M., Anderson, R.V., Baillie, D.L., Beckenbach, K., Curran, J. & Rutherford, T.A. (1990) DNA probes for differentiating isolates of the pinewood nematode species complex. *Revue de Nematologie*, 13, 255–263.
- Wharton, D.A. (1999) Parasites and low temperatures. Parasitology, 119, S7-S17.
- Wharton, D.A. (2002) Nematode survival strategies. *In*: Lee, D.L. (ed) *The biology of nematodes*. Taylor & Francis, London, pp 389–411.
- Whitmore, D., Gaimari, S.D., Nihei, S.S., Evenhuis, N.L., Kurina, O., Borkent, C.J., Sinclair, B.J., O'Hara, J.E., Zhang, Z.-Q., Moulton, J.K., Ribeiro, G.C., Bickel, D.J., Gilka, W., Andersen, T., Rossaro, B., Whittington, A.E., Lamas, C.J.E., Heller, K., Kehlmaier, C., Courtney, G.W., Kerr, P.H. & Blagoderov, V. (2021) Twenty years of Dipterology through the pages of Zootaxa. Zootaxa, 4979, 166–189.
  - https://doi.org/10.11646/zootaxa.4979.1.17
- Wilkins, J. S. (2003) The origins of species concepts: History, characters, modes and synapomorphies. PhD thesis, University of Melbourne, Melbourne
- Wilkins, J.S. (2009) Species: A History of the Idea. University of California Press, Berkeley, 303 pp.
- Wilkins, J.S. (2011) Philosophically speaking, how many species concepts are there? Zootaxa, 2765, 58-60.
- Xie, Y., Zhao, B., Hoberg, E.P., Li, M., Zhou, X., Gu, X., Lai, W., Peng, X. & Yang, G. (2018) Genetic characterisation and phylogenetic status of whipworms (*Trichuris* spp.) from captive non-human primates in China, determined by nuclear and mitochondrial sequencing. *Parasites & Vectors*, 11, 516.
- Xu, Y.M., Zhao, Z.Q. & Wang, J.M. (2013) An index to new genera and species of Nematoda in Zootaxa from 2007 to 2012. *Zootaxa*, 3646, 160–170.
- Ye, W., Giblin-Davis, R.M., Davies, K.A., Purcell, M.F., Scheffer, S.J., Taylor, G.S., Center, T.D., Morris, K. & Thomas, W.K. (2007) Molecular phylogenetics and the evolution of host plant associations in the nematode genus *Fergusobia* (Tylenchida : Fergusobiinae). *Molecular Phylogenetics and Evolution*, 45, 123–141.
- Yong, H.-S., Song, S.-L., Eamsobhana, P., Goh, S.-Y., Lim, P.-E., Chow, W.-L., Chan, K.-G. & Abrahams-Sandi, E. (2015) Mitochondrial genome supports sibling species of *Angiostrongylus costaricensis* (Nematoda: Angiostrongylidae). PLOS

One, 10, e0134581.

- Yoshida, K., Herrmann, M., Kanzaki, N., Weiler, C., Rödelsperger, C. & Sommer, R.J. (2018) Two new species of *Pristionchus* (Nematoda: Diplogastridae) from Taiwan and the definition of the pacificus species-complex sensu stricto. *Journal of Nematology*, 50, 355–368.
- Zeppelli, D., Vanreusel, A. & Danovaro, R. (2011) Cosmopolitanism and biogeography of the Genus *Manganonema* (Nematoda: Monhysterida) in the Deep Sea. *Animals*, 1, 291–305.
- Zhang, J., Kapli, P., Pavlidis, P. & Stamatakis, A. (2013) A general species delimitation method with applications to phylogenetic placements. *Bioinformatics*, 29, 2869–2876.
- Zhang, Y., Lin, R., Zhao, G, Yuan, Z., Song, H. & Zhu, X. (2009) ITS rDNA sequencing and phylogenetic analysis of the new species *Contracaecum rudolphii* C (Nematoda: Anisakidae). *Chinese Veterinary Science*, 39, 298–302.
- Zhang, Z.-Q. (2011) Accelerating biodiversity descriptions and transforming taxonomic publishing: the first decade of *Zootaxa*. *Zootaxa*, 2896, 1–7.
- Zhang, Z.-Q. (2013) Animal biodiversity: An update of classification and diversity in 2013. Zootaxa, 3703, 5–11.
- Zhang, Z.-Q. (2021) Contributions of *Zootaxa* to biodiversity discovery: an overview of the first twenty years. *Zootaxa*, 4979, 6–16.
- Zhang, Z.-Q., Schatz, H., Pfingstl, T., Goldschmidt, T., Martin, P., Pešić, V., Ramírez, M., Schmidt, K.-H., Fan, Q.-H., Mironov, S., Seeman, O. & Halliday, B. (2021) Discovering and documenting Acari: the first twenty years in Zootaxa. *Zootaxa*, 4979, 115–130.

https://doi.org/10.11646/zootaxa.4979.1.13

Zhao, Z.Q. (2007) An index to new species of Nematoda in Zootaxa from 2003 to 2006. Zootaxa, 1541, 65-68.

- Zhao, Z.Q., Davies, K.A., Alexander, B.J.R. & Riley, I.T. (2013a) Zeatylenchus pittosporum gen. n., sp n. (Tylenchida: Anguinata), from leaves of *Pittosporum tenuifolium* (Pittosporaceae) in New Zealand. *Nematology*, 15, 197–212.
- Zhao, Z.Q., Davies, K.A., Brenton-Rule, E.C., Grangier, J., Gruber, M.A.M., Giblin-Davis, R.M. & Lester, P.J. (2013b) Diploscapter formicidae sp n. (Rhabditida: Diploscapteridae), from the ant Prolasius advenus (Hymenoptera: Formicidae) in New Zealand. Nematology, 15, 109–123.
- Zhao, Z., Li, D., Davies, K.A. & Ye, W. (2015) *Schistonchus zealandicus* n. sp (Nematoda: Aphelenchoididae) associated with *Ficus macrophylla* in New Zealand. *Nematology*, 17, 53–66.
- Zullini, A. (1973) Su alcumi nematodi di alta quota del Nepal. Khumbu Himal, 4(3), 401-411.