



Updated checklist of the order Thysanoptera in Colombia

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Abstract

We present here the first comprehensive checklist of Thysanoptera in Colombia. This study is based on a literature review, examination of specimens deposited in museums, and fieldwork conducted from 2012 to 2018 across various agroecosystems in the Andean, Caribbean, and Orinoco regions. Our study reports 38 new records of Thysanoptera for Colombia, bringing the total number of documented species to 183.

Key words: Thripidae, diversity, new records, biogeography

Introduction

Thysanoptera is a diverse order, although relatively small when compared to other groups of insects. It comprises 6,458 species in 788 extant genera of which 187 species in 70 genera are fossils; the order is organized into two suborders and fifteen families (ThripsWiki 2023). Advances in knowledge about Thysanoptera diversity and biological interactions in the Neotropics have been published by Silva and Del-Claro (2010), who noted that most of the published studies focus on pest thrips species. Studies carried out in Panama (Goldaracena *et al.* 2012), Brazil (Silva & Del-Claro 2010; Cavalleri & Mound 2012; Lima & Mound 2016) and Argentina (De Borbón 2013; Alejo & Zamar 2021), demonstrate a great diversity of thrips species with significant implications for biological functions, ecological interactions, and their roles in both natural and agricultural systems. Mound (2002) reports an estimated richness of 2,000 species of Thysanoptera in the Neotropics. Specifically, 300 species have been reported in Mexico and Costa Rica (Mound & Marullo 1996), 246 species in Panama (Goldaracena *et al.* 2012), 627 species in Brazil (Monteiro & Lima 2023), and 154 species in Argentina (De Borbón & Zamar 2024).

Colombia is located in northern South America with a continental area of 1,141,748 km² (Ocampo *et al.* 2007; Hernández *et al.* 1992; Giraldo 2013) and is considered the second most biodiverse country in the world due to the uplift of the Andes. This created new habitats and local isolates that favored high speciation rates and endemisms of many taxa (Antonelli & Sanmartin 2011; Josse *et al.* 2012). Additionally, the intricate interplay of biogeographic mosaics spanning the Andean, Orinoco, Guiana, Caribbean, Pacific, and Amazon regions further contributes to Colombia's exceptional biodiversity (Hernández *et al.* 1992; IGAC 2012). Despite Colombia's remarkable floristic diversity, it remains one of the few neotropical nations lacking a comprehensive inventory of its Thysanoptera fauna.

As with other South American countries (Monteiro 2002; Mound 2014) the first studies on Colombian Thysanoptera date from the beginning of the XXth century, when North American and European researchers casually described new species after receiving material. After that, most records came from sporadic reports primarily focused on agroecosystems (Cárdenas & Corredor 1993; Arévalo *et al.* 2003) or specific taxa, such as those documented by Calixto (2005, 2007). Calixto (2005) published a list of 79 species in three families of Terebrantia collected in the

Bogotá plateau. Her master's thesis (2007) comprises a list of 144 species, 28 genera and four families of Terebrantia in Colombia. The latter research includes 34 species of *Frankliniella* Karny, along with several species identified at the genus level. Unfortunately, these results were never published and lack species confirmation by recognized experts in the field.

In Colombia, the richness of Thysanoptera is little known; however, Ebratt *et al.* (2019), reported 81 species in 20 genera and three taxonomic families, related to 10 cultivated plants in three geographic regions of Colombia. This was the latest broad work on thrips diversity in the country, as an effort to conduct wide surveys in several landscapes, including agroecosystems. Ebratt *et al.* (2019) also found that Thysanoptera species richness gradually decreases as altitude increases in inverse correlation with temperature. However, it was in the altitudinal range between 1,000 and 2,000 masl, where 45.5% of the total of the species was found, possibly due to greater floristic diversity, ecosystems with landscape heterogeneity, greater number of life zones and climatic variability.

The present paper provides the first extensive list of Thysanoptera records for Colombia, based on a literature review, examination of specimens deposited in entomological collections, and fieldwork conducted from 2012 to 2018 across various regions of the country. This checklist provides several new records and will serve as a foundation for carrying out future research on Thysanoptera in the second most diverse nation in the world.

Materials and methods

Our study involved a review of different sources of information about Thysanoptera from Colombia, namely (i) national digital databases (ICN–Instituto de Ciencias Naturales, Universidad Nacional de Colombia, Bogota; CTNI–AGROSAVIA–Colección Taxonomica Nacional de Insectos "Luis María Murillo", Bogota; Instituto Alexander von Humboldt–IAvH, Bogota; CEUA–Colección Entomológica Universidad de Antioquia, Medellin; MHN–UPN–Museo de Historia Natural, Universidad Pedagógica Nacional, Bogota); (ii) international museums (USNM–Smithsonian National Museum of Natural History housed in the United States Department of Agriculture, Beltsville, MD, USA; MNCN–Museo Nacional de Ciencias Naturales, Madrid, Spain; NHM–Natural History Museum, London, United Kingdom); and (iii) scientific literature reports, catalogs, taxonomic keys, and species descriptions.

Finally, the checklist provided here includes those taxa collected in Colombia by the present authors. From 2012 to 2018 we conducted 40 sampling excursions, each lasting 10 consecutive days in 23 of the 32 national departments, including 13 life zones and 73 agricultural systems in the Andean, Caribbean and Orinoco biogeographic regions (IGAC 2012) (Fig. 1). Altitude ranges of 500 meters were chosen, from 0 to 3,500 meters above sea level, using the climatic zonation by Caldas–Lang thermal floors or zones and Holdridge life zones (Holdridge 1967; Eslava *et al.* 1986; Pabón *et al.* 2001; Gutiérrez 2002).

We collected insect specimens of Thysanoptera from cultivated and native plants of different botanical families that we found in rural agricultural areas and those surrounding natural biological reserves of the National Natural Parks System (SPPN) and the Regional Autonomous Corporations of Colombia (Barbosa 1992; Giraldo 2013; CAR 2020; Resnatur 2024). In areas with commercial and backyard agroecosystems surveys on arboreal plants were included, according to the inventories conducted by Cárdenas *et al.* (1970), Cárdenas *et al.* (1972), Gómez & Rivera (1987), Escobar *et al.* (1993), Bermúdez (1997), Gamboa (2003), Arrieta *et al.* (2004), Fuentes *et al.* (2011). We determined the taxonomy of the plant material using taxonomic keys from the Flora of Colombia (Gentry 1996; Bernal *et al.* 2016); and we compared the material to specimens in the herbarium of the Instituto de Ciencias Naturales, Universidad Nacional de Colombia. At the end of the study, we obtained 4,495 samples in agroecosystems and wild plants in the Andean (2,563 samples), Caribbean (1,532 samples), and Orinoco (400 samples) regions (Fig. 1).

The thrips were sorted by morphotypes and identified. The specimens were cleared in cold 5% KOH, followed by washing with distilled water. Subsequently, adults were mounted in Hoyer's medium in slides, and dried in the oven for 48 h at 35–40°C, according to Mound & Marullo (1996). A few specimens of each species were mounted in Canada Balsam for the permanent collection according to Mound & Marullo (1996). The taxonomic determination of adults was carried out based on morphological characters according and available keys (Moulton 1948; Sakimura & O'Neill 1979; Sakimura 1981; Mound & Nakahara 1994; Mound & Marullo 1996; Monteiro 1999; Berzosa & Maroto 2003; Hoddle & Mound 2003; Berzosa 2006; De Borbón & Cardello 2006; Mound &

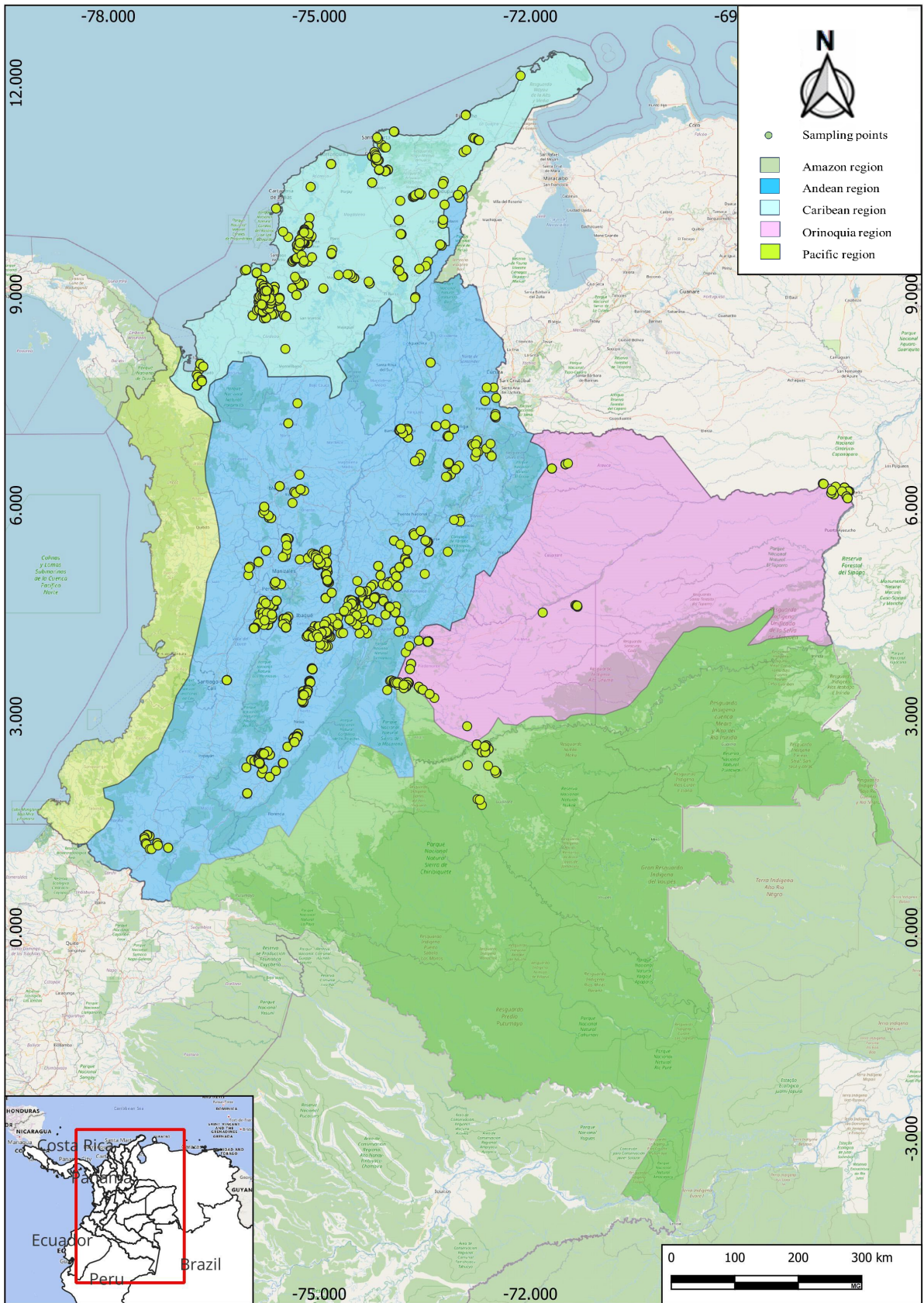


FIGURE 1. Total distribution of sampling points ($n = 4,495$) and geographic regions of Colombia, from 2012–2018.

Ng 2009; Cavalleri & Mound 2012; De Borbón 2013; Kumar *et al.* 2013; Lima & Mound 2016) with the help of a Nikon SMZ800 stereoscope and a Nikon Type-119YS2-T microscope with phase contrast and the image analysis programs NIS elements F (v. 4.6) and ToupView-x86. We also carried out direct comparisons with specimens kept in the private collection of Dr. Arturo Goldarazena at MNCN. All information regarding the material collected during fieldwork, including collecting data and associated plants, is available at the repository file of the MNCN (<https://doi.org/10.5281/zenodo.12595794>).

We deposited the Thysanoptera specimens in the Colección Taxonomica Nacional de Insectos "Luis María Murillo" (CTNI) of AGROSAVIA in Tibaitata research center in Mosquera Cundinamarca-Colombia.

Results

Our survey reports 183 Thysanoptera species for Colombia, belonging to the families Aeolothripidae, Heterothripidae, Phlaeothripidae, and Thripidae (Table 1). The fieldwork revealed 38 new species records for the country, most of which are *Frankliniella* species (14 spp.).

TABLE 1. Checklist of the Thysanoptera from Colombia compiling new records (*) identified through fieldwork conducted between 2012 and 2018, as well as specimens reviewed in collections and literature reports (Moulton 1933; USDA 1937, 1941; Díaz *et al.* 1978; Sakimura & O'Neill 1979; Cárdenas & Corredor 1993; Zapata *et al.* 1994; Mound & Marullo 1996; Marullo & Mound 2000; Arévalo *et al.* 2003; Calixto 2005; Berzosa 2006; Goldaracena & Mound 2010; NHM 2014; Cavalleri & Kaminski 2014; Ebratt *et al.* 2018, 2019; Pereyra *et al.* 2019).

Family	Subfamily	Species
Aeolothripidae		<i>Aeolothrips surcalifornianus</i> Johansen
		<i>Ambaeolothrips microstriatus</i> (Hood)
		<i>Erythrothrips diabolus</i> Priesner
		* <i>Erythrothrips stygicus</i> Hood
		<i>Franklinothrips orizabensis</i> Johansen
		<i>Franklinothrips vespiformis</i> (Crawford DL)
		* <i>Stomatothrips flavus</i> Hood
Heterothripidae		<i>Aulacothrips tenuis</i> Cavalleri & Kaminski
		<i>Heterothrips alvarezii</i> Johansen
		<i>Heterothrips analis</i> Hood
		<i>Heterothrips bolivianus</i> Stannard
		* <i>Heterothrips condei</i> Moulton
		<i>Heterothrips decoratus</i> Hood
		* <i>Heterothrips flavidus</i> Hood
		* <i>Heterothrips pedicellatus</i> Pereyra & Cavalleri
		* <i>Heterothrips prosopidis</i> Crawford JC
		<i>Heterothrips sericatus</i> Hood
		<i>Heterothrips watsoni</i> Bailey & Cott
		<i>Scutothrips byrsonimae</i> Mound & Marullo
		<i>Scutothrips incaensis</i> Stannard
	* <i>Scutothrips peruvianus</i> (Hood)	
Phlaeothripidae	Idolothripinae	<i>Diceratothrips picticornis</i> Hood
		* <i>Elaphrothrips laevicollis</i> (Bagnall)
	Phlaeothripinae	* <i>Adraneothrips pulchellus</i> Hood
		<i>Eupathithrips dentipes</i> Bagnall
		<i>Gomphiothrips mercedes</i> Mound & Marullo

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TABLE 1. (Continued)

Family	Subfamily	Species
Thripidae	Panchaethripinae	<i>Gynaikothrips ficorum</i> (Marchal)
		<i>Haplothrips gowdeyi</i> (Franklin)
		<i>Haplothrips graminis</i> Hood
		* <i>Holopothrips</i> sp.
		<i>Hoplandrothrips flavipes</i> Bagnall
		<i>Hoplandrothrips nigriceustus</i> Hood
		* <i>Karnyothrips flavipes</i> (Jones)
		<i>Karnyothrips merrilli</i> (Watson)
		<i>Leptothrips columbianus</i> Johansen
		<i>Liothrips salti</i> Moulton
		<i>Macrophthalthothrips hemipteroides</i> (Priesner)
		<i>Psalidothrips conciliatus</i> Hood
		<i>Psalidothrips sturmi</i> zur Strassen
		<i>Rhinoceps jansei</i> Faure
		<i>Trachythrips albipes</i> Hood
		* <i>Caliothrips phaseoli</i> (Hood)
		<i>Dinurothrips hookeri</i> Hood
		<i>Heliothrips haemorrhoidalis</i> (Bouché)
		<i>Parthenothrips dracaenae</i> Heeger
		<i>Selenothrips rubrocinctus</i> (Giard)
		Sericothripinae
	<i>Hydatothrips tricinctus</i> (Hood)	
	<i>Hydatothrips volcano</i> Mound & Marullo	
	<i>Neohydatothrips basilaris</i> (Hood)	
	<i>Neohydatothrips burungae</i> (Hood)	
	<i>Neohydatothrips gracilipes</i> (Hood)	
	<i>Neohydatothrips humberto</i> Mound & Marullo	
	* <i>Neohydatothrips inversus</i> (Hood)	
	<i>Neohydatothrips mirandai</i> (Johansen)	
	<i>Neohydatothrips rapoportii</i> Johansen	
	<i>Neohydatothrips signifer</i> (Priesner)	
	<i>Neohydatothrips tibialis</i> (Priesner)	
	Thripinae	<i>Anaphothrips obscurus</i> (Müller)
		<i>Anaphothrips sudanensis</i> Trybom
		<i>Apterothrips apteris</i> (Daniel)
		<i>Aptinothrips rufus</i> Haliday
		<i>Arorathrips fulvus</i> (Moulton)
		<i>Arorathrips mexicanus</i> (Crawford DL)
		<i>Arorathrips xanthius</i> (Hood)
		<i>Aurantothrips orchidaceus</i> (Bagnall)
		<i>Aurantothrips orchidearum</i> (Bondar)
		<i>Baileyothrips limbatus</i> (Hood)
* <i>Bolacothrips striatopennatus</i> (Schmutz)		

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TABLE 1. (Continued)

Family	Subfamily	Species
		<i>*Bravothrips kraussi</i> (Crawford JC)
		<i>*Bravothrips mexicanus</i> (Priesner)
		<i>Ceratothripoides brunneus</i> Bagnall
		<i>Ceratothripoides chilticus</i> Johansen
		<i>*Ceratothripoides claratris</i> (Shumsher)
		<i>Ceratothripoides funestus</i> (Hood)
		<i>Ceratothripoides silvestris</i> (Hood)
		<i>Chaetanaphothrips orchidii</i> (Moulton)
		<i>Charassothrips incomparabilis</i> (Johansen)
		<i>Charassothrips macroseta</i> Goldarazena & Mound
		<i>Charassothrips piperaffinis</i> (Johansen)
		<i>Charassothrips urospathae</i> Hood
		<i>Chirothrips manicatus</i> (Haliday)
		<i>Corynothrips stenopterus</i> Williams
		<i>Echinothrips caribbeanus</i> Hood
		<i>Frankliniella aliaepennae</i> Berzosa
		<i>*Frankliniella aureominuta</i> Johansen
		<i>Frankliniella auripes</i> Hood
		<i>Frankliniella bagnalliana</i> Hood
		<i>Frankliniella bicolor</i> Moulton
		<i>Frankliniella borinquen</i> Hood
		<i>Frankliniella brevicaulis</i> Hood
		<i>*Frankliniella breviseta</i> Moulton
		<i>Frankliniella bruneri</i> Watson
		<i>Frankliniella brunnea</i> (Priesner)
		<i>Frankliniella caseariae</i> Moulton
		<i>Frankliniella cassiae</i> Berzosa
		<i>Frankliniella caudiseta</i> Sakimura & O'Neill
		<i>Frankliniella cephalica</i> (Crawford DL)
		<i>Frankliniella chamulae</i> Johansen
		<i>Frankliniella citripes</i> Hood
		<i>Frankliniella colombiana</i> Moulton
		<i>*Frankliniella condei</i> John
		<i>Frankliniella cotubrusensis</i> Retana & Mound
		<i>Frankliniella crawfordi</i> Sakimura & O'Neill
		<i>Frankliniella curta</i> (Hood)
		<i>*Frankliniella davidsoni</i> Moulton
		<i>Frankliniella desmodii</i> Mound & Marullo
		<i>Frankliniella distinguenda</i> Bagnall
		<i>Frankliniella diversa</i> Hood
		<i>Frankliniella espeletiae</i> Berzosa
		<i>Frankliniella fallaciosa</i> Priesner
		<i>Frankliniella fulvipennis</i> Moulton

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TABLE 1. (Continued)

Family	Subfamily	Species
		* <i>Frankliniella fulvipes</i> Bagnall
		<i>Frankliniella gardeniae</i> Moulton
		<i>Frankliniella gemina</i> Bagnall
		<i>Frankliniella gossypiana</i> (Hood)
		<i>Frankliniella insularis</i> (Franklin)
		<i>Frankliniella invasor</i> Sakimura
		<i>Frankliniella kelliae</i> Sakimura
		<i>Frankliniella lorena</i> Mound & Marullo
		* <i>Frankliniella mekokara</i> Mound & Marullo
		<i>Frankliniella melanommata</i> Williams
		<i>Frankliniella minuta</i> (Moulton)
		<i>Frankliniella musaeperda</i> Hood
		* <i>Frankliniella nakaharai</i> Sakimura & O'Neill
		* <i>Frankliniella nigricauda</i> Hood
		<i>Frankliniella occidentalis</i> (Pergande)
		* <i>Frankliniella oxyura</i> Bagnall
		<i>Frankliniella panamensis</i> Hood
		<i>Frankliniella paramorum</i> Berzosa
		<i>Frankliniella parvula</i> Hood
		<i>Frankliniella peruviana</i> Hood
		<i>Frankliniella pestinae</i> Sakimura & O'Neill
		* <i>Frankliniella pulchella</i> Hood
		<i>Frankliniella regentis</i> Berzosa
		<i>Frankliniella rostrata</i> Priesner
		<i>Frankliniella salviae</i> Moulton
		<i>Frankliniella schultzei</i> (Trybom)
		<i>Frankliniella senckenbergiana</i> Berzosa & Maroto
		* <i>Frankliniella serrata</i> Moulton
		<i>Frankliniella simplex</i> Priesner
		* <i>Frankliniella sueoa</i> Mound & Marullo
		<i>Frankliniella trisetosa</i> Hood
		<i>Frankliniella tritici</i> (Fitch)
		* <i>Frankliniella tuberosi</i> Moulton
		<i>Frankliniella tympanona</i> Hood
		<i>Frankliniella valdiviana</i> Sakimura & O'Neill
		<i>Frankliniella vargasi</i> Retana & Mound
		* <i>Frankliniella varipes</i> Moulton
		<i>Frankliniella verbesinae</i> Sakimura
		<i>Frankliniella williamsi</i> Hood
		<i>Microcephalothrips abdominalis</i> (Crawford DL)
		<i>Nexothrips perseae</i> Marullo & Mound
		* <i>Oxythrips</i> sp.
		<i>Psectrothrips delostomae</i> Hood

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TABLE 1. (Continued)

Family	Subfamily	Species
		<i>Psectrothrips interruptus</i> (Hood)
		<i>Psectrothrips palmerae</i> Mound & Marullo
		<i>Pseudothrips retanae</i> Mound & Marullo
		<i>Rhamphothrips pandens</i> Sakimura
		* <i>Salpingothrips minimus</i> Hood
		* <i>Scirtidothrips torquatus</i> Hood
		<i>Scirtothrips bisbravae</i> Johansen
		<i>Scirtothrips dorsalis</i> Hood
		<i>Scirtothrips euthyntus</i> Mound & Marullo
		<i>Scirtothrips hansonii</i> Mound & Hodde
		* <i>Scirtothrips ikelus</i> Mound & Marullo
		<i>Scirtothrips lumarius</i> Mound & Marullo
		<i>Scirtothrips manihoti</i> (Bondar)
		<i>Scirtothrips multistriatus</i> Hood
		<i>Scirtothrips panamensis</i> Hood
		<i>Scirtothrips perseae</i> Nakahara
		<i>Scolothrips tenuipennis</i> zur Strassen
		<i>Stenchaetothrips biformis</i> (Bagnall)
		<i>Tenothrips frici</i> (Uzel)
		<i>Thrips australis</i> (Bagnall)
		* <i>Thrips brevipilosus</i> Moulton
		* <i>Thrips florum</i> Schmutz
		<i>Thrips helianthi</i> Morgan
		<i>Thrips palmi</i> Karny
		<i>Thrips simplex</i> (Morison)
		<i>Thrips tabaci</i> Lindeman
		* <i>Thrips trehernei</i> Priesner
		<i>Thrips vulgatissimus</i> Haliday

Thripidae is the most diverse family, with 142 species. Within this family, the richest genera are *Frankliniella* with 67 species, followed by *Scirtothrips* with 10 species and *Thrips* with nine species. This makes Colombia the South American country with the highest richness of the speciose genus *Frankliniella* (Ortiz 1977; Cavalleri & Mound 2012; Lima & Miyasato 2017; De Borbón & Zamar 2018; Cazorla-Perfetti 2019), which reflects the wide altitudinal and floristic Colombian diversity. On the other hand, only five of the almost 30 Panchaetothripinae species in South America (Lima *et al.* 2021) are recorded in Colombia. This indicates that the richness of thrips in Colombia is still underrepresented, and more surveys should be conducted in the country. All these species are primarily phytophagous, and some are mentioned as pests (Mound *et al.* 2022). The family Heterothripidae comprises 14 species in four genera in Colombia. All *Heterothrips* and *Scutothrips* species feed exclusively on flowers, whereas *Aulacothrips* species are ectoparasites of treehoppers (Mound & Marullo 1996; Cavalleri & Kaminski 2014).

Aeolothripidae is represented by seven species in five genera. This family includes facultative or obligatory predatory species, such as those in the genera *Franklinothrips* and *Stomatothrips*, and the flower-living *Aeolothrips* (Mound & Marullo 1996). Additionally, 20 species are classified in Phlaeothripidae, with 18 belonging to the large subfamily Phlaeothripinae, which includes species with a wide range of feeding behaviors. The two species recorded in the subfamily Idolothripinae are known to ingest fungal spores (Mound & Marullo 1996).

Phlaeothripidae currently comprises more than 3,700 species worldwide and is the richest group in the order (ThripsWiki 2023). The relatively low number of species in this group in our list likely results from the lack of extensive sampling in specific microhabitats such as leaf litter, dead twigs, and galls, where these insects tend to be more diverse.

The species *Frankliniella bagnalliana* Hood (catalog number NHMUK014233252, NHMUK014233188) and *Liothrips vernoniae* Moulton (catalog number NHMUK014745045, NHMUK014745063, NHMUK014745044, NHMUK014745062, NHMUK014745064, NHMUK014745049, NHMUK014745050) were included in this article, from specimens previously collected in Colombia and deposited at NHM, however, they are not mentioned in the scientific literature. Concerning the identification labels for the *L. vernoniae* slides, curiously, the species is not indicated. Only the genus is mentioned. It is necessary to conduct a subsequent review of these slides to corroborate the information.

Acknowledgements

We thank the Colombian Agricultural Institute (ICA), BIOQUALITY-AGRO, the National University of Colombia Bogota (UNAL), and the farmers and growers for their support in carrying out this study. We thank the reviewers for their comments on the early version of the manuscript and help with the English edition.

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